Effect of surface removal following bleaching on the bond strength of enamel

Yi-ling Cheng¹, Joseph Musonda¹, Hui Cheng¹, Thomas Attin², Ming Zheng¹ and Hao Yu¹,³*

Abstract

Background: A reduction in bond strength of bleached enamel has been confirmed in the literature. Although limited information is available, it is conceivable that the veneer preparation process may remove the impacted enamel and further eliminate the compromised bond strength between the composite resin and bleached enamel. This study aimed to evaluate the effect of surface removal following bleaching on the micro-shear bond strength (μSBS) of bleached enamel.

Methods: Forty-eight specimens were prepared from bovine incisors and were randomly divided into 2 groups (n = 24): group B (bleaching with 40% hydrogen peroxide for 2 × 45 min with a 1-week interval) and group C (control group without bleaching treatment). Immediately after receiving the treatments, 0.5 mm of the enamel was removed from the specimen surface, followed by bonding of composite resin to the enamel surface. Each group was further divided into 2 subgroups of 12 specimens each: subgroup T (with 5000 thermocycles in water baths at 5 °C and 55 °C), and subgroup N (without thermocycling). The μSBS values were measured using a universal testing machine and subjected to two-way analysis of variance (α = 0.05). The fracture modes of the specimens were observed using a stereomicroscope.

Results: The μSBS values of the different groups ranged from 21.42 to 25.21 MPa. Following a surface reduction of 0.5 mm, bleaching treatment and thermocycling did not significantly affect the μSBS values (P = 0.348 and P = 0.507, respectively). No significant interaction was found between the bleaching treatment and thermocycling (P = 0.514). All the groups exhibited a high percentage of mixed failures. Compared with group C, group B exhibited higher percentage of adhesive failure.

Conclusion: The results suggested that the bonding procedure could be performed on the bleached enamel following a surface reduction of 0.5 mm immediately after the bleaching treatment.

Keywords: Enamel, Hydrogen peroxide, Shear bond strength

Background

Having whiter teeth has become increasingly popular in recent years because in some communities it has a direct effect on gaining individual confidence and social recognition [1]. Depending on the varied severity and types of tooth discoloration, tooth color can be improved by tooth bleaching, tooth scaling and polishing, and veneers, etc. [2].

Although tooth bleaching has been proven to be a safe and effective treatment [3, 4], it may present certain adverse effects on the enamel, such as surface morphological changes [5, 6], compositional changes [7, 8], alterations in the surface microhardness [9–11] and surface roughness [12]. Moreover, there is evidence that tooth bleaching may lead to reduction in bond strength of composite resin applied on previously bleached enamel [13, 14]. A reduction in bond strength of 25–60% has been reported when the bonding procedure is performed on an enamel surface immediately after bleaching [14–16]. During bleaching treatment, hydrogen peroxide undergoes ionic dissociation and increases the formation of free radicals, such as nascent oxygen and the hydroxyl radical, on the enamel surface [17]. Although the mechanism is...
still not clear, the reduced bond strength of bleached enamel has been related to the presence of residual free radicals due to the breakdown of hydrogen peroxide \cite{14,18} and alterations in the enamel composition and structure \cite{6,19} following the bleaching treatment. The residual oxygen in the interprismatic spaces can hamper resin infiltration and inhibit resin polymerization \cite{20}. Moreover, morphological and compositional changes (e.g., porosity, loss of enamel prismatic form, loss of calcium, and changes in organic substances) in the enamel may weaken the adhesive interface and compromise bond strength \cite{21,22}. Therefore, bonding procedures should not be performed immediately after bleaching treatment \cite{23}. A waiting period of 1–3 weeks has been advocated by various researchers \cite{21,24,25}. In addition to the delayed bonding procedure, the application of antioxidant agents (e.g., sodium ascorbate, sodium bicarbonate, and grape seed extract) \cite{18,26,27} and laser irradiation \cite{28,29} have been proposed to restore the compromised bond strength of bleached enamel. By neutralizing residual free radicals \cite{30} and promoting micro-retentions in the enamel surface \cite{28}, antioxidant agents and laser irradiation have been shown to reverse the reduced bond strength between the composite resin and bleached enamel. However, it is important to point out that most of the above-mentioned studies measured the bond strength without thermocycling \cite{24–29}. Thermocycling is the in vitro process of subjecting a restoration and tooth to temperature limits similar to those experienced in the oral cavity \cite{31}. It would be of interest to investigate the effects of thermocycling on the bond strength between the composite resins and the bleached enamel.

