Ontogenetic phase shifts in metabolism in a flounder *Paralichthys olivaceus*

Mitsuharu Yagi & Shin Oikawa

1 Faculty of Fisheries, Nagasaki University, Bunkyo, Nagasaki 852-8521, Japan, 2 Fishery Research Laboratory, Graduate School of Bioresource and Bioenvironmental Sciences, Kyushu University, Fukuoka 811-3304, Japan.

Size-scaling metabolism is widely considered to be of significant importance in biology and ecology. Thus, allometric relationships between metabolic rate ($V_{O2}$) and body mass ($M$), $V_{O2} = aM^b$, have long been a topic of interest and speculation. It has been proposed that intraspecifically metabolic rate scales isometrically or near isometrically with body mass during the early life history in fishes, invertebrates, birds and mammals. We developed a new perspective on intraspecific size-scaling metabolism through determination of metabolic rate in the Japanese flounder, *Paralichthys olivaceus*, during their early life stages spanning approximately four orders of magnitude in body mass. With the increase of body mass, the Japanese flounder had four distinct negative allometric phases in which three stepwise increases in scaling constants ($a_i; i=1–4$), i.e. ontogenetic phase shifts in metabolism, occurred with growth during its early life stages at around 0.002, 0.01 and 0.2 g, maintaining each scaling exponent constant in each phase ($b=0.831$). These shifts in metabolism during the early life stages are similar to the tiger puffer, *Takifugu rubripes*. Our results indicate that ontogenetic phase shifts in metabolism are key to understanding intraspecific size-scaling metabolism in fishes.

Metabolic rate of living things is related to many biological traits such as body size, growth, phylogeny of species, rates of reproduction and genome evolution. Body size of animals primarily constrains the metabolic rate through the size-scaling effect and these two traits have co-evolved. It has been established that the relationship between metabolic rate ($V_{O2}$) and body mass ($M$) in animals is expressed by the allometric formula,

$$V_{O2} = aM^b,$$

where "$a$" is a scaling constant and "$b$" is the scaling exponent. However there are many cases where a simple allometric equation can not explain non-linear or curvilinear log-log metabolic scaling relationships. Furthermore intraspecific size-scaling metabolism of animals may be a complex process in which several phases can be distinguished.

Fishes are good candidates for intraspecific studies because, unlike other vertebrates such as mammals, birds or reptiles, most fishes develop from a small body size (mg level or less) and attain a mass range approximately comparable to that between a mouse and an elephant. In addition, it is not necessary to consider the phylogeny of species. Although reported scaling exponents in various species of fish are quite variable (approximately 0.4 to 1.3), a scaling exponent has often been suggested as close to unity in the early stages of development of marine fish larvae. On the other hand, in juveniles and adults, the relationship between metabolic rate and body mass is negatively allometric, and the mass exponents for simple regression of metabolic rate on body mass are usually less than 1. Consequently, a biphasic relationship where the scaling exponent changes from isometry or near isometry during the larval phase to negative allometry during the juvenile phase has been proposed in many fish species. Although these studies concluded that both the metabolic scaling exponent and scaling constant varies during ontogeny, the generality of the isometric scaling remains an open question requiring additional empirical evidence.

Recently we have demonstrated that in the larval and juvenile tiger puffer, *Takifugu rubripes*, oxygen consumption ($V_{O2}$), as a proxy for metabolism, scales with body mass as $V_{O2} = aM^b$, and that three stepwise increases in scaling constants $a_i; (i=1–4)$, i.e. ontogenetic phase shifts in metabolism, occur, maintaining each scaling exponent constant in each phase. The overall scaling exponent (0.948 ± 0.032) (estimate ± s.e.m.) was steeper than the scaling exponents for each developmental stage (0.795 ± 0.019). This result suggests that isometric or near-isometric metabolic scaling during early life stages may be produced by a combined effect of ontogenetic phase shifts.
shifts in metabolism. Thus, ontogenetic shifts in metabolism are likely to occur in other teleost fish species. In the present paper, we examine if the ontogenetic phase shifts in metabolism occur with growth in the Japanese flounder Paralichthys olivaceus (Temminck & Schlegel, 1846).

