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Title

Serum Wisteria Floribunda Agglutinin-Positive Mac-2 Binding Protein Values Predict the Development of Hepatocellular Carcinoma among Patients with Chronic Hepatitis C after Sustained Virological Response

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RESEARCH ARTICLE

Serum Wisteria Floribunda Agglutinin-Positive Mac-2 Binding Protein Values Predict the Development of Hepatocellular Carcinoma among Patients with Chronic Hepatitis C after Sustained Virological Response

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Abstract

Measurement of Wisteria floribunda agglutinin-positive human Mac-2 binding protein (WFA*-M2BP) in serum was recently shown to be a noninvasive method to assess liver fibrosis. The aim of this study was to evaluate the utility of serum WFA*-M2BP values to predict the development of hepatocellular carcinoma (HCC) in patients who achieved a sustained virological response (SVR) by interferon treatment. For this purpose, we retrospectively analyzed 238 patients with SVR who were treated with interferon in our department. Serum WFA*-M2BP values were measured at pre-treatment (pre-Tx), post-treatment (24 weeks after completion of interferon; post-Tx), the time of HCC diagnosis, and the last clinical visit. Of 238 patients with SVR, HCC developed in 16 (6.8%) patients. The average follow-up period was 9.1 years. The cumulative incidence of HCC was 3.4% at 5 years and 7.5% at 10 years. The median pre-Tx and post-Tx WFA*-M2BP values were 1.69 (range: 0.28 to 12.04 cutoff index (COI)) and 0.80 (range: 0.17 to 5.29 COI), respectively. The WFA*-M2BP values decreased significantly after SVR (P < 0.001). The median post-Tx WFA*-M2BP value in patients who developed HCC was significantly higher than that in patients who did not (P < 0.01). Multivariate analysis disclosed that age (> 60 years), sex (male), pre-Tx platelet count (< 15.0×10^3/μL), and post-Tx WFA*-M2BP (> 2.0 COI) were associated with the development of HCC after SVR.
Conclusion
Post-Tx WFA*^M2BP (> 2.0 COI) is associated with the risk for development of HCC among patients with SVR. The WFA*^-M2BP values could be a new predictor for HCC after SVR.

Introduction
Hepatocellular carcinoma (HCC) is one of the most common malignant tumors in the world [1]. Chronic hepatitis C virus (HCV) infection is a major cause of HCC. Millions of people are persistently infected with HCV globally [2–4] and these individuals are at high risk of developing HCC [5–7]. Several studies have demonstrated that interferon (IFN) treatment in chronic hepatitis C patients reduces the risk for progression of liver disease, HCC, liver-related death, and all-cause mortality [8–13], especially in patients who exhibit a sustained virological response (SVR). However, some risk for HCC—albeit a small one—remains even after achieving viral eradication [10,14–19]. Several factors have been reported to affect HCC development among patients with SVR.

Recently, an assay for the measurement of Wisteria floribunda agglutinin-positive human Mac-2 binding protein (WFA*-M2BP) was reported as a novel, noninvasive, and rapid bedside method to assess liver fibrosis [20]. M2BP has been shown to have multibranching and sialylated N-glycans. WFA is considered to recognize the GalNAc residue of N-glycans and O-glycans or the clustered LacNAc (Gal-GlcNAc) structure. Currently, we are analyzing the glycan structures of WFA*-M2BP in detail using MS-based technology [21]. Glycans can reflect the differentiation stage of cells but not necessarily the level of cellular damage, and therefore they can be very effective markers for chronic disease. Several reports performed with proteome analysis have identified Mac-2 binding protein as a potential marker of liver fibrosis progression [22–25]. Kuno et al. were the first to report that a rapid and simple glycan-based immunoassay for WFA*-M2BP can quantify fibrosis [20,26]. On the other hand, we reported that AFP and WFA*-M2BP values are noninvasive predictive markers for the development of HCC in patients with HCV [27,28]. In this report we evaluated the utility of WFA*-M2BP values to predict the development of HCC in patients who had achieved SVR after IFN treatment.

Patients and Methods
Patients
From December 1989 to December 2010, a total of 601 consecutive HCV patients who received IFN treatment and achieved SVR at the National Hospital Organization Nagasaki Medical Center were enrolled in this retrospective study. The diagnosis of chronic HCV infection was based on continuous positivity for both anti-HCV by a second or third-generation enzyme-linked immunosorbent assay (ELISA) and positivity for serum HCV RNA by polymerase chain reaction (PCR). Before treatment, HCC was definitively ruled out either by ultrasonography (US), dynamic computed tomography (CT), or magnetic resonance imaging (MRI) on enrollment. Exclusion criteria for this study were: (1) positivity for hepatitis B surface antigen; (2) positivity for human immunodeficiency virus; (3) autoimmune hepatitis or primary biliary cirrhosis; (4) a shorter follow-up period (< 12 months) after the completion of IFN treatment; (5) a history of HCC at the time of IFN treatment; (6) development of HCC within 12 months after the completion of IFN treatment; (7) administration of low dose long-term IFN treatment; and (8) absence of properly stored serum samples or insufficient archival material. After
the exclusions, 238 patients who achieved SVR were analyzed retrospectively for the risk factors of HCC.

