Estrogen and Progesterone Receptor Expression by Macrophages and Regulation of Hepatocyte Growth Factor by Ovarian Steroids in Women with Endometriosis

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Abstract

**Background:** The production of hepatocyte growth factor (HGF) by mesenchymal cells and macrophages (Mφ) in response to different inflammatory mediators and its involvement in the growth of endometriosis has been described. However, information regarding Mφ-mediated regulation of HGF by ovarian steroid hormones in women with endometriosis is limited. Therefore, we planned to investigate the regulation of HGF by steroid hormones in isolated Mφ and stromal cells derived from women with or without endometriosis. **Methods:** We isolated CD68 immunoreactive adherent Mφ in vitro from 46 women with endometriosis and 30 women without endometriosis. The expression of estrogen receptor (ER) and progesterone receptor (PR) by Mφ was demonstrated by immunohistochemistry and RT-PCR. The production of HGF in the culture media of basal and ovarian steroid-stimulated Mφ was examined by enzyme-linked immunosorbent assay. The expression of mRNA for HGF and its receptor, c-Met in the Mφ and stromal cells in response to ovarian steroid was investigated by RT-PCR. The single and combined effect of HGF and estrogen on the growth of Mφ and stromal cells was analyzed by bromodeoxyuridine (BrdU) incorporation. **Results:** ER
and PR were expressed in isolated Mφ and intact tissue both at the protein and mRNA levels. Macrophages derived from women with endometriosis produced significantly higher concentration of HGF (352.2 ± 4.9 pg/mL) in the conditioned media after treatment with estradiol (10⁻⁸M) than that of basal Mφ (221.5 ± 32.8 pg/mL, p<0.05) or women without endometriosis (170.6 ± 2.6 pg/mL, p<0.05). These effects were less evident after treatment with progesterone. An anti-estrogenic treatment of these Mφ with tamoxifen (10⁻⁶M) reversed the production of HGF and other macromolecules towards non-treated Mφ. It was interesting to observe that secretion of HGF in response to ovarian steroids was further enhanced after activation with lipopolysaccharide. The mRNA expressions of HGF and its receptor, c-Met were also detected in Mφ and stroma in response to estrogen, suggesting an autocrine regulation. HGF mRNA expression was higher in these cells of women with endometriosis than that of non-endometriosis without displaying any phase difference of menstrual cycle. The bromodeoxyuridine (BrdU) incorporation study indicated that exogenous stimulation with HGF and estrogen, either alone or in combination, significantly increased the cell proliferation of both endometrial stroma and Mφ comparing to that of non-endometriosis or non-treated cells.
**Conclusion:** These results suggested that besides other inflammatory mediators, ovarian steroids also participate in the production of HGF by peritoneal Mφ which may be involved in the growth of endometriosis either alone or in combination with estrogen.

**Key words:** estrogen receptor / progesterone receptor / macrophage / peritoneal fluid / hepatocyte growth factor / ovarian steroids / lipopolysaccharide / endometriosis
Introduction

Endometriosis, the presence of functional endometrium outside of the uterine cavity, is a common disease, causing abdominal pain, dysmenorrhea, dyspareunia and infertility in about 10% of the female population (Strathy et al., 1982). Besides metaplastic transformation of endometrial and peritoneal mesothelial cells, the transplantation, implantation and growth of exfoliated menstrual debris on the peritoneal and ovarian surfaces are the widely accepted mechanisms of endometriosis (Sugawara et al., 1997; Ishimaru et al., 2004; Sampson J, 1927; Thomas et al., 1992). A number of literatures have already demonstrated the potential role of ovarian steroid hormones in the regeneration of endometrium after menstruation and the growth of endometriosis (Fujishita et al., 1997; Nisolle et al., 1997). However, as a non-self lesion in pelvic environment, the growth or persistence of endometriosis can also be regulated by innate immune system. The mitogenesis or angiogenesis of eutopic and ectopic endometrium possibly involves an extensive interplay between endometrial cells, inflammatory cells, ovarian hormones, soluble factors and the extracellular matrix (Folkman et al., 1987).

As a cell component of innate immune system, peritoneal fluid (PF) and intact
tissue derived from women with endometriosis have been shown to contain higher numbers of activated macrophages (Halme et al., 1987; Khan et al., 2002a, 2004a) than that found in women without endometriosis. This results in the secretion of higher concentrations of growth factors including hepatocyte growth factor (HGF) and other cytokines in PF as produced by the stimulated-Mφ in these patients (Halme et al., 1988, 1989, Khan et al., 2002b). This indicates that the growth or persistence of endometriosis is a normal inflammatory response.

Since mesenchymal cells retain estrogen receptor, production of different cytokines by endometrial stromal cells and its modulation by estrogen has been demonstrated (Tabibzadeh et al., 1989). Considering that infiltrated Mφ is one of the cell components of endometriotic lesion in pelvic environment, reports describing expression of steroid receptors by Mφ and the secretion of different macromolecules in response to steroid hormones are scanty.