In cases of severely discolored dentition and the need for tooth shape corrections, esthetic reconstructions, such as laminate and composite veneers, are often required after the bleaching treatment. Given that most of the negative effects of bleaching have been found on the enamel surface \cite{9}, it is conceivable that the veneer preparation process may remove the impacted enamel and further eliminate the compromised bond strength between the composite resin and bleached enamel. However, few studies have addressed this issue.

Therefore, the aim of the present study was to evaluate the effect of surface removal following bleaching on the micro-shear bond strength (\(\mu\)SBS) between composite resin and bleached enamel immediately after bleaching treatment. The following null hypotheses were tested: (1) that the bleaching treatment would not affect the bond strength between the composite resin and the enamel when a surface reduction of 0.5 mm was performed immediately following bleaching; (2) that the bond strength between the composite resin and the enamel would be the same after thermocycling.

Methods

The research protocol was reviewed and approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015–CX-31).

Specimen preparation

Forty-eight specimens were prepared from freshly extracted bovine incisors. The bovine teeth were collected as anonymous by-products of regular slaughtering of the cattle for human food consumption. Enamel blocks (4 mm × 4 mm × 4 mm) were cut from the middle third of the buccal surfaces using a low speed diamond saw (Isomet, Buehler, Lake Bluff, IL, USA) under water cooling. The enamel blocks were further embedded in acrylic resin (ZiRan, Nissin, Hangzhou, Zhejiang, China) using a polyvinylchloride ring mold of a diameter of 10 mm. After ultrasonically cleaning in distilled water for 1 min, the specimens were randomly divided into 2 groups of 24 specimens each, according to the bleaching treatment performed.

In group B (bleaching group), the specimens were bleached with 40% hydrogen peroxide gel (Opalescence Boost 40%, Ultradent Product Inc., South Jordan, Utah, USA). Two bleaching sessions with a 1-week interval were performed in a wet chamber at 37 °C. For each session, bleaching gels were applied to the enamel surface at a thickness of 1 mm for 15 min and were renewed three times. After each bleaching session, the bleaching gels were washed away from the specimen surfaces with distilled water. During the bleaching intervals, the specimens were stored in artificial saliva at 37 °C. The artificial saliva was mixed according to the formulation described by Klimek et al. \cite{32}. 500 ml of artificial saliva contained 0.001 g ascorbic acid, 0.015 g glucose, 0.290 g NaCl, 0.085 g CaCl\(_2\), 0.080 g NH\(_4\)Cl, 0.635 g KCl, 0.080 g NaSCN, 0.165 g KH\(_2\)PO\(_4\), 0.100 g carbamide, 1.350 g mucin and 0.170 g Na\(_2\)PO\(_4\).

In group C (control group), no bleaching treatment was performed on the specimen surfaces. The specimens were immersed in artificial saliva during the experimental period.

Bonding procedure

Immediately following the abovementioned treatments, 0.5 mm of the enamel was removed from the specimen surface with the aid of a depth-cut diamond bur (DC 5, Premier Dental Products, Plymouth Meeting, PA, USA) and a series of carborundum discs (400#, 600#, Buehler), under water irrigation. Enamel reduction was controlled with a micrometer (Mitutoyo, Kyoto, Japan). The specimens were then ultrasonically cleaned in distilled water for 5 min to remove any debris. The surfaces were observed with a stereomicroscope (MM400, Nikon, Tokyo, Japan), and the specimens exposing the cracks or dentin surface were discarded.

After cleaning with pumice, the enamel surface of each specimen was etched with 35% phosphoric acid (Scotchbond Etchant, 3 M ESPE, St Paul, MN, USA) for 15 s, rinsed with...
distilled water for 20 s, and dried thoroughly. Two consecutive coats of adhesive agents (Adper Single Bond 2, 3 M ESPE, St Paul, MN, USA) were applied and light polymerized for 10 s with an LED light-curing unit (Elipar S10, 3 M ESPE, St Paul, MN, USA). A polyethylene tube with an internal diameter of 1 mm and a height of 1.5 mm was mounted on the enamel surface and filled with the composite resin (RelyX Veneer, 3 M ESPE, St Paul, MN, USA). The composite resin was then light cured using the abovementioned light-curing unit for 40 s. The excess material surrounding the tube was removed with a sharp scalpel. The specimens were stored at room temperature for 1 h prior to removal of the polyethylene tubes. All the specimens were then examined with a stereomicroscope to exclude any specimens with air bubbles, evident interfacial gaps, or any other bonding defects.