For all patients in our cohort, a blood sample was taken on the days of the administration of IFN treatment (pre-treatment; pre-Tx), 24 weeks after completion of IFN treatment (post-treatment; post-Tx), and on the days of HCC diagnosis and last clinical visit. All separated serum samples were stored at -20°C until use. Medical histories, along with the results of routine tests for blood cell counts, liver biochemistry and HCV viral load/genotype at the time of IFN treatment and thereafter, were retrieved from medical records. Complete blood cell counts and biochemical tests were performed using automated procedures in the clinical pathologic laboratories of our hospital.

**Histological evaluation**

Liver biopsies were undertaken using fine-needle aspiration (16G or 18G sonopsy) guided by US. Liver tissue specimens were fixed in 10% formalin, embedded in paraffin, and stained with hematoxylin and eosin. The histological assessment was made by two independent pathologists according to the classification of Desmet et al. [29].

**Interferon treatment**

Among the 238 patients, 123 received IFN monotherapy for 24 weeks, 28 patients received pegylated (PEG-)IFN monotherapy for 48 weeks, and 87 patients received IFN plus ribavirin or PEG-IFN plus ribavirin combination therapy for 48–72 weeks.

**HCV RNA and HCV genotypes**

The presence of HCV RNA was determined by reverse transcriptase (RT-) PCR using a commercial kit (Amplicor HCV; Roche Diagnostic Systems, Basel, Switzerland). Genotypes of HCV were determined by RT-PCR with genotype-specific primers (HCV RNA core genotype; Roche Diagnostics, Tokyo, Japan) [30,31]. In patients treated before the availability of PCR, the presence of HCV RNA was investigated by using sera stored at -20°C.

**Definitions of response to interferon treatment**

SVR was defined as the absence of detectable HCV RNA at 24 weeks after the end of IFN treatment. There was no relapse of viremia after 24 weeks among the patients who achieved SVR.

**Measurement of Wisteria floribunda agglutinin-positive human Mac-2 binding protein (WFA+-M2BP)**

WFA+-M2BP quantification was performed based on a lectin-antibody sandwich immunoassay using a fully automatic HISCL-2000i immunoanalyzer (Sysmex Co., Hyogo, Japan) [20]. The measured values of WFA+-M2BP conjugated to WFA were indexed with the values obtained using the following equation:

\[
\text{Cutoff index (COI)} = \left( \frac{[\text{WFA}^+\text{-M2BP}]_{\text{sample}} - [\text{WFA}^+\text{-M2BP}]_{\text{NC}}}{[\text{WFA}^+\text{-M2BP}]_{\text{PC}} - [\text{WFA}^+\text{-M2BP}]_{\text{NC}}} \right)
\]

Here, \([\text{WFA}^+\text{-M2BP}]_{\text{sample}}\) represents the WFA+-M2BP count of the serum sample (PC, positive control; NC, negative control). The positive control was supplied as a calibration solution preliminarily standardized to yield a COI value of 1.0 [26].
Follow-up and diagnosis of hepatocellular carcinoma

All patients were followed up at an interval of 1–12 months by measurement of blood count and liver biochemistry, along with quantitative detection of HCV RNA, AFP, AFP-L3, and DCP. Diagnostic imaging either by US, CT, or MRI was performed at least once per year. A diagnosis of HCC was made based on positive results of typical vascular patterns, as revealed by either contrast-enhanced CT, contrast-enhanced MRI or angiography. Otherwise, the pathological diagnosis was made by fine-needle biopsy of space-occupying lesions detected in the liver.

Ethical considerations

Informed consent to utilize medical records and specimens was obtained from each patient. We obtained the written consent of participants at the time of serum collection. These processes and the study protocol were approved by the Ethical Committee of National Hospital Organization Nagasaki Medical Center (confirmation number: 25102), and conformed with the 1975 Declaration of Helsinki and the Japanese Ethical Guidelines for Clinical Research (Ministry of Health, Labor, and Welfare of Japan, Ethical Guidelines for Clinical Research, 2008). Our research is available on the National Hospital Organization Nagasaki Medical Center website (http://www.nagasaki-mc.jp/).

Statistical analysis

Continuous variables (AST, ALT, albumin, total bilirubin, γ-GTP, fasting blood sugar, HbA1c, triglyceride, total cholesterol, BMI, platelet counts, AFP, WFA⁺-M2BP) were dichotomized with respect to the median value or clinically meaningful values in the multivariate analysis. Statistical analysis was performed using a Wilcoxon signed rank test and Mann-Whitney U-test. To estimate the cumulative risk of developing HCC, the Kaplan-Meier method and the log-rank test were used. Cox proportional hazards regression analysis was performed to evaluate risk factors for HCC. The diagnostic performances of WFA⁺-M2BP and AFP for censored development of HCC were assessed by examining the area under the time-dependent receiver operating characteristic (ROC) curves (AUROC) [32]. Inclusion of variables was assessed using a stepwise selection method. A P value of 0.05 was considered statistically significant. Data analysis was performed with SPSS ver. 22.0 (SPSS, Chicago, IL).