**Results**
Rates of oxygen consumption ($V_{O_2}$ in $\mu$O$_2$ fish$^{-1}$ min$^{-1}$) in relation to body mass ($M$ in g) are plotted in Figure 1. Mass-specific rates of oxygen consumption ($V_{O_2}/M$ in $\mu$O$_2$ g$^{-1}$ min$^{-1}$) are also presented. $V_{O_2}$ increased daily from just after hatching to 7 days after hatching (DAH), with virtually no increase of body mass. After 7 DAH, body mass increased. There was no substantial difference between the results of the two different methods of respirometry (Figure 1). Therefore, the values obtained were used without any distinction for the relationship between oxygen consumption and body mass.

Two models were applied to compare the four negative allometric relationships:

\[ V_{O_2} = a_i M^b, \]  
\[ V_{O_2} = a_i M^b, \]

for each incidence of negative allometry, and

\[ V_{O_2} = z M^b \]

for the overall line constituting these allometries. $a_i$ represents an intragroup scaling constant of the $i$th group, and $z$ the intergroup one of the groups. Equations (2) and (3) were rewritten as

\[ y_{ij} = \log_{10} a_i + b x_{ij} + e_{ij}, \]

and

\[ y_{ij} = \log_{10} V_{O_2} + e_{ij}, \]

where $y_{ij}$ is $\log_{10} V_{O_2}$, $x_{ij}$ is $\log_{10} M$ and $e_{ij}$ and $E_i$ represent the random intra- and intergroup variation in metabolism. To estimate $b$ and $\log_{10} z$, we used the ordinary least-squares regression to minimise the sum of squares of $\mu_i$, which is the vertical distance of the group mean $(\bar{y}_i, \bar{y}_i)$ from the overall line (intergroup line)\textsuperscript{5}. Because $\log_{10} a_i$ is equal to ($\log_{10} \mu_i + (b - b \bar{y}_i)$), Equation (4) was rewritten as follows\textsuperscript{3,36}:

\[ y_{ij} = \log_{10} V_{O_2} + \mu_i + (b - b \bar{y}_i) \bar{y}_i + b x_{ij} + e_{ij}. \]

The scaling exponents of individual lines were significantly smaller than unity for all regression lines ($P<0.05$; two tailed t-test). Regression analysis of each group, except the transitional phases is given in rows 1 to 4 of Table 1. The slopes of the individual lines in each of the four groups were not significantly different ($F_3, 94 = 1.95, P=0.127$; one-way ANCOVA). The intragroup scaling exponent $b$ was estimated to be $0.831 \pm 0.0026$ (estimate $\pm$ S.E.M.), and logarithm of the scaling constants in each group at $b = 0.831$ was estimated as follows: $\log_{10} a_1 = 0.373 \pm 0.083$, $\log_{10} a_2 = 0.425 \pm 0.062$, $\log_{10} a_3 = 0.539 \pm 0.034$, and $\log_{10} a_4 = 0.584 \pm 0.018$. The scaling constants were calculated as follows: $a_1 = 2.36$, $a_2 = 2.66$, $a_3 = 3.46$, and $a_4 = 3.84$. Regression analysis of the intergroup is given in rows 5–7 of Table 1. The intergroup scaling exponents $b$ was estimated to be $0.915 \pm 0.0011$.

A one-way ANCOVA was carried out to clarify the validity of Equation (6), and the results are provided in Table 2. The $\mu_i$‘s were not different from zero ($F_3, 97 = 1.71, P=0.186$; one-way ANCOVA), meaning that the intragroup means might lie on the intergroup means.

Figure 1 | Ontogenetic changes in the rate of respiration ($V_{O_2}$, diamonds) and the mass-specific rate of respiration ($V_{O_2}/M$, circles) with increase of body mass ($M$) in Japanese flounder. Symbols for $V_{O_2}$ indicate the two methods of respirometry (open: closed method and solid: semi-closed method). Symbols for $V_{O_2}/M$ signify the year in which data were collected (open: 2005 and solid: 2006). The vertical broken lines at around 0.0003 g represent $V_{O_2}$ and $V_{O_2}/M$ which increased daily from just after hatching to 7 days after hatching, with virtually no increase in body mass. Ranges covered by the four solid lines each for $V_{O_2}$ and $V_{O_2}/M$ indicate intragroup phases of negative allometry. The broken lines both on $V_{O_2}$ and $V_{O_2}/M$ represent the intergroup lines. Small symbols represent values during the transitional phases. Regression analysis of each line for $V_{O_2}$ is presented in Table 1, and ANCOVA in Table 2.
what was observed in the previous study: scaling constants for the Japanese flounder to the tiger puffer ones, the transitional phases.