Thermocycling
Each group was further divided into 2 subgroups (n = 12), according to the thermocycling procedure used.

In subgroup T, the specimens were thermocycled using a thermocycling machine (TC-501F, Weier, Tianjin, China) and then subjected to the μSBS test. Thermocycling was performed for 5000 cycles at bath temperatures of 5 °C and 55 °C, with a dwell time of 30 s in each bath and a transfer time of 10 s.

In subgroup N, the specimens were stored in distilled water at 37 °C for 24 h before undergoing the μSBS test.

μSBS test
The μSBS test was performed using a universal testing machine (AGS-X, Shimadzu, Tokyo, Japan) with a cross-head speed of 1.0 mm/min until failure occurred. The maximum failure load (in N) was recorded and converted into MPa by dividing the failure load by the bonding area (0.785 mm²) [33].

After the test, the fracture surfaces were observed with a stereomicroscope under ×40 magnification. The failure modes were classified into 3 types: 1) adhesive failure: failure between the adhesive and the enamel or between the composite resin and the enamel; 2) cohesive failure: failure within the enamel or within the composite resin; and 3) mixed failure: fractures involving adhesive and cohesive failures simultaneously [20].

Statistical analysis
The data were analyzed using SPSS software (version 20.0 for Windows, SPSS, Chicago, IL, USA) at α = 0.05. The assumption of normality was confirmed using the Kolmogorov-Smirnov test, and the equality of variances was confirmed with the Levene test. Two-way analysis of variances (ANOVA) was used to evaluate the effects of both the bleaching treatment and the thermocycling process and their interactions on the μSBS.

Results
Table 1 presents the means and standard deviations of the μSBS values for the different groups. The group CN (unbleached specimens without thermocycling) showed the highest μSBS value (25.21 ± 5.53 MPa), while the group BT (bleached specimens with thermocycling) exhibited the lowest μSBS value (21.41 ± 7.22 MPa). Two-way ANOVA indicated no significant effects of either the bleaching treatment or thermocycling as well as no significant interaction between the two factors (Table 2).

The frequencies of different failure modes are shown in Fig. 1. Mixed failure was the most prevalent failure mode, followed by adhesive failure and cohesive failure. Compared with group C, group B exhibited higher percentage of adhesive failure.

Discussion
Based on the current results, the null hypotheses that the bleaching treatment would not affect the bond strength between the composite resins and the enamel when the surface is reduced by 0.5 mm immediately after bleaching and that the bond strength between the composite resin and the enamel remains the same after thermocycling were accepted.

The μSBS test was used to measure the bond strength of bleached enamel and to avoid the non-uniform distribution of interfacial stress that occurs during the macro-shear bond tests [21, 33]. Weerasinghe et al. [33] reported a μSBS value for grounded enamel of ~25 MPa, which is consistent with the present finding. The bond strength and failure mode differed between the aged and non-aged samples. The aged samples exhibited lower μSBS values compared with the non-aged samples, although the differences were not statistically significant. Temperature changes during the thermocycling process may amplify the coefficient of thermal expansion mismatch of the bonded materials, which generates mechanical stresses at the bonded interface, resulting in bonding degradation [34].

Veneers and other restorative procedures are often required after the bleaching treatment. Given that very limited information is available, the present study aimed to compare the bond strength of bleached and unbleached enamel after a surface removal of 0.5 mm, which was performed to simulate the tooth preparation process for laminate or composite veneers [33, 35]. After removing 0.5 mm of the superficial

Table 1 Means and standard deviations of the μSBS values (MPa) for different groups