Results

Patient characteristics

The baseline characteristics of the 238 patients are summarized in Table 1. The median age was 55.0 years; 147 (61.8%) patients were male; and 104 (43.7%), 68 (28.6%), 42 (17.6%) and 24 (10.1%) patients were diagnosed histologically with fibrosis stage F1, F2, F3 and F4, respectively. The median value of the pre-Tx platelet count was 16.0 (range: 6.4 to 33.2 ×10^3/μL) and that of the post-Tx platelet count was 16.8 (range: 6.5 to 36.3 ×10^3/μL). The median value of pre-Tx AFP was 5 (range: 1 to 200 ng/mL) and that of post-Tx AFP was 3 (range: 1 to 46 ng/mL). The average follow-up period was 9.1 years.

Cumulative incidence of HCC

During the follow-up period, HCC developed in 16 (6.8%) of the 238 patients. The cumulative incidences of HCC at 5 and 10 years were 3.4% and 7.5%, respectively.
Risk factors for HCC

Univariate analysis demonstrated factors that increase the risk for HCC development after SVR. Cox regression analysis was performed on 20 variables: age, sex, BMI, alcohol intake, fibrosis stage, degree of steatosis, pre-Tx platelet counts, post-Tx platelet counts, albumin, pre-Tx AST, post-Tx AST, pre-Tx ALT, post-Tx ALT, $\gamma$-GTP, T.bilirubin, HbA1c, pre-Tx AFP, post-Tx AFP, pre-Tx WFA$^+$-M2BP, post-Tx WFA$^+$-M2BP. Cutoff values for AFP and WFA$^+$-M2BP were determined by time-dependent ROC analysis as 5 ng/ml and 2.0 COI, respectively.

Table 1. Characteristics of Patients Enrolled in the Present Study.

<table>
<thead>
<tr>
<th>Factors</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patients, n</td>
<td>238</td>
</tr>
<tr>
<td>Age, year (y)</td>
<td>55.0 (18–75)</td>
</tr>
<tr>
<td>Male, n (%)</td>
<td>147 (61.8)</td>
</tr>
<tr>
<td>BMI (kg/m$^2$)</td>
<td>23.20 (16.7–34.9)</td>
</tr>
<tr>
<td>Alcohol intake (&gt; 20g/day), n (%)</td>
<td>64 (29.9)</td>
</tr>
<tr>
<td>Fibrosis stage (≥ 1/2/3/4)</td>
<td>104 (43.7)/68 (28.6)/42 (17.6)/24 (10.1)</td>
</tr>
<tr>
<td>Steatosis (≥ 10%), n (%)</td>
<td>25 (10.5)</td>
</tr>
<tr>
<td>Pre-Tx platelet counts ($\times 10^3/\mu L$)</td>
<td>16.0 (6.4–33.2)</td>
</tr>
<tr>
<td>Post-Tx platelet counts ($\times 10^3/\mu L$)</td>
<td>16.8 (6.5–36.3)</td>
</tr>
<tr>
<td>Albumin (g/dL)</td>
<td>4.30 (2.9–5.5)</td>
</tr>
<tr>
<td>Pre-Tx AST (IU/mL)</td>
<td>60.0 (12–365)</td>
</tr>
<tr>
<td>Post-Tx AST (IU/mL)</td>
<td>20.0 (10–54)</td>
</tr>
<tr>
<td>Pre-Tx ALT (IU/mL)</td>
<td>100.0 (12–519)</td>
</tr>
<tr>
<td>Post-Tx ALT (IU/mL)</td>
<td>17.0 (7–64)</td>
</tr>
<tr>
<td>$\gamma$-GTP (IU/L)</td>
<td>37.0 (7–1790)</td>
</tr>
<tr>
<td>T. Bilirubin (mg/dL)</td>
<td>0.70 (0.3–1.9)</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>5.70 (4.4–8.1)</td>
</tr>
<tr>
<td>Pre-Tx AFP (ng/mL)</td>
<td>5.0 (1–200)</td>
</tr>
<tr>
<td>Post-Tx AFP (ng/mL)</td>
<td>3.0 (1–46)</td>
</tr>
<tr>
<td>Pre-Tx WFA$^+$-M2BP (COI)</td>
<td>1.70 (0.28–12.04)</td>
</tr>
<tr>
<td>Post-Tx WFA$^+$-M2BP (COI)</td>
<td>0.80 (0.17–5.29)</td>
</tr>
<tr>
<td>HCV serogroup, n (%)</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>111 (46.6)</td>
</tr>
<tr>
<td>2</td>
<td>101 (42.4)</td>
</tr>
<tr>
<td>Unknown</td>
<td>26 (11.0)</td>
</tr>
<tr>
<td>IFN regimen, n (%)</td>
<td></td>
</tr>
<tr>
<td>IFN monotherapy</td>
<td>123 (51.6)</td>
</tr>
<tr>
<td>PEG-IFN monotherapy</td>
<td>28 (11.8)</td>
</tr>
<tr>
<td>IFN/PEG-IFN+RBV</td>
<td>87 (36.6)</td>
</tr>
<tr>
<td>Observation period, years</td>
<td>9.1 (5.6) *</td>
</tr>
</tbody>
</table>

Data are given as the medians with ranges.

