A homozygous splice site ROBO1 mutation in a patient with a novel syndrome with combined pituitary hormone deficiency

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Running title: A ROBO1 mutation in a patient with hypopituitarism

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Abstract
The genetic causes of combined pituitary hormone deficiency remain elusive in most patients. Recently, incompletely penetrant heterozygous mutations in ROBO1 have been described in patients with pituitary stalk interruption syndrome. Herein, we identified a novel homozygous slice site mutation in ROBO1 (c.1342+1G>A) using a trio whole-exome sequencing strategy in a five-year-old Japanese boy who had combined pituitary hormone deficiency, psychomotor developmental delay, severe intellectual disability, sensorineural hearing loss, strabismus, and characteristic facial features, including a broad forehead, micrognathia, and arched eyebrows. Magnetic resonance imaging delineated anterior pituitary hypoplasia, ectopic posterior pituitary, invisible pituitary stalk, thinning of the corpus callosum, and hypoplasia of the pons and midbrain. The phenotypically normal parents (first cousins) were heterozygous for the mutation. The results provide further evidence of ROBO1 being involved in the development of the pituitary gland. A recessive mutation of ROBO1 is a potential novel cause of a syndromic disorder associated with combined pituitary hormone deficiency.
Introduction

Normal pituitary development requires a complex genetic cascade of transcription factors and signaling molecules, either intrinsic or extrinsic to the developing Rathke's pouch [1]. Mutations of genes involved in these processes, including POUIF1, PROP1, HESX1, LHX3, LHX4, OTX2, GLI2, and SOX2, are associated with a wide range of pituitary phenotypes, such as isolated growth hormone (GH) deficiency and combined pituitary hormone deficiency (CPHD), which is defined as the presence of hormone deficits affecting at least two anterior pituitary hormone lineages [1, 2]. However, the definitive genetic causes remain obscure in the majority of patients with CPHD [2, 3].

Congenital hypopituitarism is frequently associated with other extrapituitary abnormalities, such as anophthalmia/microphthalmia, optic nerve hypoplasia, dysgenesis of the corpus callosum, absence of the septum pellucidum, and holoprosencephaly, suggesting that defects in signaling molecules or transcription factors involved in the development of the forebrain result in such syndromic disorders [4].

The roundabout guidance receptors (ROBOs) and their Slit guidance ligands play critical roles in axonal guidance, which is essential for the formation of the neuronal network in the central nervous system. ROBO1 acts as the gatekeeper controlling the midline crossing of axons [5].

In the present study, we identified a homozygous splice-acceptor site mutation in the ROBO1 gene in a patient with a characteristic syndromic disorder associated with CPHD. Our study implies that recessive ROBO1 null mutations cause a novel neurodevelopmental syndrome associated with CPHD.

Materials and methods

Case reports

This Japanese male patient was born at 38 weeks of gestation as the first child of consanguineous phenotypically normal parents (first cousins) with no other significant family history. At birth, his length was 50.0 cm (+0.9 standard deviation [SD]), his weight 3.28 kg (+1.2 SD), and his head circumference 37.5 cm (+3.4 SD). He had distinct facial features with a broad forehead, micrognathia, a broad philtrum, and arched eyebrows (Fig. 1a). He also had hypotonia, micropenis, cryptorchidism, strabismus, and sensorineural deafness. Brain magnetic resonance imaging delineated hydrocephalus, anterior pituitary hypoplasia,
ectopic posterior pituitary, invisible pituitary stalk, thinning of the corpus callosum, and hypoplasia of the pontine and midbrain (Fig. 1b, c).

At 20 days of age, he developed recurrent hypoglycemia and conjugated hyperbilirubinemia. A hormonal examination for critical samples obtained at the time of spontaneous presentation of hypoglycemia showed central hypothyroidism and low serum cortisol and plasma ACTH levels, suggesting the presence of CPHD (Table 1). He was therefore started on thyroid hormone and hydrocortisone replacement therapies. At 18 months of age, his height was 64.5 cm (-5.1 SD). Endocrine studies at that time confirmed the diagnosis of CPHD (associated with deficiencies of GH, TSH, prolactin, LH, FSH, and ACTH) (Table 1), and recombinant human GH therapy was started.

