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Context-dependent activation of Wnt signaling by tumor suppressor RUNX3 in gastric cancer cells

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RUNX3 is a member of the runt-related transcription factor RUNX family and was originally identified as a tumor suppressor of gastric cancer development.¹⁻⁴ In approximately 80% of gastric cancers, RUNX3 expression is lost due to epigenetic silencing and mislocalization in the cytoplasm.¹⁻³,⁵ Moreover, expression of RUNX3 in gastric cancer cells results in suppression of tumorigenicity, while expression of the mutant form of RUNX3 R122C found in human gastric cancer does not affect tumorigenicity.¹⁻⁶ Consistently, gastric epithelial cells derived from Runx3⁻⁻ mouse intestinal mucosa without any alteration of the expression levels of TCF4 and β-catenin, and Runx3⁺⁺ mice develop intestinal tumors.⁶⁻⁻ Notably, the association of the mutant RUNX3 R122C with TCF4 is weaker than wild-type RUNX3; thus, RUNX3 plays a tumor-suppressing role in gastric cancer cells through a Wnt-independent mechanism. These results indicate that RUNX3 can either suppress or activate the Wnt signaling pathway through its binding to the TCF4/β-catenin complex by cell context-dependent mechanisms.

RUNX3 is a tumor suppressor for a variety of cancers. RUNX3 suppresses the canonical Wnt signaling pathway by binding to the TCF4/β-catenin complex, resulting in the inhibition of binding of the complex to the Wnt target gene promoter. Here, we confirmed that RUNX3 suppressed Wnt signaling activity in several gastric cancer cell lines; however, we found that RUNX3 increased the Wnt signaling activity in KatoIII and SNU668 gastric cancer cells. Notably, RUNX3 expression increased the ratio of the Wnt signaling-high population in the KatoIII cells, although the maximum Wnt activation level of individual cells was similar to that in the control. As found previously, RUNX3 also binds to TCF4 and β-catenin in KatoIII cells, suggesting that these molecules form a ternary complex. Moreover, the ChIP analyses revealed that TCF4, β-catenin and RUNX3 bind the promoter region of the Wnt target genes, Axin2 and c-Myc, and the occupancy of TCF4 and β-catenin in these promoter regions is increased by the RUNX3 expression. These results suggest that RUNX3 stabilizes the TCF4/β-catenin complex on the Wnt target gene promoter in KatoIII cells, leading to activation of Wnt signaling. Although RUNX3 increased the Wnt signaling activity, its expression resulted in suppression of tumorigenesis of KatoIII cells, indicating that RUNX3 plays a tumor-suppressing role in KatoIII cells through a Wnt-independent mechanism. In contrast to these findings, we present the unexpected finding that RUNX3 activates Wnt signaling in KatoIII and SNU668 gastric cancer cells. Interestingly, RUNX3 binds TCF4 and β-catenin also in the KatoIII cells, and binding of the complex to Wnt target gene promoter is more stable in the presence of RUNX3, which may cause Wnt signaling activation. Accordingly, it is possible that RUNX3 can either suppress or activate Wnt signaling activity by binding to the TCF4/β-catenin complex, and the direction of Wnt signaling modulation may be regulated by a cell context-dependent mechanism.
Materials and Methods

Cell culture experiments. Human gastric cancer cell lines, AGS (ATCC), AZ521, MKN45, KatoIII, (RIKEN, BioResource Center, Tsukuba, Japan), SNU216, SNU484, SNU601, SNU638, SNU668 and SNU719 (Korean Cell Line Bank, Seoul, Korea) were cultured in RPMI1640 supplemented with 10% FBS. The cell proliferation rate was examined using the Alamar Blue Cell Viability Reagent (Invitrogen, Carlsbad, CA, USA). For the soft agar colony formation assay, cells were suspended in 0.33% agarose contained in the medium and seeded on 0.5% bottom agar. After 21 days of culture, soft agar was stained with Giemsa solution (Wako, Osaka, Japan) and colony numbers were scored.

Cells were transfected with pcDNA3, pcDNA-Flag-RUNX3 or pcDNA-Flag-RUNX3(R122C) vector. (6) KatoIII-R3 stable cell line was constructed by transfection with pcDNA-RUNX3 and selected with G418 (Wako) at 100 µg/mL. To knock down gene expression, cells were transfected with Silencer Select siRNA for RUNX3 or β-catenin (Ambion, Cambridge, MA, USA).

