Malaria Vectors in Lake Victoria and Adjacent Habitats in Western Kenya

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Abstract

The prevalence of malaria among the residents of the Lake Victoria basin remains high. The environment associated with the lake may maintain a high number of malaria vectors. Lake habitats including water hyacinths have been suspected to be the source of vectors. This study investigated whether malaria vectors breed in the lake habitats and adjacent backwater pools. Anopheline larvae were collected within the littoral zone of the lake and adjacent pools located along approximately 24.3 km of the lakeshore in western Kenya, and their breeding sites characterized. Three primary vector species, Anopheles arabiensis, Anopheles gambiae s.s. and Anopheles funestus s.s., and three potential vectors, were found in the lake habitats. Unexpectedly, An. arabiensis was the most dominant vector species in the lake sampling sites. Its habitats were uncovered or covered with short grass. A potential secondary malaria vector, Anopheles rivulorum, dominated the water hyacinths in the lake. Most breeding sites in the lake were limited to areas that were surrounded by tall emergent plants, including trees, and those not exposed to waves. Nearly half of adjacent habitats were lagoons that were separated from the lake by sand bars. Lagoons contained a variety of microhabitats. Anopheles arabiensis dominated open habitats, whereas An. funestus s.s. was found mainly in vegetated habitats in lagoons. The current study confirmed that several breeding sites are associated with Lake Victoria. Given that Lake Victoria is the second largest lake in the world, the related habitats must be extensive; therefore, making targeted vector control difficult. Further exploration is necessary to estimate the effects of lake associated habitats on malaria transmission so as to inform a rational decision-making process for vector control.

Introduction

While malaria transmission has been reduced considerably in some Kenyan regions, it still remains high among the residents in the Lake Victoria basin [1,2]. Intense malaria transmission is often observed in villages near lakes and large reservoirs in Africa [3,4]. Studies have shown that malaria cases increase with decreasing distance to the shores of large water bodies [5,6]. Construction of dams may shift seasonal transmission to perennial transmission [7]. Thus, it is reasonable to hypothesize that the environment associated with the shoreline of Lake Victoria supports a high density of malaria vectors given the year-round availability of water.

The major African malaria vectors, Anopheles arabiensis and Anopheles gambiae s.s., often inhabit small and sunlit temporary water pools [8–10]. Numerous small water pools in the wetlands along the lakeshore may become suitable vector habitats as has been demonstrated from the presence of malaria vectors in stagnant water pools on the lakeshore where tall vegetation has been cleared [11]. Although recent studies suggest that breeding occurs in permanent and semi-permanent water pools [12,13], the characteristics of their habitats suggest that they do not breed within the lake water itself [14].

Another important malaria vector, Anopheles funestus s.s., usually inhabits large, stable water pools covered with aquatic vegetation that can be found in wetlands adjacent to Lake Victoria [8,9,15]. These habitats may exist throughout the year because of a constant supply of seepage water from the lake. Malaria vectors also inhabit large back-water pools (lagoons) along the shore of Lake Victoria [16]. Lagoons appear on strips of land that emerge following reductions in the lake water levels. Waves transport sands, which build up along the shore. The body of water that becomes enclosed behind the mounds of sand creates a lagoon. If a lagoon is large and stable, emergent and floating plants colonise the area. Previously, three major vector species have been recorded in such sites, although the micro-habitats of these vectors within lagoons have not been documented in detail [16].

Given that An. funestus s.s. is closely associated with aquatic vegetation [8,9,15], it has been proposed that the infestation of water hyacinths (Eichhornia crassipes) in African lakes has increased breeding site availability for this vector species [17,18]. A potential secondary vector, Anopheles rivulorum, may also occur in water hyacinths, because the larvae of this species are often associated with floating plants such as Nile cabbage (Pistia stratiotes) [19]. The water hyacinths originated from South America, and were first reported in Lake Victoria in 1989. Since then, they have quickly spread throughout the lake, covering 80% of the Ugandan coastline by 1995 [20]. One study found no malaria vectors amongst water hyacinth supported sites in Lake Victoria [17],
whilst another reported larvae of the *An. funestus* complex within a water hyacinth mat at the margin of the lake [18].

Lake Victoria is the second largest lake in the world, and the effects of the lake habitats on local malaria transmission could be substantial if they support vector breeding. This threat would not be limited to Lake Victoria, as water hyacinths have already invaded several lakes and reservoirs in malaria endemic areas of Africa [21]. Here, we investigated whether the habitats associated with Lake Victoria serve as active breeding sites for malaria vectors.