The details of HGF and its receptor, c-Met and the biological activity of this ligand-receptor has already been described (Khan et al., 2005). HGF production by mesenchymal cells and Mφ in response to different inflammatory mediators and its
involvement in the growth of endometriosis has been reported (Sugawara et al., 1997; Jiang et al., 1999; Khan et al., 2003a, 2005). However, information regarding ovarian steroid-mediated production of HGF by peritoneal Mφ is yet to be determined. Therefore, we report for the first time the production of HGF by the peritoneal Mφ in response to ovarian steroids in women with or without endometriosis and examined the effect of HGF on the growth of endometrial cells either alone or in combination with estrogen.

Materials and Methods

Reagents. Culture media: RPMI-1640 medium for macrophage and Dulbecco’s Modified Essential Medium (DMEM):Ham’s F12 medium for stromal cells and were supplemented with 100 IU/mL of penicillin G, 50 mg/mL of streptomycin, 2.5 μg/mL of amphotericin B (GIBCO, Grand Island, NY). Fetal bovine serum (FBS), 17β-estradiol (E2), progesterone (P), hydroxytamoxifen (TMX), lipopolysaccharide (LPS, derived from Escherichia coli, serotype 0111:B4) were purchased from Sigma Chemical Co. (St. Louis, MO). Recombinant hepatocyte growth factor (HGF) was from R&D
system (Quantikine, R & D system, Minneapolis, MN).

**Subjects.** Women between 20 and 42 years old who were undergoing either diagnostic laparoscopy for dysmenorrhea or elective laparoscopy for infertility were recruited for this study. Ectopic and corresponding eutopic endometrial tissue samples were collected from eight women containing blood-filled red lesions and two women harboring mixed black and white lesions. Eutopic endometrium from ten control women was also collected for parallel study by immunohistochemistry. Peritoneal fluid (PF) was obtained from 46 women with endometriosis and cycle matched 30 women with non-endometriosis. The control group, between 18 to 32 years old, consisted of fertile women without any evidence of endometriosis and operated for dermoid cyst by laparoscopy.

The production of macromolecules in the culture media of basal and stimulated macrophages and cell proliferation assay were studied in six women with endometriosis and six women without endometriosis (three each in proliferative phase and three in secretory phase). The extent of the disease was staged according to the revised-classification of American Society of Reproductive Medicine (revised-ASRM,
1997). All biopsy specimens and PF were collected in accordance with the guidelines of the Declaration of Helsinki. This study protocol was approved by the Institutional Review Board (IBR) of Nagasaki University. An informed consent was obtained from all women.

The distribution of patients in different revised-ASRM staging of endometriosis was as follows: stage I-II, n=24 and stage III-IV, n=22. Neither the study group nor the endometriosis free group had been on hormonal medication in the 3 months prior to the surgical procedure. All control women and women with endometriosis had regular menstrual cycles (28-32 days). The phase of the menstrual cycle was determined by histological dating of eutopic endometrium samples taken simultaneously with the peritoneal fluid samples. Menstrual dating was carried out by an independent pathologist. All induced menstrual cycles were excluded from the current study. The distribution of patients in different menstrual cycles is as follows: control women, proliferative phase, n=10; secretory phase, n=20; endometriosis women, proliferative phase, n=20; secretory phase, n=23 and menstrual phase, n=3.

Peritoneal lesions of endometriosis were diagnosed by their macroscopic appearance according to published criteria (Jansen et al., 1996) and categorized as red,
black and white lesions as proposed in the latest revision of the ASRM classification (ASRM, 1997). As we described recently (Khan et al., 2004b), the grouping of patients according to color appearance of endometriosis for our current study was done as follows: (1) women with red lesion (n=9), women containing blood-filled opaque red lesions or non-opaque transparent and/or translucent red lesions; (2) women with combined black and white lesions (n=22), women containing dominant distribution of black or white lesions without existence of any opaque red lesion; (3) women with chocolate cysts (n=15), women containing variable sizes of chocolate cysts with scanty distribution of either black lesion or white lesion or both but without any evidence of red lesion.

**Isolation of peritoneal fluid macrophages.** Peritoneal fluid was aspirated from the posterior cul-de-sac. Peritoneal fluid Mϕ were isolated by the method of Halme et al. (1987), and this method has been extensively characterized before (Helme et al., 1987; Khan et al., 2005). It results in an enriched population of Mϕ (>95%) which are not activated during the isolation procedure and are capable of being maintained as viable cultures for up to 72 hour. Isolated PF macrophages for immunohistochemistry were spotted onto untreated four-well chamber slides (Nunc, Naperville, IL) and the purity of
the preparation was determined using immunohistochemical staining using CD68 (KP1), a mouse monoclonal antibody for M\(\phi\) (1:50 dilution) from Dako, Denmark. The detail procedures of immunohistochemical staining are described elsewhere (Khan et al., 2004a, 2005). The adherent cells on the slides routinely contained >95% macrophages. Non-immune mouse immunoglobulin (Ig) G1 antibody in 1:50 dilution was used as a negative control. A counter staining of M\(\phi\) was also performed and we did not find any contaminating cells in isolated M\(\phi\). The concentration of M\(\phi\) in PF was counted by a hemocytometer counter.