To examine the Wnt activation level, cells were cotransfected with super 8× TOPFlash or Super 8× FOPFlash (Addgene, Cambridge, MA, USA), together with pcDNA3, pcDNA-Flag-RUNX3 or pcDNA-Flag-RUNX3(R122C). (6) At 24 h after transfection, the luciferase activity was measured using a Luciferase assay system (Promega, Madison, WI, USA).

Wnt suppression and activation. To inhibit Wnt signaling, cells were treated with 10 µg/mL of C59 (provided by Dr David Virshup), which inhibits porcupine, a membrane-bound O-acyltransferase required for Wnt palmitoylation. (14) To activate Wnt signaling, conditioned media including Wnt3a and Rspondin were prepared from L cells expressing Wnt3a and Rspondin, respectively (provided by Dr David Virshup), which inhibits porcupine, a membrane-bound O-acyltransferase required for Wnt palmitoylation. (14)

To activate Wnt signaling, conditioned media including Wnt3a and Rspondin were prepared from L cells expressing Wnt3a and Rspondin, respectively (provided by Dr Marc Leushacke), and the conditioned media were supplemented at 10% volume in the culture medium.

Western blotting. A total of 10 µg of protein samples were separated in 10% SDS-polyacrylamide gels. Antibodies for RUNX3(5) or unphosphorylated β-catenin (Millipore, Billerica, MA, USA) were used as the primary antibodies. The anti-β-actin antibody (Sigma, St. Louis, MO, USA) was used as an internal control, and the ECL detection system (GE Healthcare, Buckinghamshire, UK) was used to detect the signals.

Real-time RT-PCR. Total RNA was extracted using ISOGEN (Nippon Gene, Tokyo, Japan) and cDNA was constructed using the Prime Script RT Reagent Kit (Takara, Tokyo, Japan). Real-time RT-PCR was performed using the SYBR Premix Ex TaqII (Takara) and Stratagene Mx3000P (Agilent Technologies, Santa Clara, CA, USA). The primers were purchased from Takara.

Flow cytometry analysis. To examine the intracellular RUNX3 and β-catenin levels, permeabilized cells were incubated with the primary antibodies for total β-catenin (Sigma) or RUNX3, (5) followed by the secondary antibodies for rabbit IgG-conjugated with Alexa 488 (Molecular Probes, Grand Island, NY, USA) or mouse IgG-conjugated with Alexa 633 (Invitrogen), and examined using FACS Canto II (BD Biosciences, San Jose, CA, USA). Cells were transfected with a pcDNA-RUNX3-ires-mGFP expression vector, in which internal ribosome entry site (IRES) fragment from pTRE3G-IRES (Clontech Laboratories, Mountain View, CA, USA) and maxGFP cDNA from pmaxGFP (Lonza, Allendale, NJ, USA) were subcloned to pcDNA-Flag-RUNX3, and RUNX3-expressing cells were isolated using the FACS Aria cell sorter (BD Biosciences, San Jose, CA, USA) to collect GFP-expressing cells.

Immunocytochemistry. The cells were seeded on cover slips and fixed in 4% paraformaldehyde, then permeabilized with 0.5% Triton X-100 in PBS. Antibodies against total β-catenin (Sigma) or RUNX3 were used as the primary antibodies, and anti-rabbit IgG Alexa 594 or anti-mouse IgG Alexa 488 (Molecular Probes) were used as the secondary antibodies. (5)

Immunoprecipitation. KatoIII cells were transfected with the pcDNA-Flag-RUNX3 or pcDNA-Flag, and the cell lysates were used for immunoprecipitation with anti-FLAG M2 aga-rose (Sigma). Western blotting was performed using antibodies against unphosphorylated β-catenin (Millipore), TCF4 (Santa Cruz Biotechnology, Santa Cruz, CA), FLAG peptide or β-actin (Sigma).

ChIP. The cells were treated with formaldehyde solution (Wako) for crosslinking. ChIP was performed using the ChIP Assay kit EZ ChIP (Millipore) and antibodies against TCF4 (Santa Cruz Biotechnology), unphosphorylated β-catenin (Millipore), RUNX3(5) and mouse normal IgG. The primer sequences for the c-Myc promoter were: 5′TTGCTGGGTTATTTTAATCAT-3′ and 5′ACTGGTTTGGCAGACCCATCC-3′. (12)

For the Axin2 promoter, conserved TCF/LEF binding sites are localized in intron 1. (16) and Simple ChIP Human Axin2 Intron 1 Primers (Cell Signaling, Danvers, MA, USA) were used.

Statistical analysis. Statistical analyses were performed using the unpaired Student’s t-test, with P-values <0.05 considered significant.