At the final examination at 5 years of age, his motor and mental development was severely retarded. He was unable to speak any meaningful words and sit alone. Diffusion tensor imaging and fiber tractography, performed after the genetic diagnosis, demonstrated thinning of the corpus callosum and the anterior commissure but showed the presence of transverse pontine fibers (Fig. 1d).

**Molecular studies**

This study was approved by the Institutional Review Board at Nagasaki University Graduate School of Biomedical Sciences. Trio whole-exome sequencing was performed using a SureSelect Human All Exon V5 (Agilent Technologies, Santa Clara, CA, USA) on a HiSeq 2500 platform (Illumina, San Diego, CA, USA). DNA was obtained from peripheral blood samples of the patient and the parents after written informed consent was obtained from the parents. The reads in the FASTQ files were aligned to the human reference genome using Novoalign version 3.0 (http://www.novocraft.com/). The mean depth of the RefSeq coding region was 140.54 with 97.2% of total coding sequences covered by 20 reads or more in the proband. Trio-based genomic variation information was detected by the Genome Analysis Toolkit software version 3.4-46 [6]. Subsequently, de novo, homozygous, compound heterozygous, and X-linked variations in exons and canonical splice sites (±2 bp) were extracted and annotated by the ANNOVAR software [7]. This process excluded variants with allele frequencies >0.5% in any of the Exome Aggregation Consortium (ExAC) (http://exac.broadinstitute.org/), NHLBI GO Exome Sequencing Project (http://evs.gs.washington.edu/EVS/), Human Genetic Variation Database (http://www.hgvd.genome.med.kyoto-u.ac.jp), the 1KJPN database of Tohoku Medical
Megabank (http://www.dist.megabank.tohoku.ac.jp), and in-house exome data. Heterozygous variations sharing the same GENCODE v19 genes were also extracted to detect compound heterozygous mutations. The candidate variants identified in the strategy were confirmed via Sanger sequencing.

Results

Using the trio-based strategy and the filtering methods, we identified eight candidate variants consisting of six homozygous and two X-linked hemizygous variants (Supplemental Table 1). Of these, a homozygous splice-acceptor site mutation in ROBO1 (c.1342+1G>A, NM_002941) was proposed as the best candidate by the Online Mendelian Inheritance in Man database information of known diseases (www.omim.org) (Fig. 2a, b). This splice mutation is predicted to cause exon skipping and frameshift mediating nonsense-mediated mRNA decay (http://www.fruitfly.org/seq_tools/splice.html) [8]. The father and the mother of the proband were heterozygous for the mutation. The patient had no other pathogenic mutations in genes known to cause CPHD [2].

Discussion

We identified a homozygous ROBO1 splice-acceptor site mutation in a patient with syndromic CPHD and summarized the genetic and clinical features of patients previously reported to have ROBO1 mutations (Table 2). To our knowledge, the combination of his clinical manifestations has not been reported thus far. Therefore, we propose the null homozygous mutation of ROBO1 as the likely genetic cause of a novel syndrome associated with CPHD, based on the following: First, Robo1 null mice, which die shortly after birth, show defects in axon pathfinding with dysgenesis of the corpus callosum and the hippocampal commissure [5], phenotypically similar to those of patients with biallelic ROBO1 mutations (the present patient and Case 6 in Table 2). Second, heterozygous mutations in ROBO1 have been recently reported in five patients (Cases 1 through 5 in Table 2) from three independent families with pituitary stalk interruption syndrome [9] and variable pituitary phenotypes ranging from isolated GH deficiency to CPHD, indicating that ROBO1 is involved in the pituitary development and function (Table 2). However, the penetrance of the dominant ROBO1 mutations seems to be incomplete, as phenotypically normal members in the pedigrees also had the same mutation. Indeed, the present parents, harboring a heterozygous ROBO1 mutation, seem phenotypically normal. Furthermore, heterozygous
ROBO1 loss-of-function variants, including nonsense, frameshift, and splice site mutations, are described in the ExAC database. Third, not only homozygous but also heterozygous patients exhibit various ophthalmological phenotypes, such as strabismus, optic nerve hypoplasia, and hypermetropia (Table 2) [9, 10]. This may not be surprising, considering that Robo/Slit signaling plays a critical role in the extension of the retinal ganglion cell axons from the eye to the brain and formation of the optic chiasm [11]. Fourth, a patient with biallelic compound heterozygous missense variants in ROBO shared some phenotypes with the present patient, such as intellectual disability and thinning of the anterior commissure and corpus callosum [10]. However, the previously reported patient did not exhibit any abnormalities of the pituitary gland, indicating that the pituitary phenotypes in patients with biallelic ROBO1 mutations may be variable. Taken together, these findings imply that the homozygous ROBO1 null mutations cause a characteristic neurodevelopmental disorder with CPHD and defects in axon pathfinding, and heterozygous mutations may also cause diverse clinical features, ranging from nearly normal to pituitary stalk interruption syndrome and showing a wide range of penetrance, with expressivity depending on other genetic and environmental factors.