**Immunoeexpressions of estrogen receptor (ER) and progesterone receptor (PR) by isolated macrophages and in intact tissue.** In order to immunolocalize ER and PR in the CD68 immunoreactive isolated M\(\phi\) and to demonstrate that ER and PR are being synthesized and expressed by M\(\phi\) in intact tissue, we performed immunohistochemistry using respective antibody and using serial section of eutopic and ectopic endometrium derived from women with or without endometriosis. The details of ER and PR antibody and procedure of immunohistochemistry were described previously (Fujishita et al., 1997; Nisolle et al., 1997). Non-immune mouse immunoglobulin (Ig) G1
antibody (1:50) was used as a negative control.

**Human endometrial cell cultures.** Primary endometrial cell cultures were prepared from the biopsy specimens of the eutopic and ectopic endometrium derived from six women with endometriosis and six women without endometriosis (three each in the proliferative phase and three in the secretory phase). Glandular epithelial cells were separated from stromal cells and debris by filtration through narrow gauge sieves. The characteristics of stromal cells were determined by morphological and vimentin-positive immunocytochemical studies. Stromal cells were sub-cultured to eliminate contamination by Mφ or other leukocytes, and experiments were performed at passage 1. The detail procedures for the isolation of endometrial stromal cells were described previously (Sugawara et al., 1997; Osteen et al., 1989, Khan et al., 2005). The specificity of the immunocytochemical staining was confirmed by the deletion of the first antibody. A counter-staining of stroma was also performed to exclude the contamination by epithelial cell or endothelial cells in isolated stromal cell culture (data not shown).

**Activation and ovarian steroid stimulation of peritoneal fluid macrophages.** Isolated peritoneal fluid Mφ as derived from six women with
endometriosis and six women without endometriosis were plated in 96-well microplates and allowed to attach for 24 hour in phenol red free media plus 5% FBS. After this time the cells were washed with phosphate-buffered saline (PBS) and incubated in serum free and phenol red free RPMI-1640 media containing either E2 (10^{-8}\text{M}), P (10^{-6}\text{M}) or a combination of both, for another 24 hr. These doses of E2 and P were used according to a previous study protocol (McLaren et al., 1996). Control cells were incubated in just phenol red free RPMI-1640 media. In order to examine the blocking effect of estrogen receptor on the secretion of different macromolecules in the M\textit{\phi} condition media, we performed extended experiment with tamoxifen (10^{-6}\text{M}). This dose was used as described in a previous report (Gockerman et al., 1986). Activation studies involved the addition of 5 ng/mL of LPS just before commencement of the 24 hr incubation with E2 and P. This dose of LPS was selected from a previous dose-dependent study from our laboratory that showed a maximum activation of M\textit{\phi} (Khan et al., 2005). The condition media were collected in triplicate, pooled, and frozen at -70\textdegree C until testing.

**Cytokine assays in the condition media.** The concentrations of HGF and VEGF in the culture media of treated- and non-treated-M\textit{\phi} were measured in duplicate
using a commercially available sandwich enzyme linked immunosorbent assay (ELISA) developed by R & D system in a blind fashion (Quantikine, R & D system, Minneapolis, MN). The antibodies used in HGF and VEGF determination do not cross-react with other cytokines. The limits of detection were 40.0 pg/mL for HGF and 9.0 pg/mL for VEGF. Both the intra-assay and inter-assay coefficients of variation were <10% for all these assays.

**Reverse transcription-polymerase chain reaction (RT-RCR).** Ribonucleic acid (RNA) was extracted from each of $10^6 M \phi$ and stromal cells cultured in 60 mm petridish (Greiner) using the monophasic solution of 40% phenol and ISOGEN method (Molecular Research Center, Tokyo), according to the manufacturer’s protocol.

The presence of mRNA encoding the estrogen and progesterone receptors in basal M \( \phi \) was determined using forward and reverse primers synthesized to anneal with cDNA for respective receptors. Amplification of cDNA reaction mixture for ER and PR was done in two stages and as described previously (McLaren et al., 1996). The mRNA expressions of HGF and its receptor c-Met were also analyzed in isolated peritoneal fluid M \( \phi \) and endometrial stromal cells by RT-PCR and using sense and antisense primers of
HGF and c-Met as previously described (Jiang et al., 1999, Khan et al., 2005). PCR generated bands were cloned and found to match the published sequences for the expected products. Human oligonucleotide primers of ER, PR, HGF, c-Met, and β-actin which we used for our current study are shown in Table 1. A scanner densitometer was used to determine the ratio of intensity of each band relative to β-actin which was used as an internal control.

Because quantitative application of this method is contingent upon the analysis of the PCR products during the amplification phase before the plateau, cycle relationships and dilution curves for cDNA of each target molecule and the housekeeping gene β-actin were determined.

