### Title
In vitro activity of garenoxacin against Streptococcus pneumoniae mutants with characterized resistance mechanisms.

### Author(s)
Yamamoto, Kazuko; Yanagihara, Katsunori; Sugahara, Kazuyuki; Imamura, Yoshifumi; Seki, Masafumi; Izumikawa, Koichi; Kakeya, Hiroshi; Yamamoto, Yoshihiro; Hirakata, Yoichi; Kamihira, Shimeru; Kohno, Shigeru

### Citation
Antimicrobial agents and chemotherapy, 53(8), pp.3572-3575; 2009

### Issue Date
2009-08

### URL
http://hdl.handle.net/10069/22297

### Copyright
© 2009, American Society for Microbiology. All Rights Reserved.
Title-

In vitro Activity of Garenoxacin against Streptococcus pneumoniae Mutants with Characterized Resistance Mechanisms

Running title-

Activity of Garenoxacin against S. pneumoniae mutants

Authors-

Kazuko Yamamoto¹, Katsunori Yanagihara*², Kazuyuki Sugahara², Yoshifumi Imamura¹, Masafumi Seki¹, Koichi Izumikawa¹, Hiroshi Kakeya¹, Yoshihiro Yamamoto¹, Yoichi Hirakata³, Shimeru Kamihira², and Shigeru Kohno¹

Institutions-

¹Department of Infection and Immunology, ²Department of Laboratory Medicine, Nagasaki University Graduate School of Biomedical Sciences, 1-7-1 Sakamoto, Nagasaki City 852-8501, Japan

³Department of Infection Control and Laboratory Diagnostics, Internal Medicine, Tohoku University Graduate School of Medicine, 1-1 Seiryo-machi, Aoba-ku, Sendai 980-8574 Japan
Corresponding Author-

Katsunori Yanagihara, M.D., Ph.D.

Department of Laboratory Medicine,

Nagasaki University Graduate School of Biomedical Sciences,

1-7-1 Sakamoto, Nagasaki City 852-8501, Japan

Phone: +81-95-849-7418

FAX: +81-95-849-7257

e-mail: kyana-ngs@umin.ac.jp
ABSTRACT

We evaluated garenoxacin potency in selection of *Streptococcus pneumoniae* resistance compared to other quinolones, by determining the mutant prevention concentration, using strains with and without topoisomerase genes mutations. Garenoxacin is potently active against pneumococci including topoisomerase mutations-containing strains, at a level significantly greater than other quinolones. The genetic analysis of *S. pneumoniae* mutants created by garenoxacin exposure revealed that *gyrA* gene is a primary target of garenoxacin.
Background

The emergence of *Streptococcus pneumoniae* strains with resistance to the β-lactams and macrolides has complicated the treatment of pneumococcal respiratory tract infections and created a need for new agents. Recently developed compounds within the quinolone group have demonstrated enhanced potency against *S. pneumoniae*. In particular, agents such as moxifloxacin, gatifloxacin, and levofloxacin have been recommended and used for therapy (16). However, *S. pneumoniae* strains exhibiting quinolone resistance have been observed in several countries (3,9,11,15,18). Furthermore, evidence suggests that increased usage of these compounds could lead to further resistance development and treatment failure (5,7,19).

Quinolone resistance in *S. pneumoniae* is mediated by amino acid substitutions within the quinolone resistance determining regions (QRDRs) of DNA gyrase (GyrA or GyrB) and/or topoisomerase IV (ParC or ParE), sometimes in combination with efflux (1,12,14).

The mutant prevention concentration (MPC), the drug concentration that prohibits the growth of mutants from a susceptible population of more than $10^{10}$ cells, is a novel approach in the evaluation of FQ potency (2). Additionally, mutant selection window (MSW), which is defined as the range between MIC and MPC, provides a means of defining the ability of antibiotics in preventing the emergence of mutants (21).

Garenoxacin (GAR) is a novel des-F(6) quinolone with a broad spectrum of activity against respiratory tract pathogens, including *S. pneumoniae* with elevated resistant-level
fluoroquinolone MIC values (4,10,13,20). The aim of this study was to evaluate GAR potency in selection of *S. pneumoniae* resistance compared to other quinolones, by determining the MPC and the MSW, using strains with and without mutations of QRDR in topoisomerase genes. Additionally, we examined the intrinsic development of *S. pneumoniae* resistant mutants created by garenoxacin exposure, with detailed evaluation of additional QRDR mutations and efflux, for determining the target of GAR.

**Materials and Methods**

A total of eight *S. pneumoniae* clinical isolates were used in this study. QRDR genetic backgrounds and quinolones MICs of isolates are summarized in Table 1. All strains were wild-type with GyrB and with ParE. Isolates W001 and W002 are quinolone-susceptible isolates with wild-type ParC and GyrA. An isolate S001 have a Asp83Asn mutation in ParC with wild-type GyrA. An isolate S002 have a Ser79Phe mutation in ParC with wild-type GyrA. An isolate S002 have a Ser79Phe mutation in ParC with wild-type GyrA. Isolates S003 and S004 have a GyrA single mutation (Ser81Phe in S003 and Gly85Asn in S004). Isolates D001 and D002 have both GyrA and ParC mutations (D001; Ser79Phe in ParC and Gly85Lys in GyrA, D002; Ser79Phe in ParC and Ser81Phe in GyrA). All strains were exposed to two, four, eight, sixteen, thirty-two, and sixty-four times the MICs of ciprofloxacin (CIP), levofloxacin (LVX), gatifloxacin (GAT), moxifloxacin (MXF), and GAR for 48 hr to 72 hr of incubation at 37°C in 5%CO₂. The MPCs were measured with the
procedure previously described (2). Briefly, 200μl of a culture containing 10 log_{10} CFU/ml was applied to Mueller Hinton II Agar plates containing 5% sheep blood and drugs at various concentrations. MPCs were recorded as the lowest antibiotic concentration that prevented bacterial colony formation at 48 hr. All determination was done in duplicate, and the results were identical.

Genomic DNA was extracted from growing mutants (maximum eight mutants per one plate were purely cultured, see Table 2) using QIAGEN® Blood Mini Kit (QIAGEN, Hilden, Germany). All DNA extracts of mutants were screened for possessing QRDR mutations of \textit{parC} and \textit{gyrA} genes using a novel PCR-melting curve analysis (PCR-MCA) method which we previously reported (8). Briefly, probes labeled with LC-Red 640 and fluorescein were used with designated primers (8), which targeting four QRDR positions (Ser79 and Asp83 of the \textit{parC} gene and also Ser81 and Gly85 of the \textit{gyrA} gene). PCR was performed in a total volume of 20 μl containing 5 μl of DNA template, 4 μl of LightCycler® 480 Genotyping Master (Roche Diagnostics, Basel, Switzerland), 3mM MgCl₂, 0.2 μM each probes, and 0.5 μM each primers. Thermal cycling was performed with an initial hold for 10 min at 95°C, followed by 35 cycles of 5 sec at 95°C, 10 sec at 55°C and 12 sec at 72°C. Melting curve was generated by cooling to 40°C for 30 sec, followed by heating to 80°C at a rate of 2.0°C/s. The PCR-MCA assay was performed using LightCycler480® analysis software (Roche Diagnostics, Basel, Switzerland). The total assay time was approximately 1 hr. Nucleotide
mismatches between the sequence and the hybridization probe result in a lower mutant melting temperature ($T_m$) than that of the wild-type. The assay makes it possible to quickly and easily differentiate a mutant strain from a wild-type strain.

The QRDR sequence of topoisomerase genes ($parC$, $parE$, $gyrA$, and $gyrB$) of mutant strains were confirmed by all directly sequenced at the nucleotide level using the BigDye® Terminator ver.3.1 Sequencing Standard Kit and an ABI PRISM™ 310 Genetic Analyzer (both by Applied Biosystems, CA, USA) with published primers (17). The QRDR DNA sequencing results were compared with the strain R6 (GenBank accession no. NC_003098).

MIC determination was done in parallel both with or without an efflux inhibitor (10µg of reserpine/ml). Efflux was considered as present when a $\geq 2$-fold reduction of the MIC was observed.

Results

The MPCs and MSW ranges of $S. pneumoniae$ isolates were summarized in Figure 1. The MPCs which shown were averaged between W001 and W002, S001 and S002, S003 and S004, D001 and D002. The MPC order in $S. pneumoniae$ strains was CIP > LVX > GAT $\geq$ MXF > GAR. GAR potency was 16- to 64-fold greater than LVX or CIP, and 4-fold superior to GAT or MXF. GAR had a significantly narrow MSW compared to other quinolones, even against strains with QRDRR mutations. Compared between strains, MSW range was, wild-type
strains (W001 and W002) < strains with QRDR single mutation (S001-004) < strains with
double QRDR mutations (D001 and D002). Strains with parC single mutation showed narrow
MSW compared to strains with gyrA single mutation which exposed to LVX, GAT and GAR,
while CIP and MXF were in reverse. Number of pneumococcal picked up mutants which
created by GAR exposure were shown in Table 2. QRDR wild-type strains created mutants
with GAR exposure from MIC to 8×MICs, while strains with QRDR mutations created
mutants from MIC to 16-32×MICs. QRDR genetic changes in S. pneumoniae mutants created
by GAR exposure were shown in Table 3. Results of PCR-MCA assay of mutants from
wild-type strains (W001 and W002) created by GAR exposure revealed high percentage of
additional gyrA mutations. With regard to the strains with parC single mutation (S001 and
S002), high percentages of additional gyrA mutation in codon 81 was seen. On the other
hand, strains with gyrA single mutation (S003 and S004), additional parC mutation in codon
79 was seen in high percentage. Strains with double QRDR mutations (D001 and D002)
showed additional gyrA mutaions resulted from GAR exposure. None of the mutants created
from GAR exposure showed existence of efflux, by MIC determination with reserpine. DNA
sequence analysis using mutants without additional QRDR mutations with GAR exposure,
showed two mutants from W001, and one mutant from W002 with additional parE mutation
(Asp435Asn). In addition, two mutants from W001, one mutant from W002, two mutants
from S003, and one mutant from S004 showed additional parC mutation (Lys137Asn). With
regard to gyrB gene, only one mutant from S001 showed Asp435Asn mutation (data not shown).

Discussion

In the present study, the MIC results showed that GAR was potently active against pneumococci including QRDR mutations-containing strains, at a level significantly greater than other quinolones. In addition, significantly narrow range of MSW, and the low MPC values demonstrated that pneumococcus hard to acquire resistance to GAR, compared to other quinolones.

We used an original PCR-MCA assay to enable a detailed genetic analysis of QRDR mutations using vast numbers of mutants. This was difficult in previous studies due to the time-consuming nature of analyzing the DNA sequence of mutants.

A high proportion of the mutants that were from isolates with a parC single mutation acquired secondary mutations in gyrA and became highly resistance to all of the quinolones we used in our study. With regard to wild type pneumococcal strains, the high percentage of additional GyrA mutations was seen in GAR-exposed mutants. This may indicate that GAR may have a more balanced affinity for the two target enzymes, with a slight initial preference for GyrA as an initial target.

A total of 72 out of 231 mutants (31%) were not detected of additional gyrA or parC
mutations in this study. These remaining mutants were expected to acquire additional other
QRDR mutations (parE or gyrB) or efflux, however, only ten mutants (14%) were detected by
the sequence of topoisomerase genes. These mutants may have acquired other resistant
mechanisms, such as plasmid-based resistance. Further studies are needed to clarify
pneumococcal resistant mechanisms to GAR.

Although quinolone resistance in S. pneumoniae isolates remains low, the opportunity for the
treatment of respiratory tract infections rises. The potential for resistance formation should
thus be considered when specific fluoroquinolones are selected for treatment. Including MPCs
as part of a dosing strategy may be one means of limiting the selection of quinolone-resistant
mutants and preserving this class of antibiotic.

In conclusion, a novel des-F(6) quinolone, Garenoxacin showed the low MPC value and the
narrow MSW for QRDR mutation containing pneumococcal strains, suggested that will be
useful in minimizing the selection of quinolone-resistant mutants.
References


<table>
<thead>
<tr>
<th>S. pneumoniae strains</th>
<th>QRDR genetic background</th>
<th>MIC (µg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>par*C</td>
<td>gyr*A</td>
</tr>
<tr>
<td>W001</td>
<td>wt*a</td>
<td>wt</td>
</tr>
<tr>
<td>W002</td>
<td>wt</td>
<td>wt</td>
</tr>
<tr>
<td>S001</td>
<td>Asp83Asn</td>
<td>wt</td>
</tr>
<tr>
<td>S002</td>
<td>Ser79Phe</td>
<td>wt</td>
</tr>
<tr>
<td>S003</td>
<td>wt</td>
<td>Ser81Phe</td>
</tr>
<tr>
<td>S004</td>
<td>wt</td>
<td>Gly85Asn</td>
</tr>
<tr>
<td>D001</td>
<td>Ser79Phe</td>
<td>Gly85Lys</td>
</tr>
<tr>
<td>D002</td>
<td>Ser79Phe</td>
<td>Ser81Phe</td>
</tr>
</tbody>
</table>

Abbreviation: *a wt, wild type. CIP, ciprofloxacin; LVX, levofloxacin; GAT, gatifloxacin; MXF, moxifloxacin; GAR, garenoxacin.
<table>
<thead>
<tr>
<th>( S.\ pneumoniae ) strains</th>
<th>GAR MIC (( \mu \text{g/ml} ))</th>
<th>No. of picked up mutants (n=231)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>GAR concentrations (( \times \text{MICs} ))</td>
<td>2( \times )</td>
</tr>
<tr>
<td>W001</td>
<td>0.015</td>
<td>8</td>
</tr>
<tr>
<td>W002</td>
<td>0.015</td>
<td>8</td>
</tr>
<tr>
<td>S001</td>
<td>0.03</td>
<td>8</td>
</tr>
<tr>
<td>S002</td>
<td>0.06</td>
<td>8</td>
</tr>
<tr>
<td>S003</td>
<td>0.06</td>
<td>8</td>
</tr>
<tr>
<td>S004</td>
<td>0.06</td>
<td>8</td>
</tr>
<tr>
<td>D001</td>
<td>0.25</td>
<td>8</td>
</tr>
<tr>
<td>D002</td>
<td>0.5</td>
<td>8</td>
</tr>
</tbody>
</table>

Abbreviation: GAR, garenoxacin.
Table 3.

<table>
<thead>
<tr>
<th>S. pneumoniae strains</th>
<th>QRDR genetic background</th>
<th>ParC79 (n=31)</th>
<th>ParC83 (n=31)</th>
<th>GyrA81 (n=31)</th>
<th>GyrA85 (n=31)</th>
</tr>
</thead>
<tbody>
<tr>
<td>W001</td>
<td>wt^a, wt</td>
<td>0 / 24 (0%)</td>
<td>0 / 24 (0%)</td>
<td>12 / 24 (50%)</td>
<td>3 / 24 (12%)</td>
</tr>
<tr>
<td>W002</td>
<td>wt, wt</td>
<td>0 / 24 (0%)</td>
<td>0 / 24 (0%)</td>
<td>16 / 24 (67%)</td>
<td>0 / 24 (0%)</td>
</tr>
<tr>
<td>S001</td>
<td>Asp83Asn, wt</td>
<td>0 / 31 (0%)</td>
<td>16 / 33 (48%)</td>
<td>3 / 33 (9%)</td>
<td>0 / 31 (0%)</td>
</tr>
<tr>
<td>S002</td>
<td>Ser79Phe, wt</td>
<td>0 / 33 (0%)</td>
<td>16 / 33 (48%)</td>
<td>3 / 33 (9%)</td>
<td>0 / 33 (0%)</td>
</tr>
<tr>
<td>S003</td>
<td>wt, Ser81Phe</td>
<td>21 / 32 (65%)</td>
<td>0 / 32 (0%)</td>
<td>7 / 31 (23%)</td>
<td>22 / 31 (70%)</td>
</tr>
<tr>
<td>S004</td>
<td>wt, Ser81Phe</td>
<td>18 / 32 (56%)</td>
<td>0 / 32 (0%)</td>
<td>5 / 24 (21%)</td>
<td>12 / 24 (50%)</td>
</tr>
</tbody>
</table>

* no. of mutants with QRDR mutations / no. of isolated mutants (%)

** presented mutations with original isolates

Abbreviations: ^a wt, wild type.
Figure 1. Levels of Mutant Prevention Concentrations and Mutant Selection Windows of *S. pneumoniae* Strains

<table>
<thead>
<tr>
<th>FQs</th>
<th>0</th>
<th>50</th>
<th>100</th>
<th>150</th>
<th>200</th>
<th>250</th>
</tr>
</thead>
<tbody>
<tr>
<td>GAR</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MXF</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GAT</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>LVX</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CIP</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

FQ concentrations (µg/ml)

- wild type
- `parC` mutation
- `gyrA` mutation
- `parC` and `gyrA` mutation