Materials and Methods

Study area

The study area included the entire shoreline of Lake Victoria in the Gembe West region of Mbita District, western Kenya (Figure 1). The area forms a peninsula surrounded by lake water to the north, west, and east. The eastern shore is within the Nyanza Gulf, and the western shore faces the main lake body. The total shoreline of the study area was approximately 24.3 km, with the eastern and western shores comprising 16.2 km and 8.1 km, respectively. The western shore faces the main lake body and is exposed to strong waves generated by westerly winds. Strong winds from the main lake body are common in this area in the afternoon. Sands carried by waves accumulate on the western shore, and several lagoons have formed behind the sand mounds. While the wind creates strong waves on the western side, the shadow effect of the peninsula creates a calm area along the eastern shore. Most residents depend on fishing and traditional small-scale farming. Two wet seasons occur annually from March to June and October to November, but the periods vary each year. A high prevalence of malaria is known among the local residents (≥40% *Plasmodium falciparum* parasite rate in the overall population) [2]. Three species of malaria vectors were previously identified: *An. arabiensis*, *An. gambiae s.s.*, and *An. funestus s.s.* [18,22].

Water hyacinth habitats in the lake

The distribution of water hyacinths was investigated along the lakeshore using a small engine boat in March 2008. The water hyacinth mats that were drifting across the lake were not surveyed.

When a water hyacinth mat was observed along the shore, its location was recorded using a hand-held global positioning system (GPS; GPSmap 60CSx, Garmin International, Inc., Olathe, Kansas, USA). The scattered water hyacinths were not considered to constitute a mat; only the water hyacinths that were densely assembled into a mat were assessed. When tall emergent vegetation blocked the view from the boat, the presence of water hyacinths was confirmed from the land. The lengths of short water hyacinth mats were estimated using a laser rangefinder (Nikon Laser 550AS; Nikon Vision Co. Ltd., Tokyo, Japan), while those of longer mats were estimated using a geographical information system (GIS; Arc View 9.2; Environmental Systems Research Institute, Redlands, California, USA). Both ends of each mat were located using the GPS and their locations were plotted on the map produced by a satellite image that was taken in June 2006 (0.6-m ground resolution; Quick Bird image; Digital Globe, Longmont, Colorado, USA).

Larval sampling

Each water hyacinth mat in the lake was examined for anopheline larvae using a standard mosquito dipper (350 ml; BioQuip Products, Rancho Dominguez, California, USA) in June 2008 (during the wet season). When a mat was longer than 100 m, it was divided into 100-m sections using the GIS. Each site was dipped a total of 100 times, and a density was estimated as a total number of larvae found in 100 dips per site. The same sampling procedure was repeated for the same sites in February 2009 (during the dry season), and also applied to the other habitats described below. The sampling sites were characterized based on the presence of surrounding trees or non-woody tall emergent plants as these conditions may influence the occurrence of malaria vectors. Water hyacinths were often found trapped by rows of trees (*Aeschynomene elaphroxylon*) that were planted in the lake water to discourage hippopotami (*Hippopotamus amphibius*) from approaching farms near the lake [17,18].

Non-water hyacinth habitats in the lake

During the dry season, mosquitoes were also sampled from various types of non-water hyacinth habitats in the lake. These habitats included:

- **Fishponds:** These were filled with water and often located near the shoreline. They were characterized by a high density of tall emergent vegetation (≥1 m) and the presence of water hyacinths. Three fishponds were selected for larval sampling.

- **Lagoons:** These were small bodies of water that were often enclosed by vegetation. Three lagoons were selected for larval sampling.

- **Lakeside rocks:** These were boulders of different sizes that were not washed away by waves. Two lakeside rocks were selected for larval sampling.

- **Mud flats:** These were shallow areas where water hyacinths were often found. Two mud flats were selected for larval sampling.

- **Water hyacinth bed:** These were areas where water hyacinths were densely assembled into mats. The density of larvae was estimated using a standard mosquito dipper.

- **Geographical features:** These were small geographical features such as inlets, bays, and peninsulas that were often surrounded by water hyacinths. Two geographical features were selected for larval sampling.

- **Village water source:** These were wells, brooks, and streams located near villages. Two village water sources were selected for larval sampling.

- **Water hyacinth island:** These were areas where water hyacinths were densely assembled into mats on small islands. Two water hyacinth islands were selected for larval sampling.

- **Water hyacinth roof:** These were areas where water hyacinths were densely assembled into mats on roofs. Two water hyacinth roofs were selected for larval sampling.

- **Water hyacinthaffle:** These were areas where water hyacinths were densely assembled into mats onaffle. Two water hyacinthaffles were selected for larval sampling.

- **Water hyacinth float:** These were areas where water hyacinths were densely assembled into mats on floats. Two water hyacinth floats were selected for larval sampling.