constant in each phase (growth during its early life stages, maintaining each scaling exponent constant in each phase (Figure 1). Thus, the metabolic rate is expressed by \( V_{O2} = aM^{0.83} \), in which \( a \) increased three times during the transitional phases.

Discussion

The size-scaling metabolism for the Japanese flounder during its early life stages (that is, spanning approximately four orders of magnitude in body mass) showed ontogenetic phase shifts, because the scaling constant \( a \) increased significantly from \( a_{1}=2.36 \) to \( a_{5}=3.84 \) with increase in body mass (Figure 1). Thus, the metabolic rate is expressed by \( V_{O2} = aM^{0.83} \), in which \( a \) increased three times during the transitional phases.

Table 1 | Intragroup (rows 1 to 4), intergroup (rows 5 to 7) regression analysis of the relationship between log10 \( V_{O2} \) (\( V_{O2} \): oxygen consumption in \( \mu L \ O_2 \) fish \(^{-1} \) min \(^{-1} \)) and log10 \( M \) (\( M \): body mass in g) in the Japanese flounder

<table>
<thead>
<tr>
<th>Group</th>
<th>( N )</th>
<th>Range of body mass (g)</th>
<th>Scaling (Constant)</th>
<th>Scaling exponent (Mean ± S.E.M.)</th>
<th>( P )</th>
<th>( R^2 )</th>
</tr>
</thead>
<tbody>
<tr>
<td>1(^*)</td>
<td>24</td>
<td>0.00041–0.0015</td>
<td>0.06</td>
<td>0.060 ± 0.065</td>
<td>6.26 \times 10^{-5}</td>
<td>0.832</td>
</tr>
<tr>
<td>2(^*)</td>
<td>26</td>
<td>0.0025–0.0099</td>
<td>2.57</td>
<td>0.825 ± 0.066</td>
<td>1.42 \times 10^{-2}</td>
<td>0.866</td>
</tr>
<tr>
<td>3(^*)</td>
<td>36</td>
<td>0.016–0.14</td>
<td>3.92</td>
<td>0.875 ± 0.035</td>
<td>1.12 \times 10^{-3}</td>
<td>0.948</td>
</tr>
<tr>
<td>4(^*)</td>
<td>16</td>
<td>0.24–0.90</td>
<td>3.67</td>
<td>0.782 ± 0.066</td>
<td>5.04 \times 10^{-3}</td>
<td>0.910</td>
</tr>
<tr>
<td>1–4(^*)</td>
<td>102</td>
<td>0.00041–0.90</td>
<td>( \beta = 0.915 ), ( \alpha = 0.0011 )</td>
<td>6.58 \times 10^{-9}</td>
<td>0.9999</td>
<td></td>
</tr>
<tr>
<td>1–4(^*)</td>
<td>102</td>
<td>0.0014–0.90</td>
<td>4.20</td>
<td>0.911 ± 0.0061</td>
<td>1.55 \times 10^{-26}</td>
<td>0.996</td>
</tr>
<tr>
<td>Total(^*)</td>
<td>124</td>
<td>0.00026–0.90</td>
<td>4.32</td>
<td>0.922 ± 0.0060</td>
<td>1.73 \times 10^{-24}</td>
<td>0.995</td>
</tr>
</tbody>
</table>

*Parameters were estimated to minimise the sum of squares of \( \varepsilon \), \( n \) is the number of determinations, \( P \) is the difference of scaling exponent from unity, examined using Students \( t \)-test, two tailed; \( R^2 \) is squared correlation coefficient between log10 \( V_{O2} \) and log10 \( M \).