*Results are expressed as the means ± standard deviation. Unless otherwise indicated, data were collected at pre-treatment (before administration of IFN therapy; pre-Tx). Several biochemical measurements were made at both pre-Tx and post-treatment (24 weeks after completion of IFN therapy; post-Tx).

Abbreviations: AST, aspartate aminotransferase; ALT, alanine aminotransferase; $\gamma$-GTP, $\gamma$-glutamyl transpeptidase; HbA1c, glycated hemoglobin; BMI, body mass index; AFP, $\alpha$-fetoprotein; HCV, hepatitis C virus; PEG-IFN, pegylated interferon; RBV, ribavirin.

doi:10.1371/journal.pone.0129053.t001

Risk factors for HCC

Univariate analysis demonstrated factors that increase the risk for HCC development after SVR. Cox regression analysis was performed on 20 variables: age, sex, BMI, alcohol intake, fibrosis stage, degree of steatosis, pre-Tx platelet counts, post-Tx platelet counts, albumin, pre-Tx AST, post-Tx AST, pre-Tx ALT, post-Tx ALT, $\gamma$-GTP, T.bilirubin, HbA1c, pre-Tx AFP, post-Tx AFP, pre-Tx WFA$^+$-M2BP, post-Tx WFA$^+$-M2BP. Cutoff values for AFP and WFA$^+$-M2BP were determined by time-dependent ROC analysis as 5 ng/ml and 2.0 COI, respectively.
The following seven factors were identified as posing an increased risk for HCC by the univariate analysis: age, fibrosis stage, albumin, pre-Tx platelet count, post-Tx platelet count, post-Tx AFP, and post-Tx WFA⁺-M2BP (Table 2).

Multivariate analysis was performed on these seven factors, and the following four factors were identified as independent risk factors: age (> 60 years, HR 5.42, 95% CI = 1.59–18.47, \(P = 0.007\)), sex (male, HR 4.71, 95% CI = 1.23–17.92, \(P = 0.023\)), pre-Tx platelet count (< 15.0×10³/μL, HR 4.72, 95% CI = 1.45–15.30, \(P = 0.010\)), and post-Tx WFA⁺-M2BP values (> 2.0 COI, HR 5.71, 95% CI = 1.66–19.57, \(P = 0.006\)).

**Development of HCC**

To evaluate the relation between post-Tx WFA⁺-M2BP values and development of HCC, we characterized 238 patients who achieved SVR with respect to their post-Tx WFA⁺-M2BP values. Fig 1 shows the cumulative risk of HCC and the post-Tx WFA⁺-M2BP values. The 5 and 10-year cumulative risks of HCC were 1.9% and 5.0% in the 18 patients with post-Tx WFA⁺-M2BP > 2.0 COI (post-Tx WFA⁺-M2BP > 2.0 COI group), and 22.6% and 38.1% in the 220 patients with post-Tx WFA⁺-M2BP ≤ 2.0 COI (post-Tx WFA⁺-M2BP ≤ 2.0 COI group). The incidence rates were significantly higher in the post-Tx WFA⁺-M2BP > 2.0 COI group (\(P < 0.0001\) by the log-rank test).

Fig 2 shows the relation between the cumulative incidence of HCC and the post-Tx WFA⁺-M2BP values, stratified by age. In the patients with age > 60 years, the 5- and 10-year cumulative risks of HCC were 27.3% and 45.5% for the post-Tx WFA⁺-M2BP > 2.0 COI group. On the other hand, in the patients with age ≤ 60 years, the 5- and 10-year cumulative risks of HCC were 1.3% and 2.6% for the post-Tx WFA⁺-M2BP ≤ 2.0 COI group. There were significant differences in HCC incidence between the post-Tx WFA⁺-M2BP > 2.0 COI group and post-Tx WFA⁺-M2BP ≤ 2.0 COI group for both age categories (\(P < 0.0001\) by the log-rank test).