The pathological mechanisms of invisible stalk and pituitary dysfunction in patients with ROBO1 mutations remain obscure. Since ROBO1 defects possibly lead to abnormal axon elongation of magnocellular neurons from the paraventricular and supraoptic nuclei of the hypothalamus to the posterior pituitary, ROBO1 mutations may affect close relationship and tissue interactions between oral and neural ectoderm, which are critical for development and differentiation of the pituitary gland. Therefore, it is reasonable to hypothesize that ROBO1 mutations result in pituitary dysmorphogenesis and dysfunction [1].

In conclusion, the results provide further evidence of the involvement of ROBO1 in the pituitary development. Recessive null mutations of ROBO1 may cause novel syndromic CPHD. At this time, however, the phenotypic spectrum and mechanisms underlying the development of pituitary dysfunction remain to be determined in patients with ROBO1 mutations. These matters await further investigations.

Conflicts of interest
The authors declare no conflicts of interest.

Acknowledgements
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References


**Titles and legends to figures**

**Figure 1. Clinical findings in the patient.**

(a) A front view of the patient at one year of age showing distinct facial features with strabismus, a broad forehead, micrognathia, a broad philtrum and arched eyebrows. T1-weighted sagittal (b) and coronal (c) views of the brain magnetic resonance imaging show anterior pituitary hypoplasia, ectopic posterior pituitary (white arrow), thinning of the corpus callosum, and the pontine and the midbrain hypoplasia. (d) Diffusion tensor imaging and fiber tractography showing the presence of transverse pontine fibers. The authors have obtained informed consent from his parents to publication of these images.

**Figure 2. Mutation analyses of ROBO1 in this family.**

(a) Electrochromatograms delineating the homozygous mutation in a splice-acceptor site (c.1342+1G>A, NM_002941) in the patient (asterisk) and the heterozygous ones in the parents. The mutation was confirmed by direct sequencing. (b) Pedigree of the family. The black-painted square indicates the presence of the homozygous variant. Half-black, half-white symbols represent carriers of the variant in a heterozygous form.
Table 1. Blood hormone values of the patient with a homozygous ROBO1 mutation