Results

Wnt activation by RUNX3 expression in KatoIII and SNU668 cells. RUNX3 expression was detected by western blotting in AZ521 and MKN45 cells, while it was not detected in other gastric cancer cell lines (Fig. 1a). Consistently, high levels of RUNX3 mRNA were detected in AZ521 and MKN45 cells by RT-PCR (Fig. S1). We transiently transfected the RUNX3 expression vector to all gastric cell lines, and examined Wnt signaling activity by luciferase reporter analysis. The Wnt signaling activity was significantly decreased in SNU216, SNU601, SNU638 and SNU719 cells, which was consistent with the previous results. (12) However, RUNX3 expression increased Wnt signaling activity in KatoIII and SNU668 cells (Fig. 1b). Importantly, the R122C mutant form of RUNX3 that is defective in the RUNX3 function did not change Wnt signaling activity in these cells. (1, 6) The KatoIII-R3, stable RUNX3-expressing cells (Fig. S2) also exhibited an increased luciferase activity, which was suppressed by RUNX3 siRNA transfection (Fig. 1c). Moreover, Wnt activation levels increased gradually in accordance with the amount of the RUNX3 expression vector (Fig. 1d). Consistently, the expression levels of Wnt target genes, Sox4 and Axin2, increased significantly in KatoIII and SNU668 cells by RUNX3 expression (Fig. 1e).

Notably, inhibition of endogenous RUNX3 expression by siRNA in AZ521 significantly increased Wnt signaling activity, whereas RUNX3 siRNA transfection partially suppressed Wnt signaling in MKN45 cells (Fig. 1e.f). These results, taken together, suggest that RUNX3 suppresses or activates the Wnt signaling in a cell context-dependent mechanism.

Saturation of ligand-induced β-catenin stabilization in KatoIII cells. We further examined the RUNX3-induced Wnt activation mechanism using KatoIII cells. In KatoIII cells, Wnt signaling
is activated by β-catenin gene amplification. Treatment of control KatoIII cells with a Wnt ligand secretion inhibitor C59 significantly suppressed the endogenous Wnt signaling, indicating that Wnt ligand stimulation maintains the basal Wnt activation level (Fig. 2a). C59 treatment also decreased the luciferase activity in the RUNX3-expressing KatoIII cells. However, the ratio of the RUNX3-induced increase of luciferase activity in the C59-treated cells was similar to that in the control cells; that is, approximately 4.5-fold the control levels. Accordingly, it is possible that RUNX3 increases Wnt signaling activity in KatoIII cells through a ligand-independent mechanism.

We next examined the β-catenin levels of the RUNX3-transfected cells by western blotting. Although the active β-catenin levels were slightly increased both in the KatoIII-R3 cells and the RUNX3 vector-transiently transfected KatoIII cells (Fig. 2b, Figs S2 and S3), the increase was not sufficient to explain the marked increase of the TOPflash activity (Fig. 1b).

The Wnt activation level increased significantly in 293T cells following treatment with Wnt3a and/or Rspondin (Fig. 2c). However, treatment of KatoIII cells with Wnt3a/Rspondin did not change the luciferase activity (Fig. 2d). Consistently, the β-catenin high population measured by flow cytometry was significantly increased in the 293T cells following Wnt3a/Rspondin treatment, while the β-catenin high population in the KatoIII cells did not change following stimulation with Wnt3a/Rspondin (Fig. 2e,f). Therefore, it is possible that the Wnt ligand-induced β-catenin stabilization level is saturated in KatoIII cells, and RUNX3 activates Wnt signaling through mechanisms other than β-catenin stabilization.

Increase of β-cateninHI population by RUNX3 expression in KatoIII cells. We previously found that the Wnt activation level in individual KatoIII cells oscillates between a Wnt-high and
isolated by cell sorting, corresponding to RUNX3Hi-expressing and RUNX3 Lo-expressing cells, respectively (Fig. 4b). and we were unable to establish a GFP Hi (RUNX3Hi) cell
KatoIII cells. (19) Therefore, we reexamined the role of RUNX3
been reported that RUNX3 suppresses the tumorigenicity of
Importantly, proliferation of GFP Hi cells was significantly
the RUNX3-expressing cells using flow cytometry. When the
When the RUNX3 vector was transfected, the β-cateninHi population (top 50% level of the control in Fig. 3a, [left]) increased significantly in the RUNX3Hi cells (78%: Q2/(Q1 + Q2)), but not in the RUNX3Lo cells (45%: Q4/(Q3 + Q4)) (Fig. 3a [right], b). Notably, the maximum β-catenin level in the RUNX3-transfected cells was similar to that observed in the control (Fig. 3a). We confirmed that transfection efficiency was not different between β-cateninHi and β-cateninLo KatoIII cells (Fig. S4). Immunocytochemistry of the RUNX3 vector-transfected KatoIII cells showed that approximately 73% of the RUNX3-positive cells were also positive for β-catenin, whereas approximately 44% of the RUNX3-negative cells were β-catenin positive (Fig. 3c). These results indicate that RUNX3 expression increases the number of cells in the Wnt-high population of KatoIII cells.