Fig 3 shows the relation between the cumulative incidence of HCC and the post-Tx WFA⁺-M2BP values, stratified by stage of fibrosis. In the patients with F3/4, the 5- and 10-year cumulative risks of HCC were 25.9% and 62.9% for the post-Tx WFA⁺-M2BP > 2.0 COI group. On the other hand, in the patients with F1/2, the 5- and 10-year cumulative risks of HCC were 1.3% and 3.9% for the post-Tx WFA⁺-M2BP ≤ 2.0 COI group. There were significant differences in HCC incidence between the post-Tx WFA⁺-M2BP > 2.0 COI group and post-Tx WFA⁺-M2BP ≤ 2.0 COI group with advanced fibrosis (F3/4) patients (\(P < 0.01\) by the log-rank test).

**Predictive value of HCC incidence versus WFA⁺-M2BP and AFP**

Table 3 shows the AUROC analyses for prediction of the development of HCC at 3, 5 and 10 years with AFP and WFA⁺-M2BP. The post-Tx WFA⁺-M2BP was superior to the post-Tx AFP for predicting the development of HCC at each of 3, 5 and 10 years.

**Chronological changes in the WFA⁺-M2BP and AFP values after IFN treatment**

In the 238 patients with SVR, the median values of the chronological change in WFA⁺-M2BP at pre-Tx and post-Tx were 1.70 (range: 0.28 to 12.04 COI) and 0.80 (range: 0.17 to 5.29 COI). The post-Tx WFA⁺-M2BP values were significantly decreased relative to the pre-Tx WFA⁺-M2BP values (\(P < 0.001\)).

Next, we analyzed the WFA⁺-M2BP and AFP values in the 16 patients who developed HCC. Fig 4A shows the chronological changes in WFA⁺-M2BP and AFP values at pre-Tx, post-Tx, and the time of HCC development for the 16 patients. The median WFA⁺-M2BP
Table 2. Factors Associated with Hepatocellular Carcinoma.

<table>
<thead>
<tr>
<th></th>
<th>Univariate analysis</th>
<th>Hazard Ratio (95% CI)</th>
<th>Univariate analysis</th>
<th>Hazard Ratio (95% CI)</th>
<th>P</th>
<th>Multivariate analysis</th>
<th>Hazard Ratio (95% CI)</th>
<th>Multivariate analysis</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (year)</td>
<td>≤ 60</td>
<td>1</td>
<td>&gt; 60</td>
<td>6.09 (2.03–18.26)</td>
<td>0.001</td>
<td>5.42 (1.59–18.47)</td>
<td>0.007</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Male</td>
<td>1.12</td>
<td></td>
<td></td>
<td>0.290</td>
<td>4.71 (1.23–17.92)</td>
<td>0.023</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sex</td>
<td>Female</td>
<td>1</td>
<td>Male</td>
<td>1.12</td>
<td>0.290</td>
<td>4.71 (1.23–17.92)</td>
<td>0.023</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>≤ 23.0</td>
<td>1</td>
<td>&gt; 23.0</td>
<td>2.09 (0.71–6.11)</td>
<td>0.167</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alcohol intake (g/day)</td>
<td>≤ 20</td>
<td>1</td>
<td>&gt; 20</td>
<td>1.60 (0.58–4.42)</td>
<td>0.364</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fibrosis stage</td>
<td>F1/2</td>
<td>1</td>
<td>F3/4</td>
<td>4.62 (1.67–12.81)</td>
<td>0.003</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Steatosis (%)</td>
<td>≤ 10</td>
<td>1</td>
<td>&gt; 10</td>
<td>1.12 (0.30–5.69)</td>
<td>0.561</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pre-Tx Platelet counts</td>
<td>≥ 15.0</td>
<td>1</td>
<td>&lt; 15.0</td>
<td>4.75 (1.52–14.79)</td>
<td>0.007</td>
<td>4.72 (1.45–15.30)</td>
<td>0.010</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(×10³/μL)</td>
<td>&gt; 15.0</td>
<td>1</td>
<td>&lt; 15.0</td>
<td>3.21 (1.20–12.96)</td>
<td>0.011</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Albumin (g/dL)</td>
<td>≥ 4.0</td>
<td>1</td>
<td>&lt; 4.0</td>
<td>3.40 (1.22–9.45)</td>
<td>0.018</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pre-Tx AST (IU/mL)</td>
<td>≤ 60</td>
<td>1</td>
<td>&gt; 60</td>
<td>2.13 (0.74–6.14)</td>
<td>0.146</td>
<td></td>
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<tr>
<td>Post-Tx AST (IU/mL)</td>
<td>≤ 20</td>
<td>1</td>
<td>&gt; 20</td>
<td>2.33 (0.30–18.29)</td>
<td>0.473</td>
<td></td>
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</tr>
<tr>
<td>Pre-Tx ALT (IU/mL)</td>
<td>≤ 80</td>
<td>1</td>
<td>&gt; 80</td>
<td>2.16 (0.79–5.83)</td>
<td>0.128</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Post-Tx ALT (IU/mL)</td>
<td>≤ 15</td>
<td>1</td>
<td>&gt; 15</td>
<td>2.44 (0.72–8.31)</td>
<td>0.180</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>γ-GTP (IU/L)</td>
<td>≤ 40</td>
<td>1</td>
<td>&gt; 40</td>
<td>1.73 (0.60–4.99)</td>
<td>0.297</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>T. Bilirubin (mg/dL)</td>
<td>≤ 1.0</td>
<td>1</td>
<td>&gt; 1.0</td>
<td>1.66 (0.37–7.35)</td>
<td>0.481</td>
<td></td>
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</tr>
<tr>
<td>HbA1c (%)</td>
<td>≤ 5.5</td>
<td>1</td>
<td>&gt; 5.5</td>
<td>1.07 (0.26–4.46)</td>
<td>0.929</td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Pre-Tx AFP (ng/mL)</td>
<td>≤ 5.0</td>
<td>1</td>
<td>&gt; 5.0</td>
<td>2.50 (0.89–7.06)</td>
<td>0.079</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Post-Tx AFP (ng/mL)</td>
<td>≤ 5</td>
<td>1</td>
<td>&gt; 5</td>
<td>4.60 (1.53–13.84)</td>
<td>0.006</td>
<td></td>
<td></td>
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<tr>
<td>Pre-Tx WFA⁺-M2BP (COI)</td>
<td>≤ 2.0</td>
<td>1</td>
<td>(COI)</td>
<td>1.37 (0.49–3.77)</td>
<td>0.551</td>
<td></td>
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</tr>
<tr>
<td>Post-Tx WFA⁺-M2BP (COI)</td>
<td>≤ 2.0</td>
<td>1</td>
<td>&gt; 2.0</td>
<td>7.30 (2.20–24.17)</td>
<td>0.001</td>
<td>5.71 (1.66–19.57)</td>
<td>0.006</td>
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</table>