**Cell proliferation assay.** Bromodeoxyuridine (BrdU) labeling and detection kit measures cell proliferation by quantitating BrdU incorporated into the newly synthesized DNA of replicating cells. The incorporated BrdU can be detected by a quantitative cellular enzyme immunoassay (Biotrak, Amersham Pharmacia Biotech Ltd., UK) using monoclonal antibodies directed against BrdU. It offers a non-radioactive alternative to the $[^3]H$-thymidine-based cell proliferation and carries equal sensitivity and specificity
Briefly, desired cells (endometrial stroma cells and Mφ) were cultured in 96 well microtitre plate (10^4 cell/well). The cells were phenol red starved with 5% FBS before HGF (50 ng/mL) and E2 (10^{-8}M) treatment for a period of 24 hour. After a 24hr pre-incubation period, respective cells were treated with HGF and E2 either alone or in combination in a serum and phenol red free media and incubated for an additional 24 hours. The BrdU incorporation was assayed according to the manufacturer’s instruction.

In order to confirm that growth promoting factor in the Mφ conditioned media is HGF, we used antibody to deplete HGF in the conditioned medium. The Mφ conditioned media was added to the cultured endometrial stromal cells and macrophages, and then incubated for 24 hr. HGF inhibition studies using a specific anti-human HGF neutralizing antibody (10 μg/mL, R & D system) were carried out in parallel, incubated for another 24 hr and BrdU incorporation was measured to examine the change in cell proliferation.

**Statistical analysis.** The results were evaluated by one-way analysis of variance. The data are expressed as mean ± SEM. The concentrations of the studied
cytokines were not distributed normally and the data were analyzed using non-parametric test. The differences between two groups were compared using Mann-Whitney U-test or Student’s t test. For comparison among three or more groups, the Kruskal-Wallis test was used to determine the difference among the groups. Differences were considered as statistically significant for p<0.05.

Results

We measured CD68 immunoreactive Mφ concentrations in the PF of women with endometriosis and they were distributed according to revised-ASRM staging, color appearances of endometriotic lesions and also by the different phases of menstrual cycle. Peritoneal fluid of women with early endometriosis (stage I-II) and those containing red lesions in pelvic cavity harbored abundant macrophages in PF when compared with that of control women (p<0.05), advanced endometriosis (stage III-IV, p<0.5) or other pigmented lesions or chocolate cysts (<0.05) (Fig. 1A and 1B). Although no menstrual phase difference was observed in control women, a dominant infiltration of Mφ in PF was found in the secretory or menstrual phase in women with endometriosis (p<0.05 and
p<0.01, respectively, Fig. 1C).

**Expressions of estrogen and progesterone receptors by Mϕ.** The majority of CD68 immunoreactive Mϕ as isolated from women with or without endometriosis showed strong nuclear staining for ER but were less reactive to PR (Fig. 2A). In contrast, tissue localization of ER and PR was equally demonstrated in the same position of CD68 immunoreactive Mϕ (Fig. 2B) (shown by corresponding arrow heads) in the serial section of intact tissues derived from the eutopic endometrium of woman with endometriosis. This indicates that besides glandular epithelium and stroma, ER and PR are also being synthesized and expressed by the infiltrated Mϕ in intact tissue.

This was further confirmed by RT-PCR and revealed that basal Mϕ isolated from the PF of women with or without endometriosis contained the mRNA encoding for ER and PR (Fig. 3A). No phase of the menstrual cycle dependent variation in the expression of these receptor mRNAs was evident for either group.

**Production of HGF by peritoneal fluid Mϕ in response to ovarian steroids.** Ovarian steroid concentrations are elevated in peritoneal fluid (Khan et al., 2002b; DeLeon et al., 1986) and data exist to show that these steroids can regulate the
secretion of a number of factors from animal and human peritoneal fluid macrophages (Hu et al., 1988; Frazier-Jassen et al., 1995; Chao et al., 1995). Therefore, we tried to investigate the secretion of HGF and other macromolecules by PF macrophages in response to ovarian steroids. We found that direct stimulation of Mφ in culture with E2 and P resulted in a variable increase in the secretion of HGF and VEGF by peritoneal fluid Mφ (Fig. 4). The production of HGF was significantly increased by E2 in women with endometriosis than that of control women (352.2 ± 4.9 pg/mL vs. 170.6 ± 2.6 pg/mL, respectively, p<0.05) or non-treated macrophages (221.5 ± 32.8 pg/mL, p<0.05). A marked increase in the secretion of VEGF was observed by treatment with E2 and P (p<0.01 for both) in women with endometriosis and also in women without endometriosis (p<0.05 for both) when compared with non-treated PF macrophages (Fig. 4). The production of HGF was about one half than that of VEGF in women with or without endometriosis. There was no statistical difference in response between E2 and P or a combination of E2 and P. No phase of cycle differences were seen.