Tumor suppressing function of RUNX3 in KatoIII cells. It has been reported that RUNX3 suppresses the tumorigenicity of KatoIII cells. (19) Therefore, we reexamined the role of RUNX3 in the tumorigenicity of KatoIII cells. The RUNX3 vector-transfected KatoIII cells exhibited slight but significant suppression of cell proliferation at 48 and 72 h after seeding (Fig. 4a), and RUNX3 siRNA increased proliferation KatoIII-R3 cells at 48 h after seeding (Fig. S5).

We next transfected the RUNX3-IRES-mGFP expression vector to KatoIII cells, and GFPHi and GFPLo cells were isolated by cell sorting, corresponding to RUNX3Hi-expressing and RUNX3Lo-expressing cells, respectively (Fig. 4b). Importantly, proliferation of GFPHi cells was significantly suppressed compared with that of the GFPLo cells (Fig. 4c), and we were unable to establish a GFPHi (RUNX3Hi) cell line due to the limited proliferation. Moreover, RUNX3-transfected KatoIII cells exhibited significant suppression of soft agar colony formation (Fig. 4d), and the number of colonies larger than 0.1 mm in diameter was significantly decreased to 5.6% of the control (Fig. 4e). Moreover, expression of cell cycle inhibitor p21WAF1/CIP1 (20) was increased significantly in KatoIII cells by RUNX3 expression (Fig. 4f), which was consistent with previous report. (20) These results indicate that RUNX3 plays a tumor-suppressing role in KatoIII cells, and that RUNX3-dependent Wnt activation is not sufficient to maintain the tumorigenicity of RUNX3Hi KatoIII cells.

Binding of the RUNX3 and TCF4 complex to the Wnt target gene promoters in KatoIII cells. It has previously been shown that RUNX3 binds to TCF4/β-catenin complex, which suppresses the binding of the complex to the Wnt target gene promoters. (12,13) Notably, immunoprecipitation experiments revealed that RUNX3 bound β-catenin and TCF4 also in the RUNX3-transfected KatoIII cells (Fig. 5a), suggesting that RUNX3, TCF4 and β-catenin form a ternary complex also in KatoIII cells.

We next performed ChIP analyses to examine whether the binding of the TCF4/β-catenin complex to the promoter of the Wnt target genes was suppressed in the RUNX3-expressing KatoIII cells. In the control KatoIII cells, TCF4 and β-catenin bound to the Axin2 and c-Myc gene promoters (Fig. 5b, Lanes 1 and 4). Importantly, TCF4 and β-catenin also bound to the Axin2 and c-Myc gene promoters in the RUNX3-expressing KatoIII cells, and their band intensities were higher than those of the control cells (Fig. 5b, Lanes 2 and 5). Consistently, ChIP-based real-time PCR analysis showed that the quantified DNA amount of the Axin2 and
Myc gene promoters that bound TCF4 and β-catenin was significantly higher in the RUNX3-expressing KatoIII cells compared with the control cells (Fig. 5c,d). Moreover, we confirmed that RUNX3 bound the Axin2 and c-Myc gene promoters in the RUNX3-expressing KatoIII cells (Fig. 5b, Lanes 2 and 5, and Fig. 5c,d). In contrast, the R122C mutant RUNX3 did not significantly increase the binding of TCF4 and β-catenin to the Axin2 and c-Myc gene promoters (Fig. 5b, Lanes 3 and 6, and Fig. 5c,d). These results indicate that the complex consisting of RUNX3, TCF4 and β-catenin binds to the promoter of Wnt target genes in KatoIII cells in a more stable fashion than the TCF4/β-catenin complex, which may increase the proportion of the Wnt-high cells in the RUNX3-expressing KatoIII cells.

Discussion

It has been demonstrated that RUNX3 binds to TCF4 and β-catenin, resulting in suppression of the binding of TCF4 to the Wnt target gene promoters, thereby suppressing the Wnt signaling. In the present study, we found that RUNX3 increased the Wnt activity in KatoIII and SNU668 cells. RUNX3 bound TCF4 and β-catenin also in the KatoIII cells, and the complex was stabilized to bind Wnt target gene promoter, which is in contrast to the previous findings.