Hazard ratios for the development of hepatocellular carcinoma were calculated by Cox proportional hazards analysis.

Abbreviations: AST, aspartate aminotransferase; ALT, alanine aminotransferase; γ-GTP, γ-glutamyl transpeptidase; HbA1c, glycated hemoglobin; BMI, body mass index; AFP, α-fetoprotein; HCV, hepatitis C virus; WFA⁺-M2BP, Wisteria floribunda agglutinin-positive human Mac-2 binding protein.

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values of the 16 patients who developed HCC at pre-Tx, post-Tx and the time of HCC development were 2.07 (range: 0.99 to 8.04 COI), 1.24 (range: 0.42 to 4.44 COI) and 0.79 (range: 0.41 to 2.79 COI). The median AFP values of 16 patients at pre-Tx, post-Tx and the time of HCC development were 8 (range: 2 to 63 ng/mL), 5 (range: 1 to 7 ng/mL) and 7 (range: 3 to 5463 ng/mL). The WFA+-M2BP values at the time of post-Tx were significantly lower than those at the time of pre-Tx ($P < 0.001$). Additionally, the WFA+-M2BP values at the time of HCC development were significantly lower than those at the time of post-Tx ($P < 0.001$). The AFP values at the time of post-Tx were significantly lower than those at the time of pre-Tx ($P < 0.001$). In contrast, the AFP values at the time of HCC development were significantly higher than those at the time of post-Tx ($P < 0.001$).

We also analyzed the WFA+-M2BP and AFP values in the 222 patients who did not develop HCC. The median WFA+-M2BP values of the 222 patients who did not develop HCC at pre-Tx, post-Tx and the last clinical visit were 2.07 (range: 0.99 to 8.04 COI), 1.24 (range: 0.42 to 4.44 COI) and 0.79 (range: 0.41 to 2.79 COI). The median AFP values of 16 patients at pre-Tx, post-Tx and the time of HCC development were 8 (range: 2 to 63 ng/mL), 5 (range: 1 to 7 ng/mL) and 7 (range: 3 to 5463 ng/mL). The WFA+-M2BP values at the time of post-Tx were significantly lower than those at the time of pre-Tx ($P < 0.001$). Additionally, the WFA+-M2BP values at the time of HCC development were significantly lower than those at the time of post-Tx ($P < 0.001$). The AFP values at the time of post-Tx were significantly lower than those at the time of pre-Tx ($P < 0.001$). In addition, the WFA+-M2BP values at the time of last clinical visit were significantly lower than those at the time of post-Tx ($P < 0.001$). The AFP values at the time of post-Tx were significantly lower than those at the time of post-Tx ($P < 0.001$).