<table>
<thead>
<tr>
<th>Group</th>
<th>Subgroup</th>
<th>μSBS</th>
</tr>
</thead>
<tbody>
<tr>
<td>B (bleaching)</td>
<td>T (with thermocycling)</td>
<td>21.41 (7.22)</td>
</tr>
<tr>
<td></td>
<td>N (without thermocycling)</td>
<td>21.45 (8.87)</td>
</tr>
<tr>
<td>C (control)</td>
<td>T (with thermocycling)</td>
<td>22.10 (6.05)</td>
</tr>
<tr>
<td></td>
<td>N (without thermocycling)</td>
<td>25.21 (5.53)</td>
</tr>
</tbody>
</table>
enamel, the specimens from the bleaching group exhibited similar μSBS values to those from the control group, regardless of thermocycling. Although the bond strength of bleached enamel without surface reduction was not tested in the present study, a reduction in the enamel bond strength after bleaching has been well documented by previous studies [14–16]. Evidence has also shown a reduction in resin tags as well as a lower degree of adhesive penetration into the bleached enamel substrate [13, 36]. The abovementioned phenomena have been attributed to the residual free radicals being released from the bleaching agents and to bleaching-induced structural changes in the enamel [14, 15, 25]. Although no information is available regarding the amount of residual free radicals or structural changes found at different depths of dental hard tissues, it is conceivable that changes due to bleaching occur primarily on the enamel surface. The residual oxygen and structural changes caused by bleaching would therefore be eliminated, at least to some extent, by the enamel reduction process. This hypothesis is supported by previous studies that found that the topical application of antioxidant agents [18, 26, 27] and the use of laser irrigation [28, 29] could reverse the reduced bond strength of enamel after bleaching. Interestingly, although no significant differences in the bond strength were found, the mean bond strength of bleached enamel was lower than that for unbleached enamels, and the frequency of adhesive failure in the bleaching group was higher than that in the control group. These findings may indicate that the negative effects of bleaching were not completely reversed, even after the superficial layer of the enamel was removed. However, this hypothesis should be tested in future studies.

Due to morphological and physicochemical similarities, bovine teeth are a good alternative to human teeth for experimental procedures [37, 38]. Moreover, human and bovine teeth have been reported to exhibit similar behaviors during bleaching treatments [39]. Thus, the present study used bovine teeth as the testing substrates, in accordance with the previous studies [20, 28, 29, 40].

### Table 2: Two-way ANOVA results for the μSBS values

<table>
<thead>
<tr>
<th>Source of variance</th>
<th>Type III sum of squares</th>
<th>df</th>
<th>Mean square</th>
<th>F</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bleaching</td>
<td>49.146</td>
<td>1</td>
<td>49.146</td>
<td>0.906</td>
<td>0.348</td>
</tr>
<tr>
<td>Thermocycling</td>
<td>24.371</td>
<td>1</td>
<td>24.371</td>
<td>0.449</td>
<td>0.507</td>
</tr>
<tr>
<td>Bleaching x thermocycling</td>
<td>23.621</td>
<td>1</td>
<td>23.621</td>
<td>0.435</td>
<td>0.514</td>
</tr>
<tr>
<td>Error</td>
<td>1953.251</td>
<td>36</td>
<td>54.257</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

![Fig. 1](image-url) Distribution of failure modes among different groups.
Based on the μSBS results, clinicians should be able to perform the bonding procedures using bleached enamel immediately after bleaching if 0.5 mm of the enamel is first removed from the tooth surface. The negative effects of bleaching on the bond strength of enamel have been suggested to be hydrogen-peroxide-concentration-dependent [20]. Because a 40% hydrogen peroxide gel was employed to simulate the worst-case scenario, the present findings may also be applicable to at-home bleaching, which uses a lower concentration of hydrogen peroxide or carbamide peroxide. However, further in vitro and in vivo studies are necessary to confirm this hypothesis.

Conclusions
Within the limitations of the present study, it can be concluded that bleached and unbleached enamel exhibited similar μSBS values after removing 0.5 mm of the enamel from the surface.

Abbreviations
ANOVA: analysis of variances, μSBS: micro-shear bond strength

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Availability of data and materials
Further data may be requested by contacting the corresponding author. We declare, that any data regarding the study will easily be provided.

Authors’ contributions
CYL drafted the paper and collected data. CYL and MJ performed the experiment. ZM performed the statistical analysis. AT and CH helped with the experimental design and revised the paper. YH designed the study and revised the paper. All authors have read and approved the manuscript in its current form.

Ethics approval and consent to participate
This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31). This study was approved by the Research Ethics Committee of School and Hospital of Stomatology, Fujian Medical University (No. 2015-CX-31).

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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