Table 2 | ANCOVA table for respirometry in the Japanese flounder based on the model (equation (6))

<table>
<thead>
<tr>
<th>Term</th>
<th>Sum of squares</th>
<th>Degrees of freedom</th>
<th>Mean square</th>
<th>Mean-square ratio</th>
<th>( P )</th>
</tr>
</thead>
<tbody>
<tr>
<td>log ( \alpha )</td>
<td>110.12189</td>
<td>1</td>
<td>110.12189</td>
<td>34124</td>
<td>2.26 \times 10^{-125}</td>
</tr>
<tr>
<td>( \mu )</td>
<td>0.011030379</td>
<td>2</td>
<td>0.005515</td>
<td>1.71</td>
<td>0.186</td>
</tr>
<tr>
<td>( h = b - b ) ( \xi )</td>
<td>0.031216</td>
<td>1</td>
<td>0.031216</td>
<td>9.67</td>
<td>0.00246</td>
</tr>
<tr>
<td>( \xi = )</td>
<td>79.661712</td>
<td>97</td>
<td>0.003227</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total (about mean)</td>
<td>80.016990</td>
<td>101</td>
<td>24685</td>
<td>1.42 \times 10^{-118}</td>
<td></td>
</tr>
<tr>
<td>Total (about zero)</td>
<td>190.138887</td>
<td>102</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
and 0.2 g (6–10 and 25 mm in standard length; M. Yagi and S. Oikawa, unpublished data) during the transitional phases were accompanied by morphological and behavioural changes. At approximately 0.02 g larvae, rapid increment of swimming velocity was observed\(^2\). At ~0.01 g, the transformation from larval to juvenile stage occurs\(^2\). Fukuhara\(^3\) also reported that a relatively sharp increase in maximum swimming speed was observed for ~0.01 g juvenile fish. As larvae metamorphose into juveniles, and at ~0.1 g, squamation of the body proceeded\(^2\). For the Japanese flounder, at approximately ~0.02 g larvae, the ventral fin buds are present and the base of the dorsal and anal fins (primordial) appear, at ~0.01 g, development of fin rays is completed, and at ~0.2 g, squamation on the caudal peduncle is completed, consisting of eight scale rows\(^4\). Thus, in this study, three transitional phases of VT were fitted between V1, V2, and V3 based on the developmental stages which is similar to the tiger puffer Takifugu rubripes\(^5\), interposing three transitional phases of VT: at approximately 0.002, 0.01 and 0.2 g of wet body mass for the tiger puffer, at approximately ~0.002 g, the primordia of the fin rays, at ~0.01 g post-larvae, larvae metamorphosed into juveniles, and at ~0.1 g, squamation of the body proceeded\(^2\). Thus, in this study, three transitional phases were fitted between 0.002 (0.0015–0.0020 g), 0.01 (0.010–0.015 g) and 0.2 (0.15–0.22 g) g of wet body mass were interposed. Size-scaling analyses were performed by applying the statistical model described in Yagi et al.\(^6\).

**Methods**

**Fish used.** Fish (*P. olivaceus*) were hatched from artificially fertilized eggs obtained from wild parents in the northern part of Kyushu, Japan that were captured by fishermen. Experiments were conducted at the Fisheries Research Laboratory, Kyushu University, Fukuoka, Japan in 2005 and 2006. All larvae and juveniles were maintained in aerated 50 l tanks supplied with a constant flow of seawater and fed live rotifers, *Brachionus rotundiformis* twice daily between 2 and 22 DAH (~5000 1\(^{-1}\)), live brine shrimp, *Artemia* sp., twice daily two times between 20 and 32 DAH (~2000 1\(^{-1}\)), and artificial diets three times daily thereafter. Live diets were fortified with essential fatty acids, EPA and DHA using Super Capsule Powder (Chloroella Industry, Tokyo, Japan), before feeding. The water temperature in the rearing tanks was held at a 18 °C. This temperature was chosen to approximate conditions experienced by the early life stages of the flounder in the wild and was also used when measuring the oxygen consumption.). Fish used in the respirometry study were not fed for 3 to 24 h before experiments, depending on the fish body size, and so the individuals used for respirometry did not have food in their guts. All experimental procedures were approved by a KU committee and conducted in accordance with the Guideline for the Care and Use of Laboratory Animals of Japan, with registrations of M.Y. and S.O. for Animal experiments in the Faculty of Agriculture and in the Graduate Course of KU.

