Isolation of *Acanthamoeba* Genotype T4 from a Non-Contact Lens Wearer from the Philippines

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**Abstract:** We report the case of a 76-year old Filipino male who presented with pain, redness, and blurring of vision of the right eye. Corneal scraping was done and sent to the St. Luke’s Research and Biotechnology Group for detection and identification of the infectious agent. Morphological detection was performed by allowing the organism from the scraping to grow in 1.5% non-nutrient agar plate with heat-killed *E. coli*. Trophozoites with acanthopodia and double-walled cysts characteristic of *Acanthamoeba* were observed within the first and second week of observations, respectively. Molecular identification of the amoebae at the genus level based on the presence of *Acanthamoeba*-specific amplimer S1, ASA.S1 confirmed the morphological identification. Genotyping through sequence revealed that the organism belonged to T4, which is the genotype commonly present in the eye of keratitis patients.

**Key words:** *Acanthamoeba* genotype T4, non-contact lens wearer, Philippines

We report the case of a 76-year old Filipino male from the province of Porac, Pampanga who presented with pain, redness, and blurring of vision of the right eye (oculus dexter, OD) on 25 February 2009. One and a half weeks prior to consultation, the patient complained of right eye discharge and lid swelling. He consulted an ophthalmologist and was given unrecalled topical eye medications, but no improvement of symptoms was noted. The patient complained of worsening pain and redness in the right eye. A non-contact lens wearer, he denied any trauma to the right eye. However, he admitted to washing both eyes with tap water. Visual acuity in the right eye was counting fingers at 3 feet. Slit lamp examination showed diffuse conjunctival injection. A ring-like infiltrate was noted in the cornea almost extending to the limbus. Significant anterior chamber cell reaction and hypopyon were also observed. The primary working impression was *Acanthamoeba* keratitis (AK), OD (Fig. 1).

The patient was referred to another ophthalmologist for further evaluation and management. Corneal scraping was done and sent to the St. Luke’s Research and Biotechnology Group for detection and identification of the infectious agent. Morphological detection was performed by allowing the organism from the scraping to grow in 1.5% non-nutrient agar (NNA) plate with heat-killed *E. coli*. Trophozoites with acanthopodia and double-walled cysts characteristic of *Acanthamoeba* were observed within the eye of the patient.

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The isolate was then grown axenically in 2% proteose-peptone, 0.2% yeast, 1.8% glucose (PPYG) with 100 units of penicillin G sodium and 100 μg streptomycin sulfate/ml of PYPG (Fig. 2).

The DNA from *Acanthamoeba* isolated from the infected eye of the patient was extracted using the DNeasy Blood and Tissue Kit according to the manufacturer’s instructions (Qiagen, Hilden, Germany). PCR amplification was carried out with 1X Phusion HF buffer, 0.2 mM dNTPs, 0.50 μM of genus-specific JDP1 sense (5'-GGCCCAAGATCGTTTACCGTGAA-3') and JPD2 (5'-TCTCACAAGCTGCTAGGGGAGTCA-3') antisense primer [1], 0.02 U Phusion DNA polymerase, and PCR-grade water with the following thermal profile: initial denaturation at 95°C for 7 minutes followed by 45 cycles at 95°C for 1 minute, 60°C for 1 minute, 72°C for 2 minutes, and final elongation at 72°C for 10 minutes. The PCR products were purified using the QIAquick PCR Purification Kit according to the manufacturer’s protocol (Qiagen, Hilden, Germany). The amplicons were sequenced using an automated sequencer (First BASE Laboratories, Selangor, Malaysia). Sanger dideoxy sequencing of the PCR product was carried out using conserved 892 (5'-CCAAGAATTTCACCTCTGAC-3') sense and 892C (5'-GTCAGAGGTGAAATTCTTGG-3') antisense primers to determine the sequence of diagnostic fragment (DF3) of the 18S ribosomal DNA (Rns). Nucleotide sequences were compared for identity with sequences from NCBI utilizing the BLAST program. Molecular identification of the amoebae at the genus level based on the presence of *Acanthamoeba*-specific amplimer S1, ASA.S1 (a partial 18S ribosomal DNA gene) confirmed the morphological identification (Fig. 3) [1, 2]. Genotyping through sequence revealed that the organism belonged to T4, which is the genotype commonly present in the eye of keratitis patients (Fig. 3) [1, 2].

The nucleotide sequence reported in this study has been deposited to NCBI and can be retrieved under GenBank accession number KJ995957. To the best of our knowledge, this is the first documentation of the genotype of *Acanthamoeba* directly isolated from a non-contact lens wearer patient in the Philippines. However, in 1992, Philippine *Acanthamoeba* isolate IB-1-7 was reported to have been obtained from a patient with keratitis at the Philippine General Hospital. The same report also noted isolation of *Acanthamoeba* from a deep-well water source in Quezon City. These two previously reported isolates were not genotyped [3].

*Acanthamoeba* is one of the leading causes of keratitis, a painful sight-threatening corneal infection. The ma-
Majority of AK strains are associated with genotype T4, but AK strains belonging to other genotypes have also been reported worldwide [4–5]. In the Philippines, *Acanthamoeba* isolates from contact lens wearers were found to be either genotype T4 or T5 [6]. AK cases mostly occur in contact lens wearers and people with a history of ocular trauma. The free-living amoeba is also responsible for life-threatening infections such as granulomatous amebic encephalitis, a fatal disease of the central nervous system seen among immuno-compromised patients [7].

Non-contact lens wearers comprised 3% to 15% of AK cases in the United Kingdom and United States [8]. Recently, it was reported that 27% of the recreational water samples collected in Batangas and Pampanga, Philippines were positive for *Acanthamoeba* by PCR. *Acanthamoeba* spp. was found in river, pond, lake, and tap water samples from the said provinces [9]. This study corroborates and extends previous findings which showed that risk factors other than contact lens wear, corneal trauma and contamination of the water supply can predispose infection [2, 5].

Based on the clinical findings and laboratory results, the final diagnosis was AK. The patient was treated with chlorhexidine. The corneal infection subsided and was resolved after two months of treatment. A corneal scar was noted and corneal transplant contemplated, but the patient was lost to follow-up.

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**References**