Since progesterone failed to show a significant increase in the secretion of HGF by PF macrophage, we performed a blocking experiment on E2 by using ER antagonist,
tamoxifen. It was interesting to observe that tamoxifen significantly reversed the secretion of VEGF (p<0.05) and tended to reverse the secretion of HGF (p=0.07) by the estrogen-treated PF macrophages towards the non-treated macrophages (Fig. 4). This indicates that it is the direct effect of estrogen on the PF macrophages that was able to produce significant amount of HGF and VEGF and is being mediated by ER as located on these inflammatory cells.

**Steroidal stimulations of HGF and VEGF production by LPS-activated peritoneal fluid Mφ.** To investigate the control of HGF and VEGF secretion by PF macrophages, we measured HGF and VEGF secretion by activated (LPS, 5 ng/mL) and non-activated PF macrophages, isolated from women without endometriosis. Macrophages from women with endometriosis were not used in order to eliminate any bias in the basal activation status of these cells as already observed by our previous study (Khan et al., 2005). HGF and VEGF were secreted from both activated and non-activated PF macrophages. However, activation with LPS significantly increased the amount of HGF (p<0.05) and VEGF (p<0.01) secreted by these cells (Fig. 5). We observed that activation of basal macrophages further enhanced the response of these cells to ovarian
steroids. In fact, exogenous treatment with E2 was able to further increase the amount of both HGF (p<0.05) and VEGF (p<0.05) secretion by PF macrophages when these cells were activated with LPS (Fig. 5). Although progesterone increased the secretion of VEGF by non-activated MΦ, it was unable to further enhance the secretion of either HGF or VEGF by activated PF macrophages (Fig. 5). These results confirmed that irrespective of activation status, PF macrophages were independently stimulated to produce HGF and VEGF by E2. This also indicates that an inflammatory response and ovarian steroid hormones may function either alone or in combination to regulate the production of HGF and VEGF by PF macrophages in pelvic microenvironment.

**mRNA expression of HGF and c-Met in PF macrophages and endometrial stroma.** Since VEGF mRNA expression has already been demonstrated previously in isolated PF macrophages (McLaren et al., 1996), we tried to examine the transcriptional activity of HGF and its receptor, c-Met from the isolated PF macrophages and endometrial stroma in response to ovarian steroids (Fig. 3B and 3C). In a time-dependent study, HGF response to E2 (10^{-8}M) was delayed, with an increasing level of HGF mRNA
observed from 6 hr to 24 hr after the addition of E2 in both PF macrophages and stroma (data not shown).

As shown in Figure 3B, peritoneal fluid macrophages derived from women with endometriosis and in response to E2 (10^{-8}M) displayed higher expression of HGF mRNA (3.2 to 3.8 fold) than that of women without endometriosis (1.2 to 1.6 fold) (p<0.05). No difference in c-Met mRNA expression was observed in both Mφ and stromal cells between the normal and endometriosis group (Fig. 3B and 3C). Similar to PF macrophages, an increase in the expression of HGF mRNA was observed in the stromal cells derived from women with endometriosis than that of women without endometriosis (Fig. 3C). Again no differences in the expression of HGF and c-Met mRNA were seen between proliferative and secretory phases of menstrual cycle in these two groups of women (Fig. 3B and 3C). This indicates an autocrine or paracrine regulation of HGF by PF macrophages and endometrial or endometriotic stromal cells in response to ovarian steroids which may result in the elevation of HGF in the peritoneal fluid of women with endometriosis as we reported recently (Khan et al., 2002b, 2004b).

**HGF-dependent increases in stromal cell and macrophage proliferation.**
The presence of c-Met receptor in endometrial stromal cells and Mφ might be expected to enable these cells to respond to endogenous or exogenous HGF. The effect of media conditioned by peritoneal Mφ on non-endometriotic stromal cell and Mφ proliferation was determined by BrdU incorporation assay. Neutralizing experiments using a specific anti-HGF antibody were carried out in parallel.

The culture media as conditioned by the PF macrophages of women with endometriosis resulted in a significantly greater (p<0.05 for both stroma and Mφ) incorporation of BrdU than that of media conditioned by Mφ derived from non-endometriosis women (Fig. 6A and 6B). This additional incorporation was significantly decreased by the inclusion of anti-HGF antibody to the culture medium (p<0.05 for both stroma and Mφ) that is comparable to those seen with medium conditioned by Mφ being collected from women without endometriosis (Fig. 6A and 6B). This indicates that the proliferation of stromal cells and macrophages induced by the conditioned media of endometriosis may be contributed by HGF.

We finally tried to examine the effect of exogenous HGF and E2 on the proliferation of endometrial stromal cells and Mφ. We found that stromal cells derived
from women with endometriosis significantly incorporated BrdU in response to recombinant HGF (50 ng/mL) and E2 ($10^{-8}$M) either alone or in combination when compared with BrdU incorporation of non-treated stromal cells (p<0.05 for all, Fig. 7A). Only a combination of HGF and E2 treatment was able to significantly proliferate stromal cells of control women (p<0.05). A similar pattern of BrdU incorporation was seen in stromal cells derived from ectopic endometrium (data not shown). A similar significantly greater BrdU incorporation was observed in PF macrophages derived from women with endometriosis than that of non-treated Mφ (p<0.05 for all, Fig. 7B). Besides combined treatment of HGF and E2, macrophages were also responsive to significantly incorporate BrdU after single treatment of HGF in control women (p<0.05, for both).