<table>
<thead>
<tr>
<th>Hormone</th>
<th>Stimulus (dosage)</th>
<th>Baseline</th>
<th>Peak</th>
<th>Reference values</th>
</tr>
</thead>
<tbody>
<tr>
<td>GH (ng/ml)</td>
<td>Arginine (0.5 g/kg)</td>
<td>0.17</td>
<td><strong>0.68</strong></td>
<td>&gt;6^b</td>
</tr>
<tr>
<td></td>
<td>L-Dopa (10 mg/kg)</td>
<td>0.2</td>
<td><strong>0.27</strong></td>
<td>&gt;6^b</td>
</tr>
<tr>
<td>LH (mIU/ml)</td>
<td>GnRH (2.5 µg/kg)</td>
<td>&lt;0.1</td>
<td>&lt;0.1</td>
<td>0.4-6.0^b</td>
</tr>
<tr>
<td>FSH (mIU/ml)</td>
<td>GnRH (2.5 µg/kg)</td>
<td>0.1</td>
<td><strong>0.21</strong></td>
<td>6.3-15.6^b</td>
</tr>
<tr>
<td>TSH (µU/ml)</td>
<td>TRH (10 µg/kg)</td>
<td>0.01</td>
<td><strong>0.01</strong></td>
<td>&gt;10^b</td>
</tr>
<tr>
<td>Prolactin (ng/ml)</td>
<td>TRH (10 µg/kg)</td>
<td>2.64</td>
<td><strong>3.61</strong></td>
<td>&gt;2 times of the basal value^b</td>
</tr>
<tr>
<td>ACTH (pg/ml)</td>
<td></td>
<td><strong>3.6a</strong></td>
<td></td>
<td>12.6-35.0</td>
</tr>
<tr>
<td>Cortisol (µg/dl)</td>
<td></td>
<td><strong>0.5a</strong></td>
<td></td>
<td>5-20</td>
</tr>
<tr>
<td>IGF-I (ng/ml)</td>
<td></td>
<td>&lt;0.1</td>
<td></td>
<td>14-148</td>
</tr>
<tr>
<td>Free T4 (ng/dl)</td>
<td></td>
<td><strong>0.7</strong></td>
<td></td>
<td>1.01-1.95</td>
</tr>
</tbody>
</table>

Hormone values have been evaluated by the age-and sex-matched Japanese reference data; low hormone data are boldfaced. Blood sampling during the provocation tests: 0, 30, 60, 90, and 120 minutes. ^aMeasured at two months of age in his critical samples obtained at time of spontaneous presentation of hypoglycemia (35 mg/dL). ^bPeak values during the provocation tests.
Table 2. Clinical and genetic features of patients with ROBO1 mutations

<table>
<thead>
<tr>
<th>Case #</th>
<th>Bashamboo et al (^a)</th>
<th>Calloni et al (^b)</th>
<th>Present case</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yrs)</td>
<td>2.6</td>
<td>2.6</td>
<td>1</td>
</tr>
<tr>
<td>Sex</td>
<td>Male</td>
<td>Female</td>
<td>Male</td>
</tr>
</tbody>
</table>

**ROBO1 mutations**

- **Allele 1**
  - c.2928-2929delG
  - p.Ala977Glnfs*40
- **Allele 2**
  - WT

**Birth measurements**

<table>
<thead>
<tr>
<th>Birth measurements</th>
<th>Bashamboo et al (^a)</th>
<th>Calloni et al (^b)</th>
<th>Present case</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gestational age (wks)</td>
<td>39</td>
<td>39</td>
<td>40</td>
</tr>
<tr>
<td>Weight (SD)</td>
<td>2800 g</td>
<td>2950 g</td>
<td>3580 g</td>
</tr>
<tr>
<td>Height (SD)</td>
<td>48 cm</td>
<td>49 cm</td>
<td>48.5 cm</td>
</tr>
<tr>
<td>OFC (SD)</td>
<td>34 cm</td>
<td>34 cm</td>
<td>36 cm</td>
</tr>
</tbody>
</table>

**Clinical findings**

- **Affected pituitary hormones**
  - GH
  - GH
  - GH
  - GH, TSH
  - GH, TSH, ACTH, N.D
  - GH, TSH, PRL, ACTH, LH/FSH
- **Short stature**
  - +
  - +
  - +
  - N.D
- **Ophthalmologic defects**
  - Strabismus
  - Strabismus
  - Strabismus
  - Optic tract defect
  - Strabismus
- **Developmental delay**
  - -
  - -
  - -
  - +
  - +
- **Intellectual disability**
  - -
  - -
  - -
  - +
  - +
- **Hypotonia**
  - -
  - -
  - -
  - +
  - +
- **Dysmorphic facial features**
  - -
  - N.A
  - -
  - N.A
  - N.A
- **Hearing loss**
  - -
  - -
  - -
  - -
  - +
- **Micropenis**
  - N.A
  - N.A
  - N.A
  - N.A
  - N.A
- **Cryptorchidism**
  - -
  - N.A
  - N.A
  - N.A
  - -
- **Other findings**
  - -
  - Ptosis
  - Cardiomyopathy
  - Spastic diplegia
  - Ataxia, dysmetria

