Experimental Glomerulonephritis in Rats Produced with Soluble Antigen-Antibody Complexes after Pretreatment with Aminonucleoside

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The pathogenesis of human glomerulonephritis has not been thoroughly recognized in spite of many extensive experimental examinations and clinical observations. With regard to experimental glomerulonephritis, MASUGI's nephritis and glomerulonephritis in serum sickness has been believed to play an important role. In serum sickness, glomerulonephritis develops during the immune phase of antigen elimination, and antigen-antibody complexes play an important role in the pathogenesis of glomerulonephritis and vascular lesions. It has been also reported that an aminonucleoside, 6-dimethylamino, 3-amino-d-ribose, increased permeability of the glomerular basement membrane for protein. In the present study pretreated rats with aminonucleoside received smaller amount of soluble antigen-antibody complexes in antigen excess which were produced in vitro between BSA anti-BSA rat serum or human γ-gl. anti-human γ-gl. rabbit serum. Glomerulonephritis developed in a short time after administration of immune complexes.

INTRODUCTION

The pathogenesis of experimental serum sickness has been extensively investigated since Rich and Gregory described the characteristic lesions, glomerulonephritis, arteritis and endocarditis.12345

Germuth5 showed, by careful analysis of the immunologic events as related to the histologic alterations, that the lesions developed during the immune phase of antigen elimination and regressed when free antigen appeared in the circulation. The immune phase of antigen elimination was of short duration and was associated with presence of soluble antigen-antibody complexes in the circulation. The important role of these antigen-antibody complexes in the pathogenesis of a certain hypersensitive state has only recently been recognized. It has been

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shown that they can produce anaphylaxis in guinea pigs and mice, contraction of isolated guinea pig smooth muscle and inflammatory changes in the skin. More direct evidence for the role of soluble antigen-antibody complexes in the pathogenesis of serum sickness was produced by the fact that the characteristic lesions of serum sickness were produced by intravenous injection of a large amount of soluble antigen-antibody complexes in normal mice.

Recently an experimental disease resembling human nephrosis has been produced in rats by the subcutaneous injection of an aminonucleoside.

During the early stage of the disorder by aminonucleoside, no detectable glomerular lesions can be seen under light microscopy.

In the present study, experimental glomerulonephritis was produced passively in rats by the intravenous injection of a smaller amount of soluble antigen-antibody complexes than has been used in previous experiments within 7 days after the previous subcutaneous injection of aminonucleoside.

MATERIAL and METHOD

Animals: Male rats of the Wister strain with initial body weight of 100 to 150 gm. were used. They were given free access to a commercial pellet diet (MF Oriental Yeast Mfg., Tokyo) and tap water. Antigens: Human 𝛾-globulin (H.G.G.) were obtained from Midori Juji Co., Osaka, and crystallized bovine serum albumin (BSA) from Armour Pharmaceutical Co., Kankakkee Illinois, USA. Antiserums: These were produced in white male rabbits weighing 2kg. against H.G.G., and in Wister male rats weighing 100 gm. against BSA. Each rabbit received multiple subcutaneous injections of 50 mg. of H.G.G. incorporated with complete Freund's adjuvant (Iatron Laboratories). Four months after this they were given a booster injection of antigen. The rats were immunized against BSA (10 mg. of antigen) by the same method and were given a booster injection of antigen four weeks after the multiple injections. Both the rabbits and the rats were bled on the 7th day after the booster injection.

The protein level of the samples was measured using a hand protein refractometer (Hitachi). Each antiserum sample was distilled up to one per cent with isotonic saline solution. Soluble complexes: The equivalent point was examined by Deen-Webb's method. Four times excess antigen, each antigen containing 10 mg. of protein in one ml., was added to the antiserum. No precipitation was found. The concentration of antigen and antibody protein was standardized at 10 mg. in 1 ml., so that 1 ml. contained 10 mg. of soluble complexes.

Aminonucleoside (AN): This was obtained from Nutritional Biochemi-
cals Corporation, Cleveland, Ohio. Rats received a daily subcutaneous injection of AN, (0.3 ml. of 0.5 per cent solution per 100 gm. of body weight).

Urinary protein: This was estimated by dipping the end of an Albustix and comparing the color with six color standards representing the following concentration of urinary protein: 0(−), tr, 30(+), 100 (+++), 300(+++) and over 1000 mg. (++++) protein per 100 ml. of urine. Some of the urinary specimens were pooled over night for immunoelectrophoretic study.