![Cumulative incidence of hepatocellular carcinoma (HCC) according to post-treatment WFA+-M2BP values.](image)

Cumulative incidences of HCC according to post-treatment WFA+-M2BP values were analyzed using the Kaplan-Meier method. The black solid and dotted lines indicate the stratified post-treatment WFA+-M2BP values with a COI < 2.0 and a COI > 2.0, respectively. The incidence rate differed significantly between the two groups ($P < 0.001$ by the log-rank test). The numbers of patients at risk at each time point are shown below the graphs.

doi:10.1371/journal.pone.0129053.g001
Fig 2. Cumulative incidence of hepatocellular carcinoma (HCC) according to post-treatment WFA+-M2BP values, stratified by age. (A): Age < 60 years (n = 165). (B): Age > 60 years (n = 73). Cumulative incidences of HCC according to post-treatment WFA+-M2BP values were analyzed using the Kaplan-Meier method. The black solid and dotted lines indicate the stratified post-treatment WFA+-M2BP values with a COI < 2.0 and a COI > 2.0, respectively. The incidence rate differed significantly between the two groups (P < 0.0001 by the log-rank test). The numbers of patients at risk at each time point are shown below the graphs.

doi:10.1371/journal.pone.0129053.g002

Fig 3. Cumulative incidence of hepatocellular carcinoma (HCC) according to post-treatment WFA+-M2BP values, stratified by stage of fibrosis. (A): F1/2 (n = 172). (B): F3/4 (n = 66). Cumulative incidences of HCC according to post-treatment WFA+-M2BP values were analyzed using the Kaplan-Meier method. The black solid and dotted lines indicate the stratified post-treatment WFA+-M2BP values with a COI ≤ 2.0 and a COI > 2.0, respectively. There were no significant differences in HCC incidence with F1/2 group (P = 0.09 by the log-rank test). On the other hand, the incidence rate differed significantly with F3/4 group (P < 0.01 by the log-rank test). The numbers of patients at risk at each time point are shown below the graphs.

doi:10.1371/journal.pone.0129053.g003
time of pre-Tx ($P < 0.001$). Similarly, the AFP values at the time of last clinical visit were significantly higher than those at the time of post-Tx ($P < 0.001$) (Fig 4B).

Fig 5 shows the distribution of post-Tx WFA+-M2BP values. Among the 238 patients who achieved SVR, only 18 (7.6%) patients had post-Tx WFA+-M2BP $> 2.0$ COI. During the follow-up period, 5 patients (27.8%) developed HCC in the post-Tx WFA+-M2BP $> 2.0$ COI group ($n = 18$), and 4 of these 5 cases developed HCC within 5 years after IFN treatment. In contrast, 11 patients (5.0%) developed HCC in the post-Tx WFA+-M2BP $/ < 2.0$ COI group ($n = 220$) ($P < 0.001$).

**Discussion**

Previous studies have shown various risk factors for HCC development among patients with SVR: older age [14,15,16,18,19,33], male gender [19], heavy alcohol consumption [15,16], steatosis [33], advanced fibrosis [15,16,19], and lower platelet count [18]. In addition, recent studies reported that AFP values were significantly associated with HCC [18,27,34], and were also valuable for predicting future HCC risk after IFN treatment [35]. Asahina reported that post-IFN treatment AFP values were significantly associated with hepatocarcinogenesis [36]. In the present study, we specifically analyzed whether high WFA+-M2BP values might also be risk factors for HCC in patients with SVR.

In our study, the cumulative 5- and 10-year incidences of HCC were 3.4% and 7.5%. These results were consistent with previous studies, which reported that the cumulative incidences of HCC after SVR were 1.1–5.8% and 5.5–11.1% at 5 and 10 years [14,15,18,36–39], and the annual incidences were 0.37–1.21%/year [13,40,41].

The first main finding of our study was that the post-Tx WFA+-M2BP was selected as a new predictive marker for development of HCC among patients with SVR (Table 2). The values of WFA+-M2BP for predicting the development of HCC were determined to have a COI of 2.0 by time-dependent receiver operator characteristics (ROC) analysis. The cumulative incidence was significantly higher in the post-Tx WFA+-M2BP $> 2.0$ COI group. We were able to stratify the patients into different risk groups using the post-Tx WFA+-M2BP values and another simple risk factor, for example, age (Fig 2). Older age has been reported to confer a risk for hepatocellular carcinoma [14], which is an important association in Japan due to the aging of the population. Moreover, the post-Tx WFA+-M2BP values were significant predictor for HCC among patients with F3/4. Cumulative incidence of HCC was significantly higher in patients with higher post-Tx WFA+-M2BP values when patients were stratified by the stage of fibrosis (Fig 3). The post-Tx WFA+-M2BP values are not just a marker for liver fibrosis. Elevation of post-Tx WFA+-M2BP values as a potential risk for hepatocarcinogenesis with advanced fibrosis. And the time-dependent AUROC analysis suggested that WFA+-M2BP is superior to AFP as a predictor for the development of HCC.
Fig 4. Chronological changes in the WFA*+M2BP and AFP values at pre-treatment, post-treatment, the time of HCC development, and the last visit of the 238 patients with sustained virological response. Dots represent the median serum WFA*+M2BP values at each time point, and the error bar represents the interquartile range. Diamonds represent the median serum AFP values at each time point, and the error bar represents the interquartile range. (A): Patients who developed HCC (n = 16). WFA*+M2BP values were decreased at post-treatment and increased at HCC development. And AFP values were decreased at post-treatment and increased at HCC development. (B): Patients who did not developed HCC (n = 222). WFA*+M2BP values were decreased at post-treatment and last clinical visit. And AFP values were decreased at post-treatment and last clinical visit.