Since BrdU incorporation study represents the simple incorporation of BrdU into the proliferated DNA of these cells and does not reflect the actual cell growth as accounted by increased cell number, therefore, we also examined the cell growth of stroma and Mφ by cell number (initial plating $10^5$ cells/well) under HGF and E2 stimulation. We found a parallel and significantly increased cell growth under the similar stimulation for both stromal cells and macrophages (data not shown).
Discussion

We demonstrated for the first time the production of HGF by the peritoneal Mφ in response to ovarian steroids in women with or without endometriosis. Although macrophage infiltration in PF was dominant in the secretory or menstrual phase, we found a little relation of HGF production by E2 or ER and PR expression with the phase of menstrual cycle. Our results of HGF production in response to E2 indicate that HGF production is possibly related to the activation status of macrophages in peritoneal milieu. In fact, the activation status of macrophages retains increased potentiality to produce and secrete different macromolecules in women endometriosis than that of non-endometriosis.

Our current findings are in consistent with the increased production of other cytokines and growth factors by the activated macrophages as described previously from our laboratory and by other literatures (Khan et al., 2002b, 2004b; Halme et al., 1988, 1989).

It is generally believed that ovarian steroid hormones are essential for the growth or persistence of ectopic endometrium and corresponding eutopic endometrium in women with endometriosis (Bergqvist, 1992). To date it was assumed that this was a consequence of direct actions of the steroids on the endometrial or endometriotic tissues.
Indirect actions of ovarian steroids on the inflammatory cells within the peritoneal fluid (PF) have generally been overlooked or scarcely described. Since the major cellular constituents of PF are macrophages, comprising between 82 and 99% of the total cell population (Eischen et al., 1994), it is quite reasonable to speculate that these cells may be responsive to ovarian steroids. Our current study is a further piece of evidence to describe this interaction between macrophages and ovarian steroid hormones.

We demonstrated that these inflammatory cells retain the mRNAs encoding both ER and PR and showed nuclear staining for both ER and PR in isolated M\(\phi\) and in intact tissue of endometriosis. These results are in accordance with the recently published results of McLaren et al. (1996). Our findings of increased concentrations of M\(\phi\) in the PF of women with early endometriosis and those containing active blood-filled red lesions in pelvic cavity are in parallel with the increased tissue infiltrations of M\(\phi\) in women with endometriosis as we reported recently (Khan et al., 2004a).

In this study we have demonstrated that HGF can be produced by PF macrophages derived from pelvic cavity in addition to its production by alveolar macrophages or hepatic kupffer cells (Skr tic et al., 1999; Crestani et al., 2002; Morimoto
et al., 2001). We reported that PF macrophages could be a constant source of different macromolecules including HGF and VEGF in response to ovarian steroids. We found that PF macrophages were directly stimulated to secrete HGF and VEGF by ovarian steroids, a response which was blocked by ER receptor antagonist and which was enhanced if the cells have been previously activated with LPS.

Besides ER and PR expression, PF macrophages as well as endometrial stromal cells expressed both HGF and its receptor, c-Met transcripts which were independent of menstrual cycle and the expression of HGF at both mRNA and protein levels were more higher in women with endometriosis than that of non-endometriosis. This indicates that expressions of HGF ligand-receptor by PF macrophages may depend on the activation status of these inflammatory cells. Finally, we demonstrated that the enhanced proliferation of endometrial stromal cells and PF macrophages induced by conditioned media of macrophages was abolished by anti-HGF antibody. This mitogenic effect of endogenous HGF was in parallel with the exogenous stimulation of HGF on these cells either alone or synergistic with estrogen.

These results have two-folds of biological implications, one, ovarian steroids
may influence the autocrine regulation of macrophages or stormal cell functions; two, an inflammatory response in pelvic environment and ovarian steroid hormones may function independently or in an orchestrated manner which may be involved in the growth or persistence of endometriosis. The production of HGF by the cells of mesenchymal origin and its interaction with c-Met receptor on epithelial cells, endothelial cells or mesothelial cells has been generally accepted (Nakamura et al., 1986; Tajima et al., 1992). We described here that endometrial stromal cells and infiltrated Mφ also retain c-Met receptor and the proliferation of these cells in response to exogenous HGF strengthened the notion that besides paracrine mode of action, HGF may also play an autocrine mode of action in the growth of endometriosis.

Although PF macrophages are the principal source of VEGF production in pelvic microenvironment (McLaren et al., 1996), it could also be produced by isolated endometrial cells (Shifren et al., 1996). These results are in agreement with our current results of VEGF production by PF macrophages. In fact, we found that the ovarian steroid-stimulated production of VEGF by PF macrophages was two-fold higher than that of HGF production by these cells. In addition to the main source of product by
mesenchymal cells, HGF may also be produced by peritoneal macrophages.