The important question is how RUNX3 activates Wnt signaling in KatoIII and SNU668 cells. It is known that β-catenin replaces the transcriptional repressor, Groucho, from
TCF in the nucleus, and recruits cofactors such as Bcl9 and pygopus to the TCF4/β-catenin complex, inducing target gene transcription.[21,22] It has also been reported that additional factors are required for the recruitment of β-catenin to the target gene promoters. Transducin β-like protein 1 (TBL1) and its related family member, TBLR1, recruit β-catenin to TCF on the Wnt target gene promoter.[22,23] TBL1 can bind both TCF4 and β-catenin, suggesting that it strengthens their physical association, which may contribute to Wnt activation. Jerky also recruits β-catenin to chromatin, and promotes the association of β-catenin and LEF1, resulting in the induction of Wnt target gene expression.[22,24] We found that RUNX3 increases the occupancy of the TCF4/β-catenin complex in KatoIII cells, suggesting that RUNX3 plays a role in the stabilization of the TCF4/β-catenin complex on the Wnt target gene promoter like TBL1 and Jerky. It is also possible that RUNX3 is involved in the recruitment of β-catenin to TCF4 on the Wnt target gene promoter like these molecules. We have previously shown that the Wnt activation levels are oscillating in the individual KatoIII cells,[18] and in this study, we showed that the RUNX3 expression increases the ratio of the Wnt-high population. Accordingly, it is conceivable that RUNX3 maintains the Wnt activation at a high level, suppressing the decrease of Wnt activity by stabilizing the TCF4/β-catenin complex on the Wnt target gene promoters.

Another unsolved question is how the RUNX3/TCF4 complex can bind to the DNA of the Wnt target gene promoter in KatoIII cells. As previously described, RUNX3-bound TCF4 cannot bind chromatin, possibly due to the competition for the DNA binding region of TCF4 with RUNX3. It is possible that cofactor(s) in the complex affect the conformation of the RUNX3/TCF4/β-catenin complex. Genetic polymorphisms of such cofactor(s) may cause conformational changes of the complex, which allows RUNX3-bound TCF4 to bind the Wnt target gene promoter. It would be worth examining the DNA sequences of cofactors of the TCF4/β-catenin complex in KatoIII and SNU668 cells to understand the molecular mechanism(s) responsible for the RUNX3-associated Wnt regulation.

In the present study, we also confirmed that RUNX3 transfection significantly suppressed the proliferation and tumorigenicity of KatoIII cells with induction of p21 expression. Moreover, RUNX3 has been shown to induce the apoptosis of gastric cancer cells by upregulating apoptosis-related genes.[25] It is thus possible that these gene products suppressed the proliferation and survival of RUNX3-expressing KatoIII cells, and that RUNX3-induced Wnt activation is not sufficient to protect KatoIII cells from RUNX3-induced apoptosis. It remains to be investigated whether RUNX3 suppresses tumorigenicity also in SNU668 cells. However, it has been reported that RUNX3 is upregulated and functions as an oncogene in head and neck cancer cells by increasing the proliferation and sphere formation.[26,27] Accordingly, the RUNX3-associated tumor suppressing functions are likely inactivated in head and neck cancer cells, and, therefore, it is possible that RUNX3-associated Wnt activation contributes to tumorigenesis in such cancer cells.

The present results, together with previous findings,[12,13] indicate that RUNX3 can either suppress or activate Wnt signaling through binding to the TCF4/β-catenin complex. Although it remains to be investigated, cofactor(s) that bind the RUNX3/TCF4/β-catenin complex may be involved in the decision regarding the direction of Wnt signaling modulation; that is, suppression or activation.

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References


Supporting Information

Additional supporting information may be found in the online version of this article:

Fig. S1. Relative RUNX3 mRNA levels in gastric cancer cell lines examined by RT-PCR.

Fig. S2. Western blotting results of RUNX3 and active form of β-catenin in Kat0II cells and Kat0III-R3 cells.

Fig. S3. Relative band intensities calculated from the Western blotting results (Fig. 2b) of active β-catenin and RUNX3 in the RUNX3 expression vector-transfected Kat0II cells.

Fig. S4. Flow cytometry analyses of β-catenin and GFP in the control Kat0III cells and GFP expression vector-transfected Kat0III cells.

Fig. S5. Cell proliferation rates of RUNX3 siRNA-transfected Kat0III-R3 cells.

Disclosure Statement

The authors have no conflict of interest.