**MRI findings**

- Anterior pituitary hypoplasia
  - +
  - +
  - +
  - +
  - -
- Ectopic posterior pituitary
  - +
  - +
  - +
  - -
  - +
- Invisable pituitary stalk
  - +
  - +
  - +
  - -
  - +
- Hypoplastic corpus callosum
  - -
  - -
  - -
  - -
  - +
- Hypoplastic pontine
  - -
  - -
  - -
  - -
  - +
- Hypoplastic midbrain
  - -
  - -
  - -
  - -
  - +
- Hydrocephalus
  - -
  - -
  - -
  - -
  - +

**Abbreviations:** OFC, occipitofrontal circumference; WT, wild type; PRL, prolactin; +, Present; -, Absent; N.D., not described; N.A, not applicable.

\(^a\)NM_002941, NP_002932. \(^b\)Cases 1 and 2 are nonidentical twins. \(^c\)Case 5 is a paternal aunt of Case 4.
Figure 2

(a) Exon 10 and Intron 10 sequences for the proband, father, and mother.

(b) Pedigree chart showing family relationships.
### Supplemental Table 1. Candidate pathogenic variants in the patient.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Locus</th>
<th>Inheritance pattern</th>
<th>Transcript ID</th>
<th>Mutation</th>
<th>Allele frequency</th>
<th>Number of homozygotes</th>
<th>Predicted Pathogenesity</th>
<th>Associated diseases</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>SIFT&lt;sup&gt;b&lt;/sup&gt;</td>
<td>PolyPhen-2&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>PSIS (AD), Axonal guidance disorder (AR)</td>
</tr>
</tbody>
</table>

| PKN2  | 1p22.2| AR                  | ENST00000370505.3 | missense c.2455G>C p.Asp819His | 0                 | -                     | 0.001 (D) | 0.998 (D) | Unknown |
| ROBO1 | 3p12.3| AR                  | ENST00000464233.5 | splice site c.1342+1G>A          | 0                 | -                     | -         | -         | -       |
| CCDC91| 12p11 | AR                  | ENST00000539107.1 | missense c.585T>A p.His195Glu    | 0                 | -                     | 0.015 (D) | 0.555 (P) | Unknown |
| CYP1A2| 15q24.1| AR                   | ENST00000343932.4 | missense c.130G>A p.Glu44Lys     | 0.00001688         | 0                     | 0.001 (D) | 0.118 (B) | Unknown |
| IMP3  | 15q24.2| AR                  | ENST00000314852.2 | missense c.130G>T p.Asp44Tyr     | 0.0000542          | 0                     | 0 (D)     | 0.718 (P) | Unknown |
| TTLL12| 22q13.2| AR                   | ENST00000343932.5 | missense c.1508A>G p.Asn503Ser    | 0.0000167          | 0                     | 0.056 (T) | 0.042 (B) | Unknown |
| ACE2  | Xp22.2| X linked            | ENST00000252519.3 | missense c.1402A>G p.Ile468Val   | 0.0007253          | 0                     | 0.097 (T) | 0.651 (P) | Unknown |
| DUSP9 | Xq28  | X linked            | ENST00000342782.3 | missense c.521C>T p.Ala174Val    | 0.0000704          | 0                     | 0.257 (T) | 0 (B)     | Unknown |

AR, autosomal recessive; AD, autosomal dominant; PSIS, pituitary stalk interruption syndrome.

<sup>a</sup> Data from the Exome Aggregation Consortium database.

<sup>b</sup> SIFT (http://sift.jcvi.org/); D: deleterious (≦0.05), T: tolerated (>0.05).

<sup>c</sup> PolyPhen-2 (http://genetics.bwh.harvard.edu/pph2/); D: Probably damaging (≧0.957), P: possibly damaging (0.453≦, ≦0.956); B: benign (≦0.452)