Besides a significant production of VEGF by both estrogen and progesterone, a higher production of HGF was demonstrated by estrogen only both at the transcriptional and protein levels. This indicates a variable response of PF macrophages to ovarian steroids in the production of HGF and VEGF. This can be explained by a differential interaction of ovarian steroid hormones with the estrogen (ERE) and progesterone response elements (PRE) located on the promoter region of HGF and VEGF. In fact, several half-palindromic consensus sequences for ERE and PRE were found on the promoter region of VEGF gene and claimed that VEGF may be the primary response gene for reproductive steroids in the endometrium (Shifren et al., 1996). In contrast, HGF gene retains only two putative ERE in its 3-kb 5’-promoter region in addition to having response elements for other cytokines and growth factors (Zarnegar, 1995). The existence of PRE on the promoter of HGF gene is now unknown. Therefore, the regulation of HGF gene by estrogen may be mediated by a direct interaction of the estrogen receptor complex with \textit{cis}-acting ERE elements.

We previously demonstrated that like VEGF, HGF also carries angiogenic and
mitogenic activity in endometrial tissues (Khan et al., 2003). Since HGF production and mRNA expressions of its ligand-receptor are up-regulated by E2-stimulated PF macrophages, when we extended our experiment to investigate the role of HGF in the pathogenesis of endometriosis, we found that the proliferation of both isolated stromal cells and PF macrophages derived from women with endometriosis were more enhanced by HGF either alone or in combination with estrogen.

Although not measured in our current study, we already demonstrated that the concentrations of HGF and other inflammatory mediators in PF were significantly elevated in women with early endometriosis and in those harboring blood-filled opaque peritoneal lesions comparing to that of women without endometriosis or non-opaque lesions (Khan et al., 2002b, 2004b). These results of a persistent inflammatory response in women with endometriosis and estrogen-regulated production of HGF by activated and non-activated PF macrophages further confirmed that the growth of endometriosis possibly depends on a mutual interaction between innate immune system and ovarian steroid hormones in pelvic microenvironment. The current therapeutic strategy of hypo-estrogenic medication in women with endometriosis can also be explained by its
effect on innate immune system, which may suppress different cytokines and growth factors and thereby improve the growth of endometriosis or other endocrine diseases. Further studies are required to evaluate the effect of hypo-estrogenic medication on immune cells.

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**Figure Legends**

**Figure 1.** Shows the peritoneal fluid concentrations of macrophages derived from women with or without endometriosis and are distributed according to the revised-ASRM staging (A), morphologic appearance with chocolate cysts (B) and different phases of menstrual cycle (C). The results are expressed as mean ± SEM. A, *p<0.05, stage I-II vs. stage III-IV or without endometriosis. B, *p<0.05, women containing red lesions vs. women with other lesions/chocolate cysts or without endometriosis. C, *p<0.05, secretory phase vs. proliferative phase; **p<0.01, menstrual phase vs. secretory phase or proliferative phase.

**Figure 2.** Shows the immunohistochemical staining of estrogen receptor (ER) and progesterone receptor (PR) in isolated peritoneal fluid Mφ (A) and in the serial sections of intact tissue (B). The isolated PF macrophages show strong nuclear staining for ER and weak staining for PR. The ER and PR expressions are shown in the same
position of CD68 immunoreactive Mφ and were demonstrated in the stromal component of eutopic endometrium derived from woman with endometriosis. The arrow heads show the co-localization of ER and PR at the same position of CD68 stained Mφ. Final magnification was adjusted at x50 and using light microscope connected to a camera (Olympus-VANOX, model-AHBS, Tokyo, Japan).

**Figure 3.** Expression of mRNA encoding for estrogen receptor, progesterone receptor, hepatocyte growth factor (HGF) and its receptor, c-Met in isolated peritoneal fluid (PF) macrophages and endometrial stromal cells as assayed by RT-PCR. Ethidium-stained agarose gels show representative products amplified from cDNA derived from PF macrophages or stromal cells. (A) Estrogen and progesterone receptors; endo (+), women with endometriosis; endo (-), women without endometriosis; P, proliferative phase; S, secretory phase; c-DNA (-), negative control with no cDNA, M, marker for DNA. (B and C) Effect of estradiol (10^{-8}M) on the mRNA expression of HGF and c-Met in isolated PF macrophages (B) and in endometrial stromal cells (C) derived from women with or without endometriosis. The mRNA expression of HGF was found to be significantly higher in women with endometriosis than that of non-endometriosis. No
differences in the expression of c-Met mRNA or in the phases of menstrual cycle were seen between these two groups of women. \( \beta \)-actin, internal control. Calculated molecular weights of product bands are indicated by arrows.