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Fig 5. Relationship between post-treatment WFA\(^+-\)M2BP values and HCC development. The distribution of WFA\(^+-\)M2BP values was plotted. The dashed line indicates the 2.0 COI for WFA\(^+-\)M2BP. The 16 patients who developed HCC were stratified according to the duration from SVR to HCC development in 5-year increments. Each time point is designated by a distinct symbol as indicated. 222 patients did not develop HCC, and 209 of these 222 patients (94.1%) were in the post-treatment WFA\(^+-\)M2BP /C20 2.0 COI group (white circles). During the follow-up period, 5 of 18 patients (27.8%) developed HCC in the post-treatment WFA\(^+-\)M2BP > 2.0 COI group, which was significantly higher than the rate in the post-treatment WFA\(^+-\)M2BP \leq 2.0 COI group (5.0%, \(P < 0.001\)). In the post-treatment WFA\(^+-\)M2BP > 2.0 COI group, 4 of 5 cases developed HCC within 5 years after IFN treatment (black circles).

doi:10.1371/journal.pone.0129053.g005
The second main finding of our study was that the WFA\(^+\)-M2BP values were decreased in patients who achieved SVR, even in those who developed HCC. Kuno previously reported that WFA\(^+\)-M2BP values were decreased by IFN treatment [20]; to this result we added that the WFA\(^+\)-M2BP values were decreased even in our IFN-treated patients who achieved SVR. However, the post-Tx WFA\(^+\)-M2BP values were significantly higher in the patients who developed HCC than in those who did not. This finding is particularly important because the post-Tx values of WFA\(^+\)-M2BP have not been adequately evaluated. Our data are thus the first to demonstrate the distribution of WFA\(^+\)-M2BP values at post-Tx.

The third main finding of our study was that AFP and WFA\(^+\)-M2BP values manifested different behaviors between the time of post-Tx and HCC diagnosis in patients who developed HCC. Our previous paper reported a close association between AFP values and the stage of fibrosis [27], whereas another report showed an elevation in AFP values caused by necroinflammation injury and regeneration of the liver [42]. However, WFA\(^+\)-M2BP values do not always correlate with the grade of hepatic activity as defined by HAI scoring of inflammation [20,28]. A slight elevation of post-Tx AFP values (> 5ng/mL) could indicate substantial risks for the development of HCC [36]. In the 16 patients who developed HCC in our study, AFP values were elevated from post-Tx to the time of HCC development. However, the WFA\(^+\)-M2BP values decreased after SVR and decreased further at the time of HCC diagnosis (Fig 4). The Mac-2 binding protein is secreted from many cell types, including hepatocytes, and it has been shown to modulate many processes, particularly those related to cell adhesion [22,43,44]. Alterations in the quality and quantity of the Mac-2 binding protein have been observed during the progression of fibrosis [22–24]. Hepatic stellate cells are considered the main fibrogenic cell type of the liver [45,46]. Activation of hepatic stellate cells and reversal of hepatic stellate cell activation [47] might be associated with WFA\(^+\)-M2BP values. WFA\(^+\)-M2BP has been associated with changes in both the quality and quantity of the Mac-2 binding protein due to changes in glycosylation [20]. From these considerations, we think that the WFA\(^+\)-M2BP values do not reflect the results of HCC development, but rather a pre-cancer status or hepatocellular carcinogenesis.

One of the limitations of the present study was its retrospective nature. A future prospective analysis will be needed to validate the efficacy of WFA\(^+\)-M2BP as a predictor of HCC development. Another limitation is that we analyzed a relatively small number of HCC cases after SVR. Multi-center prospective registration of patients with SVR could overcome this deficiency.

Regardless of these limitations, this is the first report to describe the relationship between WFA\(^+\)-M2BP values and HCC development after SVR. The rapid progress in the development of anti-viral agents [48,49] for hepatitis C suggests that the number of patients who achieve SVR—including elderly patients or patients with advanced fibrosis, who are regarded as being at high risk for HCC—might increase in the near future, especially in Japan. Therefore, the prediction of HCC development in patients with SVR is of increasing clinical relevance.

In conclusion, this study revealed an association between WFA\(^+\)-M2BP and the risk of HCC development in patients with SVR. The results suggested that the WFA\(^+\)-M2BP should not be limited to use in fibrosis stage screening but rather could be applied as a new predictor of HCC development after SVR.

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Author Contributions
Conceived and designed the experiments: RS KY HY. Performed the experiments: RS KY SA A. Komori SN AS SH SB YK HY. Analyzed the data: RS KY HY. Contributed reagents/materials/analysis tools: RS KY SA A. Komori SN AS SH SB YK A. Kuno MK AT MO MM HN HY. Wrote the paper: RS KY MM HN TI KN HY.

References