- **Water hyacinth bed:** These were areas where water hyacinths were densely assembled into mats. The density of larvae was estimated using a standard mosquito dipper.

- **Geographical features:** These were small geographical features such as inlets, bays, and peninsulas that were often surrounded by water hyacinths. Two geographical features were selected for larval sampling.

- **Village water source:** These were wells, brooks, and streams located near villages. Two village water sources were selected for larval sampling.

- **Water hyacinth island:** These were areas where water hyacinths were densely assembled into mats on small islands. Two water hyacinth islands were selected for larval sampling.

- **Water hyacinth roof:** These were areas where water hyacinths were densely assembled into mats on roofs. Two water hyacinth roofs were selected for larval sampling.

- **Water hyacinthaffle:** These were areas where water hyacinths were densely assembled into mats onaffle. Two water hyacinthaffles were selected for larval sampling.

- **Water hyacinth float:** These were areas where water hyacinths were densely assembled into mats on floats. Two water hyacinth floats were selected for larval sampling.
habitats were only surveyed during this period, because they were not easily accessible due to dense vegetation during the wet season. Such habitats were within the vicinity of (i.e., could be seen from) the water hyacinth sampling sites. The non-water hyacinth habitats were categorized based on the types of vegetation cover: (1) tall vegetation (greater than approximately 1 m), (2) short vegetation including trailing plants (less than approximately 1 m), and (3) minimal vegetation (uncovered). Tall vegetation included papyrus and sedges, but not trees. The uncovered habitat was a distinct open area larger than approximately 1 m². All uncovered areas were surrounded by vegetation and not directly exposed to waves. Dipping sites were limited within approximately 5 m of the land.

Habitats adjacent to the lake
All lagoons in the study area were located in February 2009, and their geographical positions were recorded using a GPS. The stagnant water pools that were not associated with a sand bar were also located along the lakeshore. The surveyed water pools were categorised based on their composition: (1) lagoons, (2) other natural pools that were not separated by sand bars from the lake, and (3) man-made pools such as ditches and ponds that were dug to draw water from the lake for farming. The survey did not include temporary water pools such as animal hoof prints or small puddles because these habitats mostly occur on the fringes of larger stagnant pools. Despite the large number of these temporary pools, their importance in malaria transmission is expected to be low [10]. The survey was limited to the strip of land (within approximately 100 m at most from the lake) that had emerged along the shore after a recent reduction in the lake water [16].

When a water pool contained multiple distinct microhabitats, the habitats were categorised based on the types of main vegetation cover: (1) tall vegetation, (2) short vegetation including trailing plants, (3) floating plants, and (4) uncovered. Anophelines were sampled from each microhabitat.

Species identification
The sampled larvae were immediately preserved in 96% ethanol at the field site and identified microscopically in the lab using a morphological identification key [23]. The specimens belonging to the An. funestus and the An. gambiae complexes were further identified by species using the ribosomal DNA-polymerase chain reaction (PCR) method [24]. The specimens that were microscopically indistinguishable were also examined using PCR. The primers specific to An. funestus s.s., An. rivulorum, Anopheles leesoni, Anopheles parnsis, An. Arabiensis, and An. gambiae s.s. were used for the PCR.

Data analyses
A generalised linear model (GLM; the ‘stat’ and ‘MASS’ packages in R version 2.13.1) was used to determine whether the surrounding vegetation type (tree, non-woody plant or short plant) influence each anopheline species in the water hyacinths. Data sets from two sampling years were included in the initial model to assess the temporal effect and fix the effect; sampling year was included as an explanatory variable. Since the response variable (numbers of larvae) was counts, a poisson regression was applied to optimize the statistical model. A negative binomial regression was applied when data were overdispersed. When a data set included excess zeroes (absence of larvae), a zero-inflated regression model and a hurdle model (the ‘pscl’ package in R) were also applied. Both models are two-component mixture models combining a count regression part and a binomial regression part [25]. An optimal statistical model was selected using the Akaike Information Criteria (AIC). The significance of each explanatory variable remaining in the optimal model was estimated using a log-likelihood ratio test.

Using the same statistical procedure, the mosquito data set from water hyacinth habitats and non-water hyacinth habitats in the lake in 2009 was analyzed together, to determine whether surrounding vegetation type and microhabitat type based on vegetation cover (water hyacinth, covered with short vegetation, covered with tall vegetation, and uncovered) affect mosquitoes. Similarly, the effects of water pool type (lagoon, other natural pool, and man-made pool) on mosquitoes were estimated. Including both data sets from the lake and adjacent pools, the effects of microhabitat and macrohabitat (lake and pool) were also estimated. The dominant vector species in the lake habitats and adjacent pools were also determined using the same statistical methods. In this case, the response variable was the number of larvae, and the explanatory variable was the species.