**Figure 4.** Shows the production of hepatocyte growth factor (HGF) and vascular endothelial cell growth factor (VEGF) in the conditioned media of peritoneal fluid macrophages (Mφ) derived from women with or without endometriosis and in response to estradiol (E2), progesterone (P), a combination of E2 and P (E2+P), and an estrogen receptor (ER) antagonist, tamoxifen (TMX). The results are expressed as mean ± SEM of six patients from each group (three proliferative and three secretory phase samples). HGF, *p<0.05 vs. non-treated Mφ or control women; VEGF, *p<0.05 vs. non-treated Mφ, **p<0.01 vs. non-treated Mφ or control women. The blocking of ER by tamoxifen tended to reverse the production of HGF and VEGF towards non-treated Mφ. VEGF, #p<0.05 vs. E2-treated Mφ.

**Figure 5.** Shows the production of hepatocyte growth factor (HGF) and vascular endothelial cell growth factor (VEGF) in the conditioned media of activated and/or estradiol (E2)- and progesterone (P)-treated macrophages (Mφ) that were derived
from six women with non-endometriosis (three proliferative and three secretory phase samples). Isolated peritoneal fluid Mφ, untreated or activated by lipopolysaccharide (LPS, 5 ng/mL), were incubated with E2 or P for 24 hr, and the resultant conditioned media were assayed for HGF and VEGF. The results are expressed as mean ± SEM of six patients. HGF, *p<0.05 vs. LPS non-treated Mφ; ¶p<0.05 vs. LPS-treated Mφ.

VEGF, *p<0.01 vs. LPS non-treated Mφ; ¶p<0.05 vs. LPS-treated Mφ; #p<0.05 vs. LPS non-treated Mφ.

**Figure 6.** Shows the effect of conditioned media of peritoneal fluid macrophages (Mφ) of women with or without endometriosis on the proliferation of non-endometriotic stromal cells (A) and Mφ (B) and was measured by bromodeoxyuridine (BrdU) incorporations in these cells. Neutralization studies were carried out using anti-human HGF neutralizing antibody. The results are expressed as percentage of control (mean ± SEM). Non-macrophage conditioned media equals 100%. Six patients were examined from each group (three proliferative and three secretory phase). *p<0.05, endometriosis vs. non-endometriosis for both stromal cells and Mφ; #p<0.05, anti-HGF antibody vs. without anti-HGF antibody for both stromal cells and M
\( \phi \). Endo (-) means women without endometriosis, Endo (+) means women with endometriosis.

**Figure 7.** Shows the exogenous single or combined effect of hepatocyte growth factor (HGF) and estradiol (E2) on the proliferation of endometrial stromal cells (A) and peritoneal fluid macrophages (B) derived from six women with endometriosis and six women without endometriosis. Results of bromodeoxyuridine (BrdU) incorporations are expressed as percentage of control (mean ± SEM). BrdU incorporations of non-treated cells equals 100%. Stroma of endometriosis, **\( p < 0.05 \) vs. non-treated cells; stroma of non-endometriosis, *\( p < 0.05 \) vs. non-treated cells. M\( \phi \) of endometriosis, **\( p < 0.05 \) vs. non-treated M\( \phi \); M\( \phi \) of non-endometriosis, *\( p < 0.05 \) vs. non-treated M\( \phi \).
Figure 4

The figure shows bar graphs comparing the concentration of HGF and VEGF in culture media under different conditions. The treatments include no treatment, E2 (10^{-8} M), P (10^{-6} M), E2 + P, and TMX (10^{-6} M). The graphs are divided into two sections: one for Endo (-) and one for Endo (+). The concentration is measured in pg/mL. Significant differences are indicated with asterisks (*) and double asterisks (**) for Endo (-) and (#) for Endo (+).
Figure 5

Graphs showing the concentration of HGF and VEGF in culture media under different conditions:

- **HGF**
  - LPS (-) compared to LPS (5 ng/mL)
  - Bars for E2/P untreated, E2 treated (10^-8 M), and P treated (10^-6 M)
  - Significant differences marked with symbols

- **VEGF**
  - Bars for E2/P untreated, E2 treated (10^-8 M), and P treated (10^-6 M)
  - Significant differences marked with symbols

Conc. in culture media (pg/mL)
Figure 6

A. stroma

B. Mφ

BrdU incorporation (% of control)

Mφ conditioned media
+ anti-HGF antibody

Endo (-)  Endo (+)

*  *

#  #
Table 1. Human oligonucleotide primers of ER, PR, HGF, c-Met and β-actin used.

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<th>Locus</th>
<th>Sense (5’-3’)</th>
<th>Antisense (5’-3’)</th>
<th>Size (bp)</th>
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<td>Primer 3: CTTTGGCAAAGCCCGCTC</td>
<td>Primer 4: TCATCATGCGGAACCCGAGCT</td>
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<tr>
<td>PR</td>
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<td>Primer 2: GACTTTCGTAGCCCCCTCCA</td>
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<tr>
<td></td>
<td>Primer 3: GGAAGGGCACAGCAACAATA</td>
<td>Primer 4: AAGGAATTGTATTAAGAAGTAA</td>
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<tr>
<td>HGF</td>
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<td>β-actin</td>
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ER, estrogen receptor; PR, progesterone receptor; HGF, hepatocyte growth factor; c-Met, receptor for HGF; β-actin, internal control.