The Histogenesis of Kaposi's Sarcoma

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The histogenesis of Kaposi's sarcoma has been controversial since its first description by Moritz Kaposi in 1872 (1). Although previously rare, the recent increase in its incidence associated with the acquired immune deficiency syndrome (AIDS) (2) has stimulated new interest in its origin. Early studies of the histogenesis have been the subject of several reviews (3,4). A remarkable list of cells of origin was proposed in these early studies. However, more recent studies have focused on an endothelial or pluripotent mesenchymal cell origin. These studies, utilizing electron microscopy, enzyme histochemistry and immunohistochemistry have also failed to unequivocally establish the cell of origin in Kaposi's sarcoma (5-27).

We have rigorously tested the prevailing hypothesis that the lesion is derived from vascular endothelial cells. We utilized a combination of enzyme histochemistry and immunohistochemistry in plastic and frozen sections. We used seven markers to characterize human endothelial cells: three antigens (Factor VIII-related antigen, HLA-DR/Ia, macrophage/endothelial antigens), three enzymes (5'-nucleotidase, ATPase, alkaline phosphatase), and lectin binding (Ulex europaeus I). We found distinctive phenotypes for normal vascular and lymphatic endothelial cells and that the abnormal cells of Kaposi's sarcoma most closely resembled the phenotype seen in lymphatic endothelium.

Materials & Methods

Biopsy specimens from 40 patients were studied with plastic embedding techniques. All of these specimens were examined histochemically for 5'-nucleotidase, ATPase, and alkaline phosphatase, immunohistochemically for Factor VIII-related antigen (FVIII RA), and HLA-DR/Ia antigen, using immunoperoxidase, and also with the Ulex europaeus I (UEA) lectin directly conjugated to peroxidase. The details of these procedures have been previously published (28-31). Biopsies from an additional 9 patients were snap-frozen, cryostat-sectioned, and stained immunohistochemically for FVIII RA, HLA-DR/Ia, and 3 macrophage/endothelial (MΦ/E) antigens (clones 63D3 and 61D3 from D. Capra, University of Texas at Dallas, and Leu M3), using previously described procedures (30,32).
**Results**

These studies revealed considerable phenotypic variation in the endothelial cells of different vessel types. This is summarized in Table 1.

<table>
<thead>
<tr>
<th>Table 1—Characterization of Normal Human Endothelium</th>
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<td>Staining With Endothelial Markers</td>
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<td>Lymphatic</td>
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<td>Arteriole</td>
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<td>Capillary</td>
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<td>Venule</td>
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FVIII RA, Factor VIII-related antigen; HLA-DR/la, HLA-DR/la-like antigen; UEA, Ulex europaeus I lectin; MΦ/E, macrophage/endothelial antigens; Alk<sup>+</sup>, alkaline phosphatase; 5'-N, 5'-nucleotidase; ATPase, adenosine triphosphatase; 0, negative; 0<sup>-</sup>, a rare weakly positive cell; +, weakly positive; ++, positive; ++++, strongly positive.

Only minor differences were apparent in the phenotypes of the different blood vessel endothelial cells, but there were significant deviations from this phenotype in the lymphatic endothelial cells. Lymphatic endothelium lacked the enzymes ATPase and Alk<sup>+</sup> but had the strongest 5'-N reaction of all vessels. There was very little staining with HLA/DR, FVIII RA, or MΦ/E antigens, but a strong reaction with UEA was seen.

The lesions of Kaposi's sarcoma are frequently a complex mixture of normal vessels and a spectrum of abnormal elements ranging from irregular vascular channels to spindle cells. The abnormal endothelial cells showed consistent staining with UEA and 5'-N, although the staining was weaker and more patchy in the spindle cells. The other endothelial markers (FVIII RA, HLA-DR/la, and MΦ/E, Alk<sup>+</sup>, ATP) generally failed to stain the abnormal endothelial cells or spindle cells. All the normal vessels in the lesion showed a staining pattern consistent with normal blood vessel endothelium and served as internal controls. These results are summarized in Table 2.

<table>
<thead>
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<th>Table 2—Characterization of Kaposi's Sarcoma</th>
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<tr>
<td>Staining with endothelial markers*</td>
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<td>Cell type</td>
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<td>Plastic sections</td>
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<td>Abnormal endothelial cells of Kaposi's sarcoma</td>
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<td>Spindle cells of Kaposi's sarcoma</td>
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<td>Frozen sections</td>
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<td>Spindle cells of Kaposi's sarcoma</td>
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FVIII RA, Factor VIII-related antigen; HLA-DR/la, HLA-DR/la-like antigen; UEA, Ulex europaeus I lectin; Alk<sup>+</sup>, alkaline phosphatase; ATPase, Adenosine triphosphatase; 5'-N, 5'-nucleotidase; ND, not done.

* Data are given as number of cases positive per number of cases tested with adequate survival of marker. Only solid clusters of abnormal spindle cells are tabulated so that confusion with reactive stromal elements is avoided.
Discussion

We have demonstrated that the phenotype of the abnormal proliferating cell in Kaposi's sarcoma differs significantly from that of non-neoplastic blood vessel endothelium. The phenotype [FVIII RA(-), HLA-DR/IA(-), Mφ/E(-), UEA((+), ATP(-), Alkφ(-), 5’-N(+)] more closely resembles that of normal lymphatic endothelium. This data is supported by the electron-microscopic studies of McNutt et al. (18), who found a striking similarity between early Kaposi's lesions and dermal lymphatics. A lymphatic endothelial origin has also been suggested by several early studies based on clinicopathologic data (3,4,7).

We conclude that the irregular vascular spaces in the earliest recognizable lesions of Kaposi's sarcoma are formed by cells closely resembling lymphatic endothelial cells. These lesions are complex with an intermixture of inflammatory cells and neovascularization by phenotypically normal blood vessels. With continued proliferation some of the abnormal endothelial cells lose the ability to form channels which is accompanied by a considerable loss of UEA binding and 5'-N. Morphologic evidence of clear transitions between these two states suggests that the spindle cell element is derived from the abnormal endothelial cells. These studies strongly suggest that the lymphatic endothelial cell is the cell of origin in Kaposi's sarcoma.

References

13. Sterry, W., Steigleder, G., and Bodeux, E.: Kaposi's sarcoma: Venous