Immunoelectrophoretic study: First, zone electrophoresis of the antigens (urine specimen and serum) was carried out for a period of 50 minutes in veronal buffer (pH 8.6, ½=0.06) using a current of 0.8 milli Amp. per cm. width of cellulose acetate filter paper. Strips of filter paper with a standard length of 6 cm. and variable width of 1 or 2 cm. were used. Antibody (anti-rat serum rabbit antibody and anti-BSA rabbit antibody) was placed on the empty half of the acetate filter paper along with the antigen, and then the paper was placed in liquid paraffin for 24 hours. The reaction between antigen and antibody was studied using Panceau 3R in staining.

Table I describes the number of experimental rats and the experimental procedure followed in each of the three main groups and also in the subgroups.

The experimental animals, as a rule, were sacrificed by bleeding from the abdominal aorta, after being anesthetized with ether. Some of these were used for perfusion fixation with isotonic 1 per cent glu-

<table>
<thead>
<tr>
<th>Group</th>
<th>Number of Rats</th>
<th>Number of AN Inj.*</th>
<th>Total Does of Sol. Complexes</th>
<th>Interval between Inj.</th>
<th>Time of Sacrifice after the Last Inj.</th>
<th>Experimental Period</th>
<th>Urinary Protein at Sacrifice</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>3</td>
<td>7</td>
<td>−</td>
<td>−</td>
<td>1 day</td>
<td>8 days</td>
<td>(−)−(++)</td>
</tr>
<tr>
<td>II (1)**</td>
<td>12</td>
<td>0</td>
<td>3 ml.</td>
<td>12 hrs.</td>
<td>3 days</td>
<td>4½ days</td>
<td>(±)−(−)</td>
</tr>
<tr>
<td></td>
<td>(2)***</td>
<td>6</td>
<td>3 ml.</td>
<td>12 hrs.</td>
<td>3 days</td>
<td>4½ days</td>
<td>(±)−(−)</td>
</tr>
<tr>
<td>II (1)** a</td>
<td>10</td>
<td>1−3</td>
<td>3 ml.</td>
<td>12 hrs.</td>
<td>3 days</td>
<td>5½−7½ days</td>
<td>(±)−(−)</td>
</tr>
<tr>
<td>b</td>
<td>7</td>
<td>4−5</td>
<td>3 ml.</td>
<td>12 hrs.</td>
<td>3 days</td>
<td>8½−9½ days</td>
<td>(−)−(−)</td>
</tr>
<tr>
<td>c</td>
<td>11</td>
<td>6−7</td>
<td>3 ml.</td>
<td>12 hrs.</td>
<td>3 days</td>
<td>10½−11½ days</td>
<td>(++++)</td>
</tr>
<tr>
<td>d</td>
<td>2</td>
<td>7</td>
<td>7 ml.</td>
<td>3 days</td>
<td>3 days</td>
<td>31 days</td>
<td>(++)−(++)−(++)</td>
</tr>
<tr>
<td>(2)***a</td>
<td>5</td>
<td>7</td>
<td>3 ml.</td>
<td>12 hrs.</td>
<td>3 days</td>
<td>11½ days</td>
<td>(++++)−(++)−(++)</td>
</tr>
<tr>
<td>b</td>
<td>2</td>
<td>7</td>
<td>3 ml.</td>
<td>12 hrs.</td>
<td>7 days</td>
<td>15½ days</td>
<td>(++)−(++)−(++)</td>
</tr>
</tbody>
</table>

*Rats received daily subcutaneous injections of aminonucleoside (AN) 0.3 ml. of 0.5% solution per 100 gm. of body weight.

**Rats in Group II (1) and Group III (1) received homologous soluble complexes.

***Rats in group II (2) and Group III (2) received heterologous soluble complexes.
Tissue: The kidney and other organs were sectioned after measuring weight and fixed in neutralized formalin and Zenker’s solution. Some of the rats were fixed using a perfusion technique with 1 per cent glutaraldehyde. H.E., PAS, and PAM were used for staining.