**Respirometry.** Oxygen consumption (*V\(_{O_2}\)) was measured in larval and juvenile fish ranging in size from 0.00026 g (wet body mass, 0 DAH) and 0.90 g (80 DAH). Resting routine rates of oxygen consumption, i.e. intermediate between the resting and routine activity states\(^6\), were determined in fasted larvae and juveniles at 18 °C using one of two methods depending on the developmental stage of the fish, a closed method and a semi-closed method, based on previous studies\(^6,7\). The larvae and juveniles were placed in the respiration chamber and allowed to settle for an appropriate acclimatization period before measurement to remove possible stress caused by handling (1 h for the closed method and 1 to 4 h for the semi-closed method). The closed method was only used for larvae just after hatching to 8 DAH, because of their poor swimming ability. The closed method was based on depletion of oxygen in water in a sealed glass oxygen bottle (20 ml). A blank chamber without fish was used to eliminate background respiration. The semi-closed method was essentially the closed method, but the glass chamber was slowly flushed with air-saturated water before determination and closed during determination. Several respiration chambers (50 to 650 ml) were used depending on the fish body size (50 ml for 8–25 DAH, 100 ml for 18–44 DAH, 250 ml for 46–63 DAH, 350 ml for 64–80 DAH, 650 ml for 63–80 DAH). In the semi-closed method, the bottle which received water flowing out of the respiration chamber was used as the blank chamber. This bottle was sealed at the beginning of determination of oxygen consumption by fish, placed in the water bath for the respiration chamber during determination, and the oxygen concentration in the bottle was determined at the end of the measurement. By using this value as the initial oxygen concentration in the respiration chamber, background respiration was cancelled.

Wet body mass of experimental fish was directly determined immediately after respirometry except for specimens used in the metabolic scaling study. In this study, wet body mass was indirectly estimated from other co-cultured individuals of similar body size, because they were fixed when the dissolved oxygen concentration of water in the respiration chamber was determined by the Winkler’s titration method.

**Data analysis.** Rates of oxygen consumption (*V\(_{O_2}\)) for the Japanese flounder were plotted on log\(_{10}\) plots against wet body mass (M), and four negative allometric regressions were fitted between *V\(_{O_2}\)* and *M* based on the developmental stages which is similar to the tiger puffer Takifugu rubripes\(^5\), interposing three transitional phases of *V\(_{O_2}\)*: at approximately 0.002, 0.01 and 0.2 g of wet body mass for the tiger puffer, at approximately ~0.002 g, the primordia of the fin rays, at ~0.01 g post-larvae, larvae metamorphosed into juveniles, and at ~0.1 g, squamation of the body proceeded\(^2\). For the Japanese flounder, at approximately ~0.02 g larvae, the ventral fin buds are present and the base of the dorsal and anal fins (primordial) appear, at ~0.01 g, development of fin rays is completed, and at ~0.2 g, squamation on the caudal peduncle is completed, consisting of eight scale rows\(^4\). Thus, in this study, three transitional phases were fitted between 0.002 (0.0015–0.0020 g), 0.01 (0.010–0.015 g) and 0.2 (0.15–0.22 g) g of wet body mass were interposed. Size-scaling analyses were performed by applying the statistical model described in Yagi et al.\(^6\).


### Acknowledgments

This work was partly supported by grants from the Sasakawa Science Research Foundation from The Japan Science Society to M.Y. [grant #23-749], and the Ministry of Education, Culture, Sports, Science and Technology of Japan to S.O. [grant #16580152]. We thank K. Taniguchi for assistance in rearing the fish.

### Author contributions

M.Y. and S.O. conceived of and designed the research. M.Y. cared for research animals. M.Y. and S.O. collected, processed and interpreted data. M.Y. drafted the manuscript. All authors read and commented on the manuscript.

### Additional information

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Yagi, M. & Oikawa, S. Ontogenetic phase shifts in metabolism in a flounder *Paralichthys olivaceus*. *Sci. Rep.* 4, 7135; DOI:10.1038/srep07135 (2014).

This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder in order to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by-nc-sa/4.0/