In addition to the univariate analysis for each species, a redundancy analysis (RDA; the ‘vegan’ package in R) was also applied to the data. This multivariate method comprehensively analyses the relationships between various species and the explanatory variables. An RDA is more useful for a data set including many zeroes than is a canonical correspondence analysis because it is less sensitive to double zeroes [26]. A square root transformation was applied to the raw species data to down-weight the effect of abundant species. Subsequently, the chord transformation was applied to minimise the double-zero problem. The significance of all explanatory variables together was assessed using permutation tests. Additionally, each explanatory variable was tested, and the optimal model was then selected by excluding variables that were not significant. The significance of each axis was also assessed using permutation tests.

Ethics statement
This study was approved by the Scientific Steering Committee and National Ethics Review Committee of the Kenya Medical Research Institute (SSC No. 1310), and the ethics review committee of Nagasaki University. No specific permits were required for the described field studies. The study area did not include a national park or other protected area of land. However, we obtained consents from land owners when we collected mosquitoes on privately-owned lands. This field study did not involve endangered or protected species.

Results
Water hyacinth habitats in the lake
A total of 43 distinct water hyacinth mats were found along the lakeshore in 2008 (Figure 1). All mats were on the eastern shore. The lengths of the mats ranged from 27.2 to 1585.2 m (mean ± SE; 184.2±18.2 m). The total length of the mats was 7.9 km. Water hyacinths covered one third (33.8%) of the total shoreline surveyed and nearly half (48.8%) of the eastern shore.

In 2008, larvae were sampled from 93 water hyacinth sites; however, this decreased to 58 owing to a reduction in water hyacinths in 2009. Of the 151 sites, 36 (23.8%) contained water hyacinth habitats that were trapped or surrounded by trees. In the first survey, 797 anopheline larvae were collected from 69 sites, while 508 larvae were collected at 29 sites in the second survey. A total of 1047 larvae were identified by species or cryptic species complex. Three primary malaria vectors, An. funestus s.s., An. arabiensis and An. gambiae s.s., were recorded from the water hyacinths (Table 1). However, An. arabiensis and An. gambiae s.s. were excluded from the analysis, as their numbers were insufficient for determining the
dominant species in water hyacinth habitats. In addition, *Anopheles rivulorum*, *Anopheles lessonii*, *Anopheles pharoensis* and *Anopheles coustani* s.l. were recorded from the water hyacinths. Since *P. falciparum* were found in *A. rivulorum* in Tanzania, this species was also included in the statistical analysis [27,28]. Although *P. falciparum* were reported from *A. lessonii* and *A. pharoensis* [29,30], their numbers were insufficient for the statistical analysis. *Anopheles coustani* s.l. was also excluded from the analysis, because the sample might include multiple species, and the species group is considered unimportant for transmission [31].

A hurdle model with a negative binomial distribution had the lowest AIC, and the optimal model included the binomial portion only. *Anopheles rivulorum* occurred in the water hyacinth mats more frequently than *A. funestus* s.s. (P = 0.017). When *A. rivulorum* was analyzed separately to reveal the important vegetation types that trapped the water hyacinths, a GLM with negative binomial distribution revealed that the vegetation variable was statistically significant (P = 0.007). A multiple comparison test revealed that the density of the species in water hyacinths trapped by trees was significantly higher than under other conditions (Table 1). For *A. funestus* s.s., the vegetation variable was only significant in the binomial portion of the hurdle model (P<0.001), and its occurrence in the water hyacinths trapped by trees was significantly greater. The sampling year was not statistically significant for both species.

An RDA also indicated that the vegetation type that trapped water hyacinths was a significant variable (P<0.001). When the occurrence of *A. arabiensis* was regressed against the microhabitat categorized by vegetation cover and the vegetation type that surrounded the sampling sites, the microhabitat variable remained in the binomial portion of the hurdle model, while the surrounding vegetation variable remained in the count portion. The occurrence of *A. arabiensis* was significantly greater in short grass and uncovered habitats compared with the habitats containing tall vegetation (P<0.001). The density of *A. arabiensis* was significantly greater in the habitats surrounded by non-woody tall plants than those exposed to waves (P = 0.022). When *A. rivulorum* was regressed against these variables, a negative binomial hurdle model revealed that both variables were significant (P = 0.012 for microhabitat, and P<0.001 for surrounding vegetation type). The occurrence of this species was significantly greater in the water hyacinths located in the habitats surrounded by trees (P<0.001). The multivariate analysis also indicated that the vegetation surrounding the habitats and the microhabitat were significant variables (P<0.015 and P<0.001, respectively) (Figure 2B). The biplot indicated that *A. arabiensis* was positively associated with short grass or uncovered habitats, while *A. rivulorum* was positively associated with water hyacinths surrounded by trees.

### Habitats adjacent to the lake

A total of 95 distinct stagnant pools were identified along the lakeshore of these, 43 (45.3%) pools were lagoons, of which 34 (79.1%) were located along the western shore. The study area also included 44 (46.3%) man-made pools and eight (8.4%) other natural pools. The length of the lagoons ranged from 4.5 to 652 m (mean ± SE; 135.3±18.2 m). The total length of all lagoons was 6.0 km, with 4.6 km belonging to the western shore (56.8%), (Figure 1). Tall plants and floating plants were more strongly associated with lagoons. All floating-plant habitats included only water hyacinths, with the exception of one habitat that included both the water hyacinth and Nile cabbage. A total of 149 microhabitats within stagnant pools were sampled for mosquitoes in 2009 (Table 3). Most lagoons harboured multiple microhabitats. Because of the strong association between the water-pool type and the microhabitat, the association of vector species was analyzed separately for these two variables. A total of 5,934 anopheline larvae were collected from 117 sites. A total of

### Table 1. Mean densities ± SE (per site) and occurrences of anopheline larvae sampled from the water hyacinths that were surrounded by trees or other tall emergent plants, or that were exposed to waves in the lake in 2008 and 2009.

<table>
<thead>
<tr>
<th>Taxa</th>
<th>Trees (n = 36)</th>
<th>Non-woody tall plants (n = 71)</th>
<th>Short plants (exposed) (n = 44)</th>
<th>2008 (n = 93)</th>
<th>2009 (n = 58)</th>
<th>Total (n = 151)</th>
<th>No. of larvae (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>A. funestus</em> s.s.*</td>
<td>2.4±0.8 (13)</td>
<td>0.0±0.0 (0)</td>
<td>0.1±0.1 (2)</td>
<td>0.8±0.3 (11)</td>
<td>0.3±0.2 (4)</td>
<td>0.6±0.1 (15)</td>
<td>91 (8.7)</td>
</tr>
<tr>
<td><em>A. lessonii</em></td>
<td>0.1±0.1 (3)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>&lt;0.1 (2)</td>
<td>&lt;0.1 (1)</td>
<td>&lt;0.1 (3)</td>
<td>3 (0.3)</td>
</tr>
<tr>
<td><em>A. rivulorum</em></td>
<td>3.5±1.1 (14)</td>
<td>0.8±0.5 (12)</td>
<td>0.3±0.2 (4)</td>
<td>1.3±0.4 (22)</td>
<td>1.3±0.7 (8)</td>
<td>1.3±0.4 (30)</td>
<td>195 (18.6)</td>
</tr>
<tr>
<td><em>A. arabiensis</em></td>
<td>0.7±0.6 (4)</td>
<td>0.1±0.0 (3)</td>
<td>0.2±0.2 (2)</td>
<td>0.1±0.0 (4)</td>
<td>0.6±0.4 (5)</td>
<td>0.3±0.1 (9)</td>
<td>38 (3.6)</td>
</tr>
<tr>
<td><em>A. gambiae s.s.</em></td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0.1±0.1 (2)</td>
<td>&lt;0.1 (1)</td>
<td>&lt;0.1 (1)</td>
<td>&lt;0.1 (2)</td>
<td>3 (0.3)</td>
</tr>
<tr>
<td><em>A. pharoensis</em></td>
<td>0 (0)</td>
<td>0.2±0.2 (3)</td>
<td>0.1±0.0 (2)</td>
<td>0.1±0.1 (1)</td>
<td>0.3±0.2 (4)</td>
<td>0.1±0.1 (5)</td>
<td>20 (1.9)</td>
</tr>
<tr>
<td><em>A. coustani</em> s.l.</td>
<td>6.2±1.4 (24)</td>
<td>3.8±0.8 (38)</td>
<td>4.6±1.4 (18)</td>
<td>5.1±0.8 (59)</td>
<td>3.9±1.2 (20)</td>
<td>4.6±0.6 (79)</td>
<td>697 (66.6)</td>
</tr>
</tbody>
</table>

The numbers in parentheses after the density value indicate positive sites. The values with the same letter were not statistically significant (P>0.05). Where letters are missing on densities, the count portion was excluded from the optimal zero-inflated regression model or hurdle model. Similarly, the binomial portion was excluded where letters are missing on the numbers of positive sites.

*Species included in the analyses.

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5,673 larvae were identified, of which 5,085 (89.6%) were *An. arabiensis*.

GLMs with negative binomial distribution had the lowest values of AIC for explaining the relationships between mosquito abundance and water-pool type. The statistical analysis was only applied to *An. funestus* s.s. and *An. arabiensis*, because the numbers of the other species were insufficient. The optimal model included the variable of water-pool type for both species (P < 0.001 for both cases). The multiple comparison test indicated that the density of *An. funestus* s.s. in lagoons was significantly higher than that in the other pool types, whereas the density of *An. arabiensis* in man-made pools was significantly higher (Table 4). An RDA also revealed a significant effect of pool habitat type (P < 0.001) (Figure 2C).

Anopheles arabiensis was positively correlated with man-made and other natural pools, whereas *An. funestus* s.s. was positively associated with lagoons.

All habitats associated with the lake

When the data sets from the lake and the land habitats in 2009 were combined, a total of 7177 anopheline larvae were recorded from 229 of 382 sites, of which 6720 were identified (Table 5). Nearly 80% of the larvae were *An. arabiensis* followed by *An. funestus* s.s. and *An. rivulorum*. The numbers of the other species identified were insufficient for statistical analyses. A GLM with a negative binomial distribution revealed that the density of *An. arabiensis* was significantly higher than that of other species (P < 0.001). A GLM with a negative binomial distribution revealed that the densities of *An. arabiensis* and *An. funestus* s.s. were significantly lower in the lake habitats than the land habitats (P < 0.001 for both species); however, this difference was not significant for *An. rivulorum*. The microhabitat variable was significant for all cases (P < 0.001). The densities of *An. funestus* s.s. and *An. rivulorum* were significantly higher in the water hyacinths. The density of *An. arabiensis* was significantly higher in uncovered habitat.

The multivariate analysis also revealed that the variables of lake or land, as well as the microhabitat were significant (P < 0.001 and P < 0.001, respectively) (Figure 2D). The biplot indicated that *An. rivulorum* was positively associated with water hyacinths, tall plants, and lake, while *An. arabiensis* was associated with short grass, uncovered habitat, and land habitat. The association of *An. funestus* s.s. with the micro- and macro-habitats was not as clear as for the other species.
Lake habitats

This study confirms that African malaria vectors breed in Lake Victoria. Surprisingly, *An. arabiensis* was the most dominant vector species in the lake sampling sites, contrary to previous studies which reported this vector to mainly breed in small and sunlit temporary pools [8–10]. This species was mostly found in the lake areas covered with short grass, as well as uncovered areas. However, *An. arabiensis* was less common in the areas that were exposed to waves. Tall emergent plants surrounding the habitats must create an isolated and sheltered environment similar to breeding sites on land. The number of positive sites and their density suggests that the occurrence of this species in the lake was extensive. While larvae may accidentally enter the lake from adjacent pools on the land during heavy rainfall [32], the possibility of oviposition in the lake habitats should not be excluded. On the other hand, the lake habitats may serve as reservoirs; migration from the lake habitats may contribute to breeding in newly formed adjacent pools.

A small number of *An. gambiae* s.s. also occurred in the lake. This species is less common in this study area, which may have been caused by the increase in bed net coverage [33,34]; therefore, low density and occurrence do not necessarily mean that this species is less adapted to the lake habitats as compared with *An. arabiensis*. Both species belong to the same species complex, and they are often sympatric [9,22]. This is the first study to report both *An. arabiensis* and *An. gambiae* s.s. in a lake. Although they are the most important malaria vectors and have been studied extensively, few studies have explored their presence in lake habitats.

Water hyacinth habitats

This study also confirmed that malaria vectors are able to breed amongst water hyacinth mats in Lake Victoria. Although larvae of the *An. funestus* complex were reported from a water hyacinth mat in the same study area [10], this is the first study to confirm the occurrence of malaria vectors within several water hyacinth mats in the lake. Other vector and potential vector species were also found in the water hyacinth mats in the lake; however, the results from the univariate and multivariate analyses suggest that, as compared with the other microhabitats in the lake, the water hyacinth habitat is more suitable for *An. funestus* s.s. than *An. rivulorum*. Conversely, fewer larvae were found in the water hyacinth traps by non-woody emergent plants and in those exposed to waves. This suggests that the trees minimize the effects of waves and thereby stabilize the habitat.

The water hyacinth habitat associated with trees was more likely to occur in such habitats, but this tendency was greater for *An. funestus* s.s. than *An. rivulorum*. Conversely, fewer larvae were found in the water hyacinth traps by non-woody emergent plants and in those exposed to waves. This suggests that the trees minimize the effects of waves and thereby stabilize the habitat.

Table 2. Mean densities ± SE (per site) and occurrences of anopheline larvae sampled from the water hyacinth and non-water hyacinth habitats that were surrounded by trees or other tall emergent plants or were exposed to waves in the lake, and from microhabitats categorized based on vegetation cover at each sampling site in 2009.

<table>
<thead>
<tr>
<th>Taxa</th>
<th>Surrounded by</th>
<th>Microhabitats</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Trees (n = 55)</td>
<td>Non-woody tall plants (n = 146)</td>
<td>Water hyacinth</td>
</tr>
<tr>
<td><em>An. funestus</em> s.s.</td>
<td>0.3 ± 0.2 (3)</td>
<td>0.1 ± 0.1 (1)</td>
<td>0.3 ± 0.2 (4)</td>
</tr>
<tr>
<td><em>An. lessoni</em></td>
<td>&lt;0.1 (1)</td>
<td>0 (0)</td>
<td>&lt;0.1 (1)</td>
</tr>
<tr>
<td><em>An. rivulorum</em></td>
<td>1.1 ± 0.7 (10)</td>
<td>0.1 ± 0.1 (4)</td>
<td>0.3 ± 0.3 (1)</td>
</tr>
<tr>
<td><em>An. arabiensis</em></td>
<td>0.8 ± 0.4 (8)</td>
<td>1.7 ± 0.5 (24)</td>
<td>0.5 ± 0.3 (5)</td>
</tr>
<tr>
<td><em>An. gambiae</em> s.s.</td>
<td>0 (0)</td>
<td>0.1 ± 0.1 (4)</td>
<td>0.1 ± 0.1 (1)</td>
</tr>
<tr>
<td><em>An. pharoensis</em></td>
<td>0 (0)</td>
<td>0.3 ± 0.1 (10)</td>
<td>0.1 ± 0.0 (2)</td>
</tr>
<tr>
<td><em>An. maculipalpis</em></td>
<td>&lt;0.1 (1)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td><em>An. coustani</em> s.l.</td>
<td>6.4 ± 1.2 (37)</td>
<td>0.9 ± 0.2 (28)</td>
<td>3.4 ± 1.8 (7)</td>
</tr>
</tbody>
</table>

The number in parentheses below the density value indicates positive sites. The values with the same letter were not statistically significant (P > 0.05). Where letters are missing on densities, the count portion was excluded from the optimal zero-inflated regression model or hurdle model. Similarly, the binomial portion was excluded where letters are missing on the numbers of positive sites.

*Species included in the analyses.

doi:10.1371/journal.pone.0032725.t002

Discussion

Lake habitats

This study confirms that African malaria vectors breed in Lake Victoria. Surprisingly, *An. arabiensis* was the most dominant vector species in the lake sampling sites, contrary to previous studies which reported this vector to mainly breed in small and sunlit temporary pools [8–10]. This species was mostly found in the lake areas covered with short grass, as well as uncovered areas. However, *An. arabiensis* was less common in the areas that were exposed to waves. Tall emergent plants surrounding the habitats must create an isolated and sheltered environment similar to breeding sites on land. The number of positive sites and their density suggests that the occurrence of this species in the lake was extensive. While larvae may accidentally enter the lake from adjacent pools on the land during heavy rainfall [32], the possibility of oviposition in the lake habitats should not be excluded. On the other hand, the lake habitats may serve as reservoirs; migration from the lake habitats may contribute to breeding in newly formed adjacent pools.

A small number of *An. gambiae* s.s. also occurred in the lake. This species is less common in this study area, which may have been caused by the increase in bed net coverage [33,34]; therefore, low

Table 3. Numbers of microhabitats sampled in the three types of water pools that were adjacent to the lake.

<table>
<thead>
<tr>
<th>Water-pool type</th>
<th>Water hyacinth</th>
<th>Short</th>
<th>Tall</th>
<th>Open</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lagoons</td>
<td>9</td>
<td>30</td>
<td>20</td>
<td>24</td>
<td>83</td>
</tr>
<tr>
<td>Other natural pools</td>
<td>0</td>
<td>7</td>
<td>0</td>
<td>8</td>
<td>15</td>
</tr>
<tr>
<td>Man-made pools</td>
<td>0</td>
<td>11</td>
<td>0</td>
<td>40</td>
<td>51</td>
</tr>
<tr>
<td>Total</td>
<td>9</td>
<td>48</td>
<td>20</td>
<td>72</td>
<td>149</td>
</tr>
</tbody>
</table>

doi:10.1371/journal.pone.0032725.t003
Habitats adjacent to the lake

Nearly half of stagnant water pools were lagoons, and most lagoons were located along the western shore and around the northern tip of the peninsula. The landscape and geomorphological features appear to largely determine where lagoons and water hyacinths occur. Most lagoons were large enough to host various types of vegetation, which, in turn, offer a variety of microhabitats.

In contrast, the other types of natural and man-made pools had neither tall emergent plants nor floating plants. As a result, lagoons had greater densities of *An. funestus* s.s. and *An. gambiae s.s.*. However, to determine if lagoons produce more *An. arabiensis* compared with other vectors, the area covered by each microhabitat within lagoons must be estimated. If the area covered by vegetated habitats is larger than the area covered by open areas, the total production of *An. funestus* s.s. in lagoons may be higher than that of *An. arabiensis*.

Similarly, the production of malaria vectors in each pool type cannot be estimated using density alone. In fact, due to the high productivity of *An. arabiensis*, vector densities in man-made pools were much greater than those in lagoons and other natural pools. Although this study only measured the lengths of lagoons, the sizes of other pool types were obviously smaller than most lagoons. For

<table>
<thead>
<tr>
<th>Taxa</th>
<th>Macrohabitats</th>
<th>Microhabitats</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Lake (n = 233)</td>
<td>Water hyacinth</td>
<td>Short</td>
</tr>
<tr>
<td></td>
<td>Pool (n = 149)</td>
<td>(n = 68)</td>
<td>(n = 112)</td>
</tr>
<tr>
<td><em>An. funestus</em> s.s.*</td>
<td>0.1 ± 0 (4)</td>
<td>1.4 ± 0.4 (26)</td>
<td>1.2 ± 0.7a (8)</td>
</tr>
<tr>
<td><em>An. gambiae</em> s.s.</td>
<td>0.1 ± 0.0 (5)</td>
<td>0.4 ± 0.2 (8)</td>
<td>0.1 ± 0.1 (2)</td>
</tr>
<tr>
<td><em>An. pharoensis</em></td>
<td>0.2 ± 0.1 (12)</td>
<td>0.2 ± 0.2 (3)</td>
<td>0.2 ± 0.2 (4)</td>
</tr>
<tr>
<td><em>An. maculipalpis</em></td>
<td>&lt;0.1 (1)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td><em>An. coustani</em></td>
<td>2.5 ± 0.4 (72)</td>
<td>1.7 ± 0.4 (40)</td>
<td>3.5 ± 1.0 (22)</td>
</tr>
</tbody>
</table>

The number in parentheses after the density value indicates positive sites. The values with the same letter were not statistically significant (P > 0.05). Where letters are missing on densities, the count portion was excluded from the optimal zero-inflated regression model or hurdle model. Similarly, the binomial portion was excluded where letters are missing on the numbers of positive sites.

*Species included in the analyses.

doi:10.1371/journal.pone.0032725.t005

Table 4. Mean density ± SE (per site) and occurrences of anopheline larvae sampled from each type of water pool in 2009.

Table 5. Mean densities ± SE (per site) and occurrences of anopheline larvae sampled from the lake and adjacent habitats, and from microhabitats categorized based on vegetation cover at each sampling site in 2009.

The number in parentheses after the density value indicates positive sites. The values with the same letter were not statistically significant (P > 0.05). Where letters are missing on densities, the count portion was excluded from the optimal zero-inflated regression model or hurdle model. Similarly, the binomial portion was excluded where letters are missing on the numbers of positive sites.

*Species included in the analyses.

doi:10.1371/journal.pone.0032725.t004
example, most man-made pools were round, and their diameters were clearly less than five metres, whereas the widths of lagoons often exceeded five metres. The greater total length and number of lagoons imply that the total production of malaria vectors is greater for lagoons than for the other pool types. Moreover, the size of the lagoons indicates that they likely persist throughout the year, which would increase their total vector production [10]. Thus, lagoons likely play a more important role in local malaria transmission than do the other pool types along the shore of Lake Victoria. This hypothesis is further supported by the finding that lagoons are a major habitat of *An. funestus* s.s.. This species is more endophilic and anthropophilic than *An. arabiensis*; therefore, the former species is considered a more efficient vector [36–38].

Although this study did not examine inland water pools, previous studies have suggested that pools along the shore are more productive in the study area [12,18]. Inland pools are mostly temporary, and they mainly produce vectors during the wet seasons [12,18,39,40]. In contrast, constant seepage water from the lake stabilizes the pools along the shore unless the lake water level fluctuates considerably [16]; therefore, pools along the shore are likely to produce malaria vectors throughout the year. Other possible vector sources include rivers and large reservoirs. However, this study area lacks permanent rivers with vegetated habitats and reservoirs. Small puddles may appear on dry beds of temporal streams, but most are temporary.

**Implications for vector control**

The current study confirmed that several breeding sites are associated with Lake Victoria. Given that Lake Victoria is the second largest lake in the world, the habitats associated with the lake must be extensive, and their effects on local malaria transmission likely to be substantial. Although, further studies are necessary to estimate the effects of these habitats and the extent to which they contribute to local transmission, adjacent pool habitats require special attention.

**References**