RESULTS

Urinary protein: Protein was either absent or present in only trace amounts in the urine of normal rats. The urinary protein excretion of the rats which was treated with AN for 1 to 3 days, indicated (−) to (±), for 4 to 5 days, (±) to (+) and for 6 to 7 days indicated (+) to (++). The rats which received injections of heterologous or homologous soluble complexes after treatment with AN for 6 to 7 days showed a rapid increase in urinary protein on the third day after the last injection of soluble complexes. (Fig. 1) However, the animals which received injections of soluble complexes after treatment with AN for 5 days had the same urinary protein content as the animals treated only with AN. Rats which received soluble complexes alone excreted mostly urine with a low protein content.

Organ weight per 100 gm. of body weight: The weights of the kidneys and hearts of the 9 rats in group II were measured. The mean weight of the kidneys was 0.87 gm. and that of the hearts was 0.45 gm. In group III (1) a, the mean weight of the kidneys was 0.81 gm. and that of the hearts was 0.35 gm.; in group III (1) b, the mean weight of the kidneys was 0.90 gm. and that of the hearts was 0.46 gm.; and in group III (1) c, the mean weight of the kidneys was 1.05 gm. and that of the hearts was 0.50 gm. The mean weights of the kidneys and hearts in group III (1) c were significantly heavier than the mean weights in other groups.
Immunoelectrophoretic result: BSA was noted in all sera of group III (1) a, b and c. The urine specimens of group III (1) c showed a high protein content. In addition, the presence of BSA was demonstrated immunologically, and multiple reaction lines were noted between the anti-rat serum rabbit antibody and urine specimen (Fig 2). However, in the urine of group III (1) a and b indicating low grade proteinuria BSA was not demonstrated, and only one line was noted between anti-rat serum rabbit antibody and urine specimen (Fig 3).

Light microscopic observation: None of the rats in groups I and II showed any pathologic alterations. The accumulation of granular eosinophilic, PSA-positive materials in the cytoplasm of glomerular epithelial cells in AN treated rats was not different from that of normal, untreated rats. (Fig 4). Acute glomerulonephritis developed in all rats of group III (1) c and III (2). The pathologic features of renal changes in these groups were similar and will be described together. All glomeruli were affected. (Fig 7). The glomeruli were hypercellular with swollen endothelial, epithelial and mesangial cells. The capillary lumens were narrow and relatively bloodless. (Fig 8, 9). Kidney sections examined by a perfusion fixation technique revealed swelling and increase of mesangial ground substance and cells. The glomerular basement membrane appeared thick in the section stained with PAS. Granular PAS-positive materials were more numerous in these rats. (Fig 10.) In some glomeruli, adhesion between glomerular tuft and Bowman’s capsule was detected. Lobulation was found frequently. (Fig 9). Protein rich eosinophilic colloid casts were found in the tubular spaces. In gross specimen, these kidneys were white and large with a fine granular surface, and ascites were noted in four rats. Sections from group III (1) d indicated sclerosing of the capillary tuft at the vascular pole. (Fig 11). There were few pathognomonic alterations in the sections from the kidneys of groups III (1) a and b. (Fig 5, 6)

DISCUSSION

An experimental lesion resembling human nephrosis has been produced in rats by subcutaneous injection of an aminonucleoside, 6-dimethylamino, 3-amino-ribose. Increased urinary protein excretion has been noted in the experimental animals by the 5th day after injections of AN and the degree of proteinuria increased progressively, reaching a peak on the 15th day. In this study, rats received daily injections of AN, 0.3 ml. of 0.5 per cent solution per 100 gm. of body weight. Urinary protein was estimated by dipping the end of an Albustix and comparing the color with six color standards. By the 3rd day of AN treatment urinary protein was (±) to (+), by the 7th day (+) to (++) . These results appeared to be similar to the observations made by other reporters. When kidney sections from AN treated rats were examined by light microscopy, no detectable glomerular lesions were identified in the early and reversible
stage of injury (within 7 days).

According to other studies, electron microscopic examination of the glomeruli from rats treated with AN has revealed definite abnormalities of the epithelial cells.\textsuperscript{15,16}

Serial studies of this lesion have indicated that the earliest change was a swelling and coalescence of the foot processes beginning on the 7th day after injection of AN. In addition, large vacuoles containing granular material, similar in appearance to precipitated protein, were observed inside the cytoplasm of epithelial cells of the nephrotic rats. Harkin reported that this granular material was the same as the PAS-positive material found in light microscopic examination. At the present time these materials have been interpreted as morphologic appearance of accelerated transport of serum protein.\textsuperscript{17}

In the present study the rats received intravenous injections of homologous or heterologous soluble antigen-antibody complexes in antigen excess by the time they indicated the early postproteinuric state (i.e., increased permeability of protein from glomerular basement membrane). Only minimal morphological changes were found.

Experimental serum sickness has been extensively investigated since Rich & Gregory, intravenously injected a large amount of horse serum in rabbits.\textsuperscript{12,13,14}

Hawn et al.\textsuperscript{18} and others\textsuperscript{19,20,21,22,23}, employing purified antigens, have shown lesions similar to those Rich & Gregory reported proliferative glomerulonephritis and lesions similar to those seen in polyarteritis nodosa and rheumatic disease.

Germuth et al., interpreting the immunologic events\textsuperscript{5} as related to morphologic alterations, stated that the lesions developed during the immune phase of antigen elimination and that the morphologic lesions of serum sickness were the result of an antigen-antibody reaction. It has been repeatedly demonstrated that this reaction occurred as the host began to form antibody while antigen was still in circulation and in the tissues. In these observations, it has been apparent that the development of serum sickness occurs at the time of antigen-antibody interaction. More direct support for the role of soluble antigen-antibody complexes in the pathogenesis of serum sickness was supplied by the study of McClusky et al.\textsuperscript{11,12}, which showed that the intravenous injections of large amount of soluble antigen-antibody complexes produced the characteristic lesions of serum sickness in normal mice. They explained how these complexes were arrested in the tissue affected by the serum sickness, that antigen-antibody complexes had been in circulation as unstable colloidal particles, that they were readily phagocytized by the cells of the reticuloendothelial system and such colloidal particles could be localized in glomeruli and vascular endothelium in various organs under the influence of agents such as histamine, known to be released by antigen-antibody complexes. For these reasons it was postulated
that in serum sickness antigen-antibody complexes produced changes
in the endothelium of glomeruli and blood vessels by releasing sub-
stances such as histamine. The alteration thus induced leads to the
arrest of the complexes in these sites and, in turn, to the tissue
damage.

McClusky reported that it required at least 12.5 mg. antibody
protein per 100 gm. of body weight to produce the lesions of serum
sickness.

Ishizaka et al., Germuth et al. and Treanin et al. reported
that soluble antigen-antibody complexes in antigen excess have biological
activity which does not have antigen or antibody alone, but also inclu-
des heavy (-Ag3 Ab2)24 and light (Ag2 Ab) complexes, the former
being more active biologically.25

Antigen excess immune complexes prepared from heterologous or
homologous antibody have produced (1) anaphylactic shock when injected
into normal unsensitized guinea pigs67 and mice,8 (2) contraction of
isolated guinea pig smooth muscles,9 and (3) passive skin reactions in
guinea pigs.10 Large, multiple doses cause transient acute proliferative
glomerulonephritis in mice.1112

McClusky et al.26 produced experimental glomerulonephritis in
mice employing two kinds of soluble complexes, crystallized ovalbumin
anti-ovalbumin rabbit antiserum system and crystallized bovine serum
albumin anti-bovine serum albumin rabbit antiserum system. They
stated that the former was more effective for development of the dis-
ease than the latter. They concluded that the difference in excretion
of these two antigen might be a contributory factor in the greater
severity of the disease observed with the ovalbumin anti-ovalbumin
rabbit serum system.

Coons et al. intravenously injected mice with 3 kinds of protein
(ovalbumin, bovine serum albumin, and human γ-globulin) and obser-
vied that the disappearance of ovalbumin from the kidney was fast and
that of human γ-globulin was slow.27 Their differing rates of disappa-
rance are partly due to the relative ease with which they pass through
the glomerular filter.28

In consideration of these events and the biological activity of imm-
une soluble complexes, it appears that soluble complexes of low mole-
cular systems are more complex at the site of glomerular filter, and
thus, biologically active complexes such as -Ag3Ab2 become numerous
in a short time.

On the other hand, it has been considered that the affiniating or
combining ability of injected protein with tissues is an important factor
contributing to some hypersensitive states.

Miller produced experimental glomerulonephritis in mice by intra-
venous injection of heterologous (ovalbumin anti-ovalbumin rabbit serum
system) and homologous (ovalbumin anti-ovalbumin mouse serum system)
immune soluble complexes. He reported that the lesions of the mice which received homologous complexes were more severe.

In the present study, rats received homologous or heterologous soluble complexes intravenously by the time they indicated urine with low protein content by aminonucleoside treatment and they were examined histologically and immunologically.

The dose of the soluble complexes injected in this study was standardized at 3 ml. of 10 mg. protein in 1 ml., a total 30 mg. of protein. This ranged from 1 mg. to 2mg. of antibody protein in terms of antibody protein per 100gm. of rat body weight. It is not necessary to inject such large amount to produce glomerulonephritis, if the rats had received aminonucleoside in advance. The glomerular basement membrane has been considered the main filter for filtration of serum protein in the normal kidney. Experimental diseases, such as aminonucleoside nephrosis, increased ferritin uptake in glomerular basement membrane and glomerular epithelial cells.

Protein is filtered more rapidly into urine, when the permeability of the glomerular basement membrane is increased by the aminonucleoside treatment. In addition to the effect of aminonucleoside, immune soluble complexes may possibly cause an increase in the permeability.

The results of the immunoelectrophoetic examination in this study are indirect support for this hypothesis. In urine with a high protein content, the presence of BSA, a component of soluble complexes, was detected immunologically. In addition, multiple reaction lines were found between urine specimens and anti-rat serum rabbit antiserum, but only a few such lines were found between anti-rat antibody and urine specimens with a low protein content.

Thus, it is possible that under these circumstances biologically active soluble complexes (-Ag3Ab2) are accumulated in great quantities, in a short time, at the site of glomerular filter membrane.

This hypothesis leads to the consideration that these biologically active complexes play an important role in the pathogenesis of experimental glomerulonephritis.

The characteristic findings of acute proliferative glomerulonephritis (hypercellularity, ischemia and swelling of glomeruli), appear to be caused by irritation by biologically active immune complexes, whereas these changes have not been seen in aminonucleoside nephrosis.

It could not be determined by the present study whether soluble complexes themselves or complexes and chemical agents together played a major part in the histogenetic effect. (Chemical agents, such as histamine, are believed to be released by immune complexes)

The role of affinity or combination ability of protein to tissue also needs to be clarified by further study.

In the present experimental disease, I could not find other lesions
such as arteritis or endocarditis which were reported by McClusky et al. and others. I suspect that smaller amounts of soluble complexes used for the injections in my experiment may cause this difference.

Comparing the present experimental glomerulonephritis with human poststreptococcal glomerulonephritis, the mechanism in the development of human glomerulonephritis would be as follows: In the first phase, a toxin, such as streptococcal toxin, causes an increase in the permeability of glomerular basement membrane. Indirect support for this suggestion is fibrile proteinuria clinically observed. In the second phase, biologically active soluble antigen-antibody complexes, such as $\text{Ag}_3\text{Ab}_2$, accumulate in great quantities in a short time at the site of glomerular basement membrane which was damaged in the first phase, and then glomerulonephritic lesions develop with hypercellularity, ischemia and swelling of glomeruli.

This hypothesis appears to be supported by the present experimental findings: Aminonucleoside treatment coincides with the first phase, and injection of soluble complexes coincides with the second phase.

Diffuse glomerulonephritis can be divided into two main groups, the nephritic type and the nephrotic type. The nephritic type mainly affects children and young adults and it is commonly a sequela of acute tonsillar or pharyngeal infections with group A streptococci associated with acute onset. Hematuria, pyuria and hypertension are often associated with acute cases of nephritic glomerulonephritis. Complete recovery is common. The glomerular lesions in the acute stage are mainly proliferative, exudative or hemorrhagic and occasionally necrotizing.

The nephrotic type, on the other hand, affects a wider age group and a history of a preceding infection is rarely elicited. The onset is insidious; nephrotic symptoms (protracted proteinuria, edema, ascites, hypoproteinemia and hyperlipemia) are prominent; histologically thickening of basement membrane and lobulation are commonly seen. However, as Allen and others emphasized, there are frequent transitions between these two types. Whether the pathogenesis of these two types of glomerulonephritis is the same or not, is an interesting question.

The present experimental glomerulonephritis is a mixed type: Hypertension (which is suspected because of the increased heart weight), and histologic findings of the kidney (hypercellularity, ischemia, and swelling of glomeruli), indicate the nephritic type of glomerulonephritis, but severe proteinuria, ascites (observed in some rats), lobulation, and glomerular basement membrane thickening indicate the nephrotic type of glomerulonephritis.

It has been considered that the autoimmune mechanism plays an important role in chronic evolution of glomerulonephritis. In this study chronic evolution of the disease was not discussed.
Two rats which received an injection of soluble complexes every 3 days for a total of 7 injections over a period of 24 days, indicated interesting lesions leading to chronic glomerulonephritis. The present study, however, does not settle the question what role the autoimmune mechanism or repeated irritation of exogenous soluble complexes or both played as a cause of the lesions.

**SUMMARY**

Experimental glomerulonephritis was produced in rats by administration of aminonucleoside for 6 to 7 days followed by 3 intravenous injections of homologous or heterologous soluble complexes in antigen excess with a 12-hour interval between each injection.

This experimental model supplies several forms of evidence leading to a hypothesis on the pathogenesis of human glomerulonephritis.

1. Smaller amounts of soluble complexes than reported by others were sufficient to cause glomerulonephritis in rats if previously treated with aminonucleoside.
2. Histologically, this type of glomerulonephritis is a mixed type.
3. Severe proteinuria was noted in this glomerulonephritis, and BSA, a component of soluble complexes, was demonstrated immunologically in the urine with a high protein content, whereas it was not present or noted in only trace amounts in the urine with a low protein content.
4. This appears to be indirect support for the hypothesis that biologically active complexes accumulate in great quantities in a short time at the site of glomerular filter membrane.
5. These complexes appear to be an important factor in the development of characteristic glomerulonephritic changes, hypercellularity, ischemia and swelling of glomeruli.
6. The increased permeability of glomerular basement membrane represents the first phase of the development of glomerulonephritis in man and irritation of biologically active complexes at the site of the glomerular filter membrane represent the second phase. The first phase coincides with AN treatment, and the second phase coincides with c injection of soluble complexes as demonstrated in this study.

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**REFERENCES:**

(1) RICH, A.R. and GREGORY, J.E.: The experimental demonstration that periar-


(20) MELLORS, R.C., ARIAS- STELLA, J., SIEGCL, M. and PRESMAN, D.: Analytical pathology. II. Histopathologic demontration of glomerular-localizing antibodies


(28) PAPPENHEIMER, J.R.: Renal function (3 conference). Josiah Macy Found, (1951)


Fig. 2: Photograph (left) and diagram (right) of immunoelectrophoresis of urine specimen indicating severe proteinuria. They show four reaction lines between anti-rat serum rabbit antibody and one line between anti-BSA rabbit antibody.

Fig. 3: Photograph (left) and diagram (right) of immunoelectrophoresis of urine specimen indicating low grade proteinuria. They show only one line between the urine specimen and rabbit serum containing anti-rat antibody.

Fig. 4: Kidney of a rat which was treated with AN for 7 days. Glomeruli are almost normal. PAS stain, low magnification.

Fig. 5: Glomerulus of a rat which received homologous soluble complexes every 12 hours for 3 injections. No detectable change is seen. Perfusion fixation with glutaldehyde, PAS stain, high magnification.
Fig. 6: Glomerulus from a rat which received homologous complexes after AN treatment for 2 days. It shows almost normal structure. Perfusion fixation, PAS stain, high magnification.

Fig. 7: Kidney from a rat which received heterologous complexes after AN treatment for 7 days. Swelling of glomeruli, hypercellularity and ischemia appear. These lesions are noted in all glomeruli. PAS stain, low magnification.

Fig. 8: Glomerulus from a rat which received homologous complexes after AN treatment for 7 days. It shows hypercellular, relatively bloodless glomerulus. PAS stain, high magnification.

Fig. 9: Photomicrography from same section illustrated Fig. 7. Hypercellularity, relatively bloodless, in addition, lobulation is observed in this glomerulus. PAS stain, high magnification.
Fig. 10: Glomerulus from a rat which received homologous complexes after AN treatment for 6 days. Adhesion between loops, thickening of basement membrane is seen. Perfusion fixation, PAS stain, high magnification.

Fig. 11: Glomerulus from a rat which received homologous complexes every 3 days for 7 injections after AN treatment for 7 days. PAS positive, sclerosing lesion is observed. PAS stain